

Supporting information

Methods

Infection with *Leishmania major*

Mice were injected in the left hind footpad with 1×10^6 promastigotes of *Leishmania major* MHOM/S U/73-5-ASKH strain (*L. major*). The thickness of the infected and contralateral uninfected footpad was measured as described (1).

To measure the immune responses to pathogens, draining popliteal lymph node cells ($3-10 \times 10^4$) were cultured in the presence of *L. major* antigen for 48 h as described (1).

(1) Tominaga N, Ohkusu-Tsukada K, Udono H, Abe R, Matsuyama T, Yui K (2003)

Development of Th1 and not Th2 immune responses in mice lacking IFN-regulatory factor-4. *Int Immunol* 15: 1-10.

Figure 6

IRF-4^{-/-} mice are resistant to *L. major* infection

Mice were injected in the left hind footpad with 1×10^6 promastigotes of *Leishmania major* MHOM/S U/73-5-ASKH strain (*L. major*). The thickness of the infected and contralateral uninfected footpad was measured as described (1). (A) BALB/c (open circle, n=8), B6 (open triangle, n=4) and IRF-4^{-/-} mice (closed circle, n=14) were infected with *L. major* promastigotes, and the increase in footpad thickness (%) was determined. The data represent mean \pm SD of each group. (B) Four weeks after infection, 1×10^5 (gray bar) and 3×10^4 (open bar) draining popliteal lymph node cells were cultured for 48 h in the presence of *L. major* antigen. The levels of IFN- γ in the supernatant were determined by ELISA. Representative results of three independent experiments are shown. *: not detectable.

Figure 7

CD4⁺ NK T cells from IRF-4^{-/-} mice produce IL-4 in response to alpha-GalCer

(1) Splenic CD4⁺ T cells were stained with FITC-anti-CD4 and with biotin-anti-DX5 mAb plus PE-streptavidin, and analyzed using FACS Aria. The proportion of cells within the square is indicated.

(2) Sorted CD4⁺ DX5⁺ cells (5×10^4) from BALB/c (open bar) and IRF-4^{-/-} (dark bar) mice were cultured with splenic dendritic cells (CD11c⁺ cells, 1×10^4) in the presence (+) and absence (-) alpha-GalCer (200ng/ml) for 48h. The cytokine levels in the supernatant were determined by ELISA. The purity of CD4⁺DX5⁺ cells from BALB/c and IRF-4^{-/-} mice was 93.3 and 70.6%, respectively. The lower purity of CD4⁺DX5⁺ cells from IRF-4^{-/-} mice was due to the lower expression of DX5 marker and their small numbers.

Figure 8

RNA expression of Th2 cytokines by naïve CD4⁺ T-cells.

Naïve (CD62L⁺) CD4⁺ T-cells from BALB/c (open circle) or IRF-4^{-/-} (closed circle)

mice were cultured with plates coated with anti-TCR mAb for 0-24 h. Messenger

RNA levels were determined by real time PCR and were expressed as the ratio of DNA

to G3PDH. Representative results of three independent experiments are shown.

Figure 9

The forced expression of IRF-4 in IRF-4^{-/-}CD4⁺ T-cells enhanced their Th2 cytokine production.

CD4⁺ T cells from IRF-4^{-/-} mice were co-transfected with pcDNA3 or pcDNA3-mIRF-4 together with pmaxEGFP at the ratio of 5:1, cultured for 3h, and stimulated with plate-bound anti-TCR mAb (0-10microgram/ml) for 48 h. The proportion of cells expressing GFP reached ~40% 24 h after the gene transfer. Cell lysate was prepared from unstimulated cells, separated by 12.5% SDS-PAGE, blotted, and probed with anti-IRF-4 Ab. The blot was stripped, and re-probed with anti-actin Ab. The levels of IL-4 and IL-5 in the supernatant were determined by ELISA. The proportion of EGFP⁺ cells was 30-45%. Mann-Whitney's *U* test for unpaired observations was used to calculate *p* values. Representative results of three independent experiments are shown.

Figure 6.

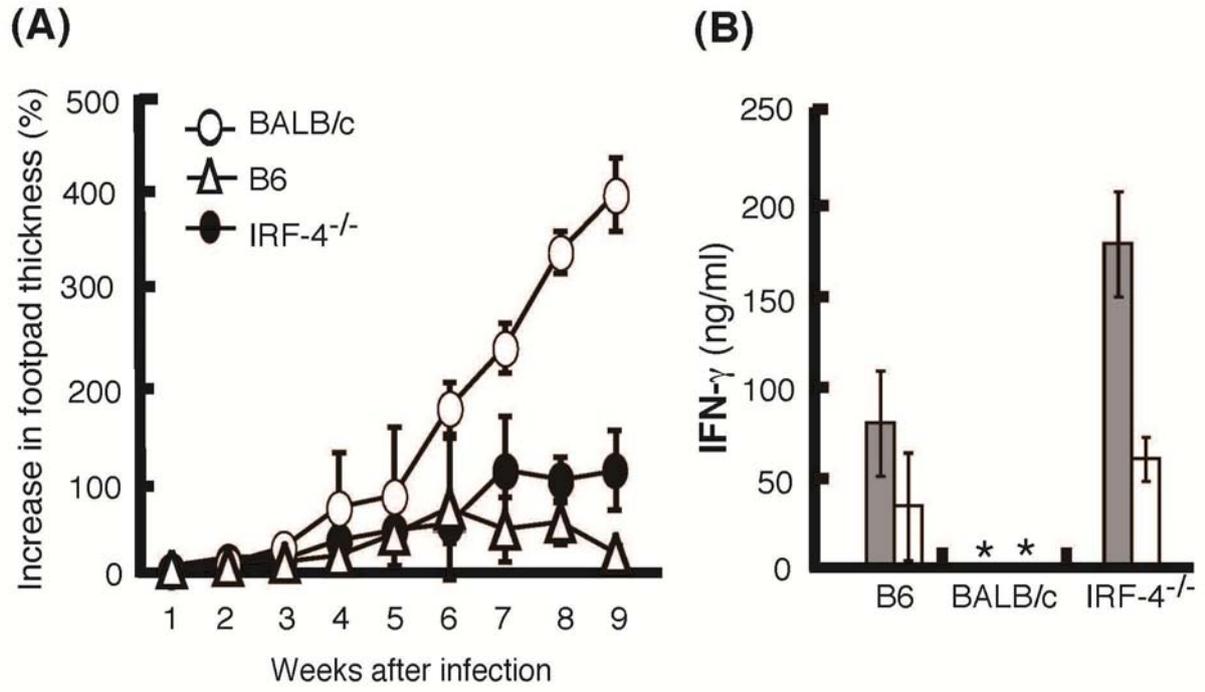


Figure 7.

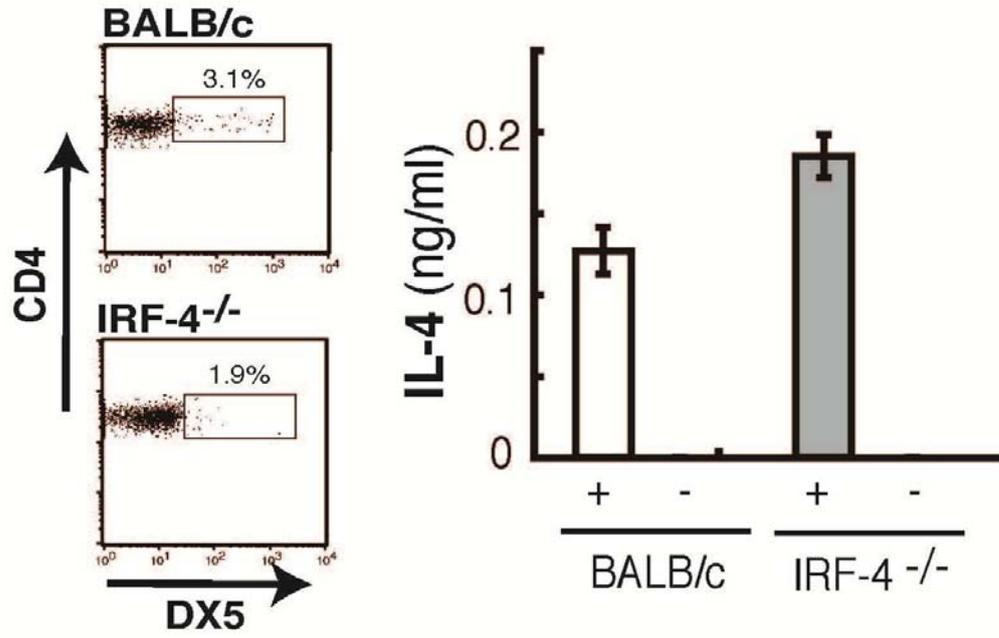


Figure 8.

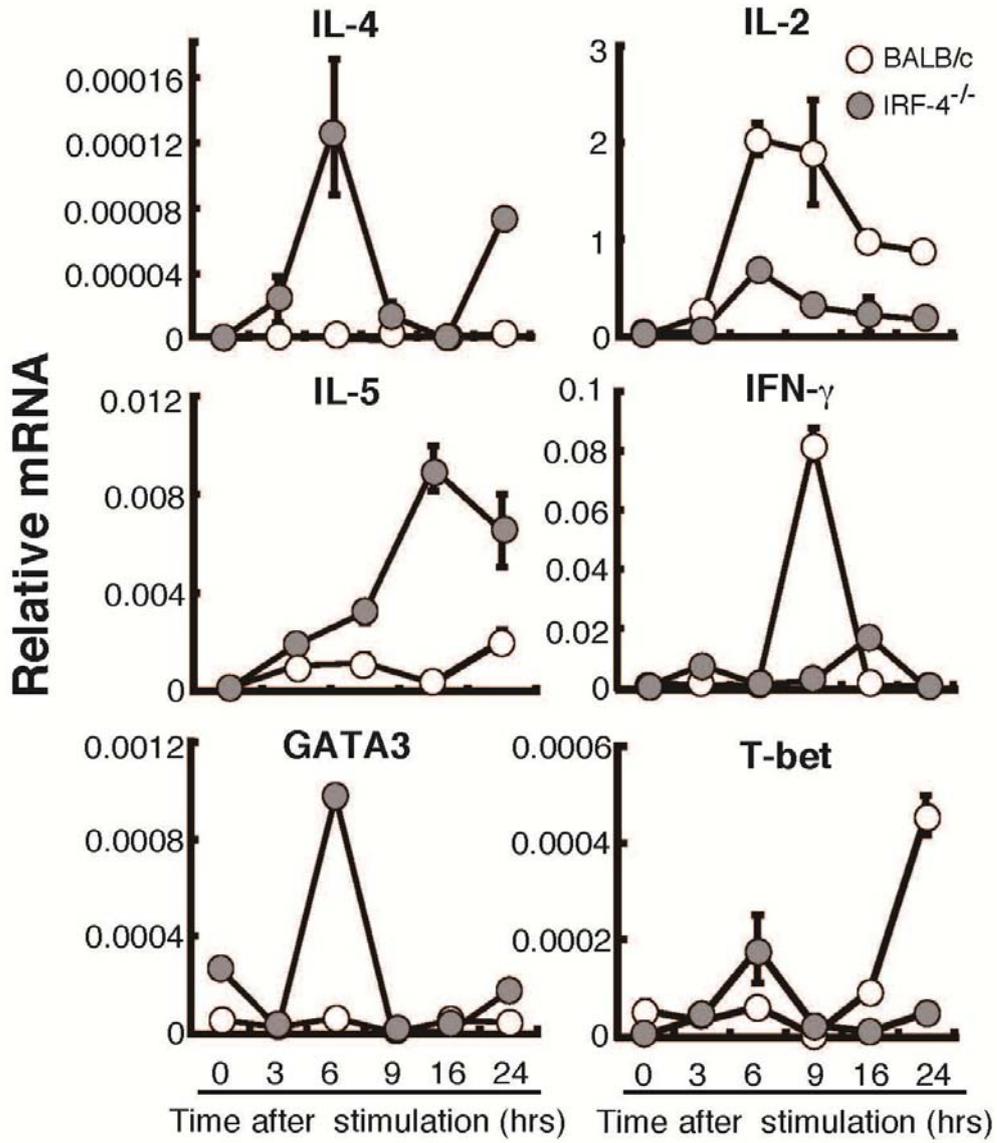


Figure 9.

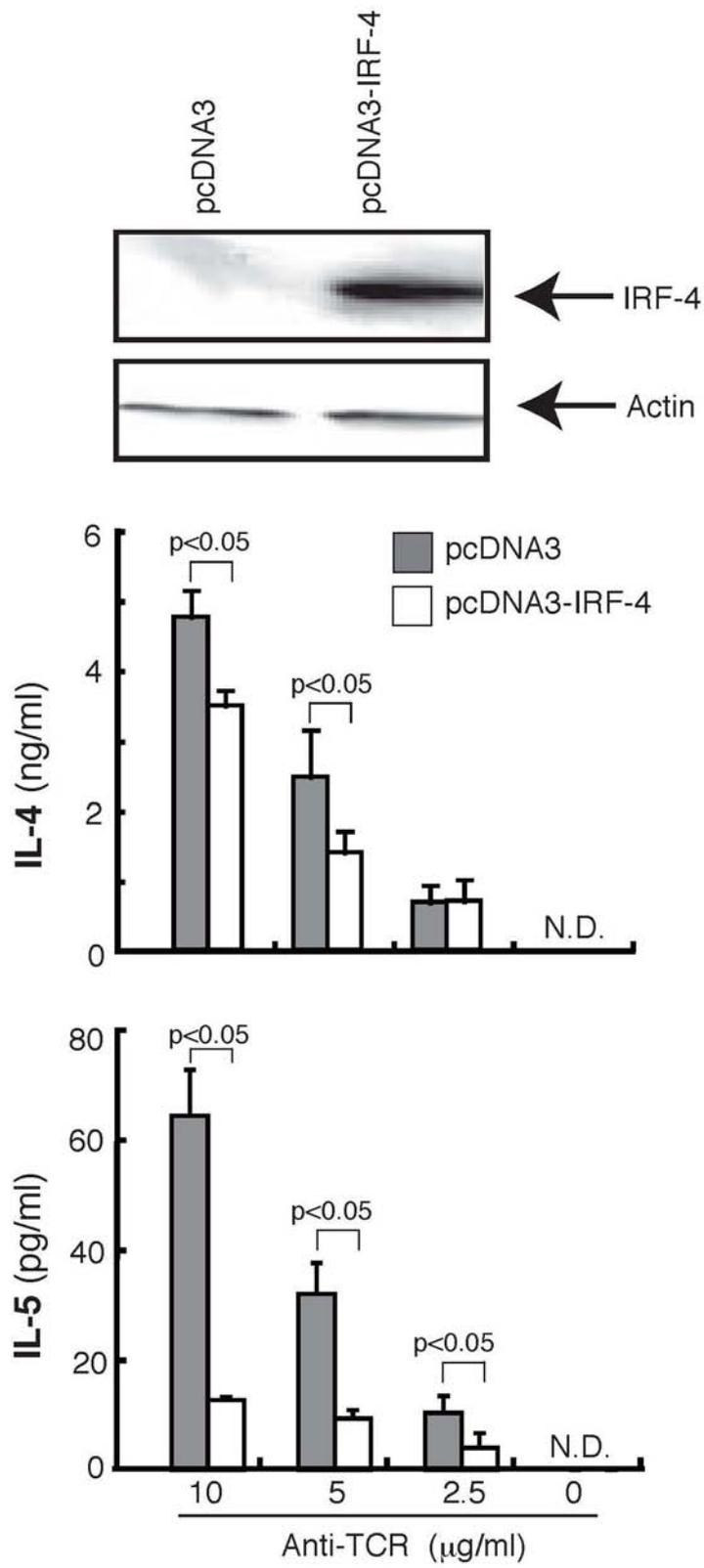


Table S1 Primer sequences used for real-time RT-PCR

name	5' primer	3' primer
IL-4	TCATCGGCATTTTGAACGAG	TTTGGCACATCCATCTCCG
IL-5	CTCTCAGCTGTGTCTGGGCC	GCTTGTCAACAGAGCTCGGTG
IFN-gamma	GCATTCATGAGTATTGCCAAGTTT	GATTCCGGCAACAGCTGGT
IRF-1	ATTCCAACCAAATCCCAGGG	CTCCGGAACAGACAGGCATC