

[CASE REPORT]

Late-onset Cerebrotendinous Xanthomatosis with a Novel Mutation in the CYP27A1 Gene

Akari Sasamura¹, Satoru Akazawa¹, Ai Haraguchi¹, Ichiro Horie¹, Takao Ando¹, Norio Abiru¹, Hajime Takei², Hiroshi Nittono², Mizuho Une³, Takao Kurosawa⁴, Tsuyoshi Murai⁴, Hiromu Naruse^{5,6}, Tomohiro Nakayama^{5,7}, Kazuhiko Kotani⁸, Alan T. Remaley⁹ and Atsushi Kawakami¹

Abstract:

Cerebrotendinous xanthomatosis (CTX) is a rare, autosomal recessive, inborn disruption in bile acid synthesis characterized by severe systemic xanthomas, cataracts and neurological injuries occurring before adolescence without elevation of the serum cholesterol or triglyceride levels. CTX is caused by a deficiency of the mitochondrial enzyme sterol 27-hydroxylase, which is encoded by the *CYP27A1* gene. We herein report a 50-year-old Japanese woman with late-onset CTX who had no relevant symptoms before the development of bilateral Achilles tendon xanthomas in middle age. A genetic analysis revealed a compound heterozygous mutation in the *CYP27A1* gene with a previously known missense mutation (NM_000784.3:c.1421 G>A) and a novel frame shift mutation of NM_000784.3:c.1342_1343insCACC.

Key words: cerebrotendinous xanthomatosis (CTX), sterol 27-hydroxylase (*CYP27A1*), cholestanol, norcholic acid, chenodeoxycholic acid (CDCA), cholesterol efflux

(Intern Med 57: 1611-1616, 2018) (DOI: 10.2169/internalmedicine.0120-17)

Introduction

Cerebrotendinous xanthomatosis (CTX) is an autosomal recessive lipid storage disease caused by disruption in bile acid synthesis induced by mutations in the *CYP27A1* gene on chromosome 2q33-qter (1). Fifty-four different mutations in *CYP27A1* have been reported thus far (2). The first case of CTX was described in 1937 by van Bogaert (3). Since then, more than 300 cases have been reported in countries around the world. There is no sex predominance. CTX is an extremely rare disease; its prevalence was estimated as 1.9 per 100,000 people among Caucasians of European ancestry (2). The estimated incidence of CTX is higher in East

Asia, at approximately 15.5 per 100,000 people (4). However, precise epidemiologic data from Japan are lacking.

The *CYP27A1* gene encodes the mitochondrial enzyme sterol 27-hydroxylase, which hydroxylates the steroid side chain at C-27 (5). Deficiency of this enzyme leads to the accumulation of cholestanol and bile alcohols in connective tissues and neuronal cells and results in systemic symptoms, such as neurological disorders, atherosclerosis, tendon xanthomas and cataracts. Patients with CTX typically develop cataracts before their 10th birthday and develop systemic xanthomas in the second or third decade of life. Although most CTX patients have normal intelligence until puberty, intellectual functioning worsens with age, and over 50% of CTX patients show an intellectual decline in their 20s.

¹Department of Endocrinology and Metabolism, Nagasaki University Hospital, Japan, ²Junshin Clinic Bile Acid Institute, Japan, ³Faculty of Pharmaceutical Science, Hiroshima International University, Japan, ⁴School of Pharmaceutical Sciences, Health Sciences University of Hokkaido, Japan, ⁵Division of Laboratory Medicine, Department of Pathology and Microbiology, Nihon University School of Medicine, Japan, ⁶Health Science Research Institute, Inc., Japan, ⁷Division of Companion Diagnostics, Department of Pathology of Microbiology, Nihon University School of Medicine, Japan, ⁸Division of Community and Family Medicine, Jichi Medical University, Japan and ⁹Lipoprotein Metabolism Section, National Institutes of Health, USA

Received: August 25, 2017; Accepted: November 6, 2017; Advance Publication by J-STAGE: February 9, 2018 Correspondence to Dr. Ichiro Horie, holy197741@me.com



Figure 1. A: Xanthoma of the Achilles tendon on T2-weighed MRI. B: Extremely thickened Achilles tendon. C: FLAIR MRI of the brain. Arrows indicate bilateral high-intensity areas in the dorsal lesion behind the cerebellar dentate nucleus. D: Hematoxylin and Eosin staining of xanthoma in the Achilles tendon. Foamy cells admixed with inflammatory giant cells infiltrated the tendinous tissue, and cholesterol clefts were visible.

Slowly progressive neurologic symptoms, such as pyramidal tract signs, cerebellar ataxia and peripheral neuropathy, are typically observed in their 20s and 30s (2). There is significant clinical heterogeneity, even among siblings with the same mutation in the *CYP27A1* gene; as such, the clinical phenotypes do not seem to be strictly correlated with the genotypes in CTX (2).

Chenodeoxycholic acid (CDCA) has been demonstrated to decrease the levels of serum cholestanol and urinary bile alcohols in CTX patients (6). Treatment with CDCA can stabilize or even reverse some of the symptoms in most patients with CTX (7). Starting CDCA during childhood or earlier is believed to be more beneficial than starting treatment in adulthood (8).

We herein report a Japanese woman diagnosed with CTX at age 50 who had no related symptoms of CTX before the development of bilateral Achilles tendon xanthomas in middle age.

Case Report

Case presentation

The patient was a 50-year-old Japanese woman from nonconsanguineous parents. She has no siblings but had a child. There was no apparent family history (parents, child and parent's siblings) of neurological disorders, cardiovascular diseases or any lipid storage diseases inducing xanthomas, including CTX. She had no history of neonatal jaundice or chronic infantile diarrhea. She had been diagnosed with idiopathic epilepsy and medicated from 7 to 10 years of age. Thereafter, she had no remarkable medical developments until middle age. She insidiously developed painless tendon thickness of the bilateral Achilles tendons and right triceps muscle at 47 years of age. The gradual enlargement of the Achilles tendons caused gait disturbance when she was 49 years of age. T1- and T2-weighted magnetic resonance imaging (MRI) revealed low-intensity lesions in the thickened bilateral Achilles tendons (Fig. 1A) and right triceps muscle,

	Case	Reference		Case	Reference
Total cholesterol (mg/dL)	203	142-248	Total bilirubin (mg/dL)	1.9	0.4-1.5
Triglyceride (mg/dL)	123	30-117	Direct bilirubin (mg/dL)	0.2	≤ 0.4
HDL-C (mg/dL)	75	48-103	AST (IU/L)	16	13-30
LDL-C (mg/dL)	103	65-163	ALT (IU/L)	9	7-23
Apo A-I (mg/dL)	167	119-155	γ -GTP (IU/L)	8	9-32
Apo A-II (mg/dL)	28.1	25.9-35.7	BUN (mg/dL)	9.0	8-20
Apo B (mg/dL)	80	73-109	Creatinine (mg/dL)	0.52	0.46-0.79
Apo C-II (mg/dL)	7.0	1.8-4.6	Calcium (mg/dL)	9.1	8.8-10.1
Apo C-III (mg/dL)	11.2	5.8-10.0	Phosphate (mg/dL)	3.0	2.7-4.6
Apo E (mg/dL)	6.3	2.7-4.3	Free T4 (ng/dL)	1.27	0.95-1.57
Free cholesterol (mg/dL)	56	34-66	TSH (µIU/mL)	0.43	0.48-5.08
Free fatty acid (mEq/L)	0.59	0.10-0.90	Intact PTH (pg/mL)	69.0	10.3-65.9
Phospholipid (mg/dL)	243	150-280	25(OH)VitD (ng/mL)	14	30-60
Lipoprotein(a) (mg/dL)	11.4	≤ 40	1,25(OH)2VitD (pg/mL)	48	20-60
MDA-LDL (U/L)	119	46-105	TRACP-5b (mU/dL)	385	120-420
SAA-LDL (µg/dL)	5.8	<20	Intact P1NP (µg/L)	66.2	17.1-64.7
Glucose (mg/dL)	80	73-109	LH (IU/L)	37.5	1.4-15
Insulin (mIU/L)	2.3	0.87-10.7	FSH (IU/L)	39.1	3.0-24.0
HbA1c (%)	5.3	4.9-6.0	Estradiol (pg/mL)	360.1	28.8-491.9

Table 1.	Serum	Lipids and	Associated	Laboratory	Findings on	Admission.
----------	-------	------------	------------	------------	-------------	------------

HDL-C: high-density lipoprotein cholesterol, LDL-C: low-density lipoprotein cholesterol, Apo: apoprotein, MDA-LDL: malondialdehyde-modified LDL, SAA-LDL: serum amyloid A-LDL, AST: aspartate aminotransferase, ALT: alanine aminotransferase, γ -GTP: gamma-glutamyltransferase, BUN: blood urea nitrogen, TSH: thyrotropin, PTH: parathyroid hormone, TRACP-5b: tartrate resistant acid phosoatase-5b, P1NP: procollagen type 1 N-terminal propeptide, LH: luteinzing hormone, FSH: fertilizing hormone

suggestive of xanthomas. She was admitted to Nagasaki University Hospital for further investigation at 50 years of age.

Clinical, laboratory and imaging examinations

The patient was lean (height 149.0 cm, weight 39.2 kg, BMI 17.7 kg/m²), and the findings of a physical examination were unremarkable except for confirmation of xanthomas on the bilateral Achilles tendons (Fig. 1B) and right triceps muscle. She had age-appropriate slight cataracts in the bilateral eyes and did not have dementia based on the Revised Hasegawa Dementia Scale (HSD-R; 26/30), Mini-Mental State Examination (MMSE; 27/30), and Frontal Assessment Battery (FAB; 16/18) scores. On a neurological examination, her gait appeared somewhat spastic, and all of her deep tendon reflexes were mildly enhanced bilaterally with positive Babinski and Chaddock reflexes, indicating impairment in the bilateral pyramidal tracts. There were no neurological findings suggesting cerebellar impairment.

Regarding the laboratory findings (Table 1), there was no elevation in the levels of serum cholesterol or triglycerides or deficiencies in apolipoproteins or thyroid hormones. As the clinical findings suggested CTX, we studied the bile acids and sterols in her serum and urine (Table 2). The primary bile acids were extremely low in the serum and urine when measured by both high-performance liquid chromatography (HPLC) and gas chromatography/mass spectrometer (GC/MS). The urinary levels of bile alcohols, especially 5βcholestane- 3α , 7α , 12α ,24,25-pentol, were markedly elevated. Norcholic acid, which is mainly derived from 5 β -cholestane-3 α ,7 α ,12 α ,23,25-pentol, was highly elevated and accounted for 78.4% of all bile acids in the urine. The serum level of cholestanol was extremely elevated, while those of sitosterol and campesterol were not elevated.

T2-weighted and fluid-attenuated inversion recovery (FLAIR) MRI of the brain showed bilateral low-intensity areas in the dentate nuclei and high-intensity areas in the peripheral white matter the cerebellum (Fig. 1C). We also found bilateral high-intensity areas along the pyramidal tracts from the pons to the cerebral crus. Brain atrophy was not remarkable. Based on these clinical, laboratory and radiological findings, the patient was diagnosed with CTX.

Ultrasound of the carotid arteries showed a slightly thickened intima-media complex at the left common carotid artery (maximum intima-media thickness, 1.2 mm). However, there was no evidence of cardiovascular disease on electrocardiogram, echocardiography and MR angiography of the brain and lower extremities.

Despite being premenopausal (Table 1), she had osteoporosis as determined by dual-energy X-ray absorptiometry, which showed a bone density of 61% and 75% the young-adult-mean in the lumbar vertebra and femoral neck, respectively.

Genetic diagnosis

Genomic DNA was isolated from peripheral blood leukocytes. All 9 exons of the *CYP27A1* gene were amplified by polymerase chain reaction (PCR) and sequenced. Two muta-

	Before	After		Before	After	
Serum bile acid (HPLC; µmol/L)			Serum bile acid (GC/MS; µmol/L)			
Glycoursodeoxycholic acid	N.D.	2.1	Total bile acid	0.10	6.82	
Tauroursodeoxycholic acid	N.D.	N.D.	Cholic acid	N.D.	N.D.	
Ursodeoxycholic acid	N.D.	1.0	Chenodeoxycholic acid	N.D.	3.80	
Glycocholic acid	N.D.	N.D.	Ursodeoxycholic acid	N.D.	2.10	
Taurocholic acid	N.D.	N.D.	Deoxycholic acid	N.D.	N.D.	
Cholic acid	N.D.	N.D.	Lithocholic acid	N.D.	N.D.	
Glycochenodeoxycholic acid	N.D.	3.4	Norcholic acid	0.10	N.D.	
Taurochenodeoxycholic acid	N.D.	N.D.	Others	N.D.	0.92	
Chenodeoxycholic acid	N.D.	1.2				
Glycodeoxycholic acid	N.D.	N.D.	Urinary bile acid (GC/MS; mm	Urinary bile acid (GC/MS; mmol/mol·Cr)		
Taurodeoxycholic acid	N.D.	N.D.	Total bile acid	0.50	1.09	
Deoxycholic acid	N.D.	N.D.	Cholic acid	0.03	N.D.	
Glycolithocholic acid	N.D.	N.D.	Chenodeoxycholic acid	0.01	0.12	
Taurolithocholic acid	N.D.	0.3	Ursodeoxycholic acid	N.D.	0.64	
Lithocholic acid	N.D.	N.D.	Deoxycholic acid	N.D.	N.D.	
			Lithocholic acid	N.D.	0.06	
			Norcholic acid	0.39	0.04	
			Others	0.07	0.23	
Serum sterol (GC; µg/mL)			Urinary bile alcohol (GC/MS;	cohol (GC/MS; mmol/mol·Cr)		
Cholestanol	25.2	7.8	Total bile alcohol	3.44	1.62	
Sitosterol	4.0	3.0	5β -cholestane- 3α , 7α , 12α ,	2.02	1.11	
Campesterol	10.1	5.1	24,25-pentol			
			Others	1.42	0.51	

Table	2.	Serum	Levels of Bile	Acids and	Associated	Biosynthetic	Products b	efore and	6 Months
after A	٨dm	inistrati	on of CDCA.						

CDCA: chenodeoxycholic acid, HPLC: high performance liquid chromatography, GC: gas chromatography, GC/MS: gas chromatography mass spectrometry, N.D.: not detectable, Cr: creatinine



Figure 2. A direct nucleotide sequence analysis of the exon 8 of *CYP27A1* gene. A: The patient was heterozygous for the missense mutation of NM_000784.3: c.1421 G>A, resulting in the amino acid substitution of arginine to glutamine at codon 474 (NP_000775.1: pArg474Gln), as previously reported (Ref. 9). B: The patient was also heterozygous for a frame-shift mutation of NM_000784.3: c.1342_1343insCACC (NP_000775.1: p.Arg448ProfsTer521), which has never been reported before.

tions were identified. One was a known missense mutation of NM_000784.3:c.1421GA in exon 8 (Fig. 2A), resulting in an amino acid substitution of arginine to glutamine at codon 474 (R474Q), as reported previously (9, 10). The other was a novel frame-shift mutation of NM_000784.3:c.1342_ 1343insCACC in exon 8 (Fig. 2B). As we did not perform genetic analyses of these two mutations in the patient's parents, we were unable to determine whether the patient was a compound heterozygote of two mutated alleles or had double mutations in one allele. Therefore, we assumed the patient to be a compound heterozygote of these two mutations based on the clinical and laboratory findings.

Cholesterol efflux assay

A cholesterol efflux assay was carried out as described previously (11). In brief, ATP-binding cassette transporter A 1 (ABCA1)-transfected cell were incubated with [³H]cholesterol. After incubation, the radioactive counts in media and cell lysates were measured by liquid scintillation counting. The ABCA1-mediated cholesterol efflux capacity, expressed as the percentage of labeled cholesterol moved from the cells to the medium, was 11.0% in the patient (reference

level: > 7.0%).

Treatments

After CTX was diagnosed, the patient was treated with CDCA at 500 mg per day, divided into 2 doses. Alendronate at 35 mg per week and alfacalcidol at 0.5 μ g per day were prescribed to treat her osteoporosis. These treatments are ongoing. She underwent bilateral Achilles-tendon plastic surgery, and the gait disturbance was improved dramatically after surgery. A pathological examination of the excised tissue revealed foamy cells admixed with inflammatory giant cells and cholesterol clefts surrounding tendinous tissues (Fig. 1D).

The serum levels of several bile acids were increased to detectable ranges, and the serum cholestanol and urinary norcholic acid levels were dramatically decreased six months after the administration of CDCA was started (Table 2). The serum cholestanol levels remained substantially low for at least 18 months after the initiation of CDCA treatment (4.5 and 5.4 μ g/mL at 12 and 18 months, respectively). The patient has shown no regrowth of the tendon xanthomas or any other complications in the post-treatment period.

Discussion

We here present a case of late-onset CTX with compound heterozygous mutations with a known missense and a novel frame shift mutation in the *CYP27A1* gene.

The pathogenesis of neuronal injury and other involvements in CTX has not been fully clarified. The leading hypothesis is that accumulation of cholestanol and cholesterol in the plasma membrane induces apoptosis of the neurons, and accumulation in other tissues causes xanthomas, juvenile-onset atherosclerosis and cataracts (12, 13). However, our patient was almost lacking in neurological and atherosclerotic findings other than the slight pyramidal-tract impairment and senile cataract. It is impossible to exclude that she had spinal cord involvements because we did not evaluate the spinal cord by MRI before starting CDCA treatment. The idiopathic epilepsy medicated in her childhood might be a symptom of CTX. It is unclear why she mainly developed xanthomas without other symptoms seen in CTX until middle age.

It was reported that sterol 27-hydroxylase expressed in macrophages enhances the cholesterol efflux capacity (14, 15). 27-hydroxycholesterol is a ligand of liver X receptor α (LXR α) (16) and plays a role in up-regulating the cholesterol efflux via the ABCA1 pathway (17). In this context, it would be of interest to know whether or not the cholesterol efflux is impaired for atherosclerotic formation in patients with sterol 27-hydroxylase deficiency, i.e. CTX. Although we performed an ABCA1-mediated cholesterol efflux assay in the present CTX patient, the efflux capacity was found to be relatively normal, which may partially account for the absence of apparent atherosclerotic disease in our case. More data are needed to clarify the relationship between sterol 27-hydroxylase and the cholesterol efflux in patients with CTX.

It has been reported that patients with CTX tend to develop osteoporosis (18). Our patient developed osteoporosis before menopause, and we found that her serum level of 25 (OH)VitD was markedly decreased. Sterol 27-hydroxylase is one of several enzymes with vitamin D 25-hydroxylase activity (19), which may be a molecular mechanism underlying the development of osteoporosis in CTX. A previous report in a small group of CTX patients suggested an association of low serum 25(OH)VitD levels with osteoporosis (20), although other studies have reported relatively normal levels of 25(OH)VitD (18, 21). Therefore, the role of 25(OH)VitD in osteoporosis in CTX remains controversial (22).

Although a fundamental treatment for CTX has not yet been established, CDCA was shown to stabilize and even reverse some of the associated symptoms of CTX (2, 8). This is because the supplementation of CDCA, a primary bile acid, induces negative feedback in the catabolism of the various sterol intermediates (6). The abnormally high production of cholestanol thereby decreases, relieving its accumulation in the plasma membranes of neural cells. Indeed, it was reported that CDCA treatment improves the neurological manifestations and abnormal findings of brain MRI in CTX patients (8, 23). Our CTX patient mainly developed xanthoma of the tendons; as such, her neurological, neuropsychiatric, and cardiovascular manifestations might have been slowly progressive compared to other reported cases (8, 24, 25). As the serum levels of cholestanol decreased after starting CDCA, we remain hopeful that further complications of CTX can be prevented in this case.

Conclusion

We reported a middle-aged patient with late-onset CTX harboring a novel mutation in the *CYP27A1* gene. It is important to begin CDCA treatment in patients with CTX before the onset of neurological dysfunction in order to prevent irreversible neuronal injury. Clinicians should consider CTX as a differential diagnosis when encountering patients with xanthomas of unknown etiology.

The authors state that they have no Conflict of Interest (COI).

References

- Cali JJ, Hsieh CL, Francke U, Russell DW. Mutations in the bile acid biosynthetic enzyme sterol 27-hydroxylase underlie cerebrotendinous xanthomatosis. J Biol Chem 266: 7779-7783, 1991.
- Waldman AT. Percy AK. Cetebrotendinous xanthomatosis. Up To Date. Wolter Kluwer, Alphen aan den Rijn, Netherlands, 2016.
- van Bogaert L, Scherer HJ, Epstein E. Une forme cérébrale de la cholestérinose généralisée. Masson et Cie, Paris, 1987 (in French).
- Appadurai V, DeBarber A, Chiang PW, et al. Apparent underdiagnosis of Cerebrotendinous Xanthomatosis revealed by analysis of ~60,000 human exomes. Mol Genet Metab 116: 298-304, 2015.
- 5. Leitersdorf E, Reshef A, Meiner V, et al. Frameshift and splice-

junction mutations in the sterol 27-hydroxylase gene cause cerebrotendinous xanthomatosis in Jews or Moroccan origin. J Clin Invest **91**: 2488-2496, 1993.

- Salen G, Meriwether TW, Nicolau G. Chenodeoxycholic acid inhibits increased cholesterol and cholestanol synthesis in patients with cerebrotendinous xanthomatosis. Biochem Med 14: 57-74, 1975.
- 7. vanHeijst AF, Verrips A, Wevers RA, Cruysberg JR, Renier WO, Tolboom JJ. Treatment and follow-up of children with cerebrotendinous xanthomatosis. Eur J Pediatr **157**: 313-316, 1998.
- Yahalom G, Tsabari R, Molshatzki N, Ephraty L, Cohen H, Hassin-Baer S. Neurological outcome in cerebrotendinous xanthomatosis treated with chenodeoxycholic acid: early versus late diagnosis. Clin Neuropharmacol 36: 78-83, 2013.
- Kim KS, Kubota S, Kuriyama M, et al. Identification of new mutations in sterol 27-hydroxylase gene in Japanese patients with cerebrotendinous xanthomatosis (CTX). J Lipid Res 35: 1031-1039, 1994.
- Gallus GN, Dotti MT, Federico A. Clinical and molecular diagnosis of cerebrotendinous xanthomatosis with a review of the mutations in the CYP27A1 gene. Neurol Sci 27: 143-149, 2006.
- Remaley AT, Thomas F, Stonik JA, et al. Synthetic amphipathic helical peptides promote lipid efflux from cells by an ABCA1dependent and an ABCA1-independent pathway. J Lipid Res 44: 828-836, 2003.
- Bjorkhem I, Meaney S. Brain cholesterol: long secret life behind a barrier. Arterioscler Thromb Vasc Biol 24: 806-815, 2004.
- 13. Bjorkhem I, Hansson M. Cerebrotendinous xanthomatosis: an inborn error in bile acid synthesis with defined mutations but still a challenge. Biochem Biophys Res Commun 396: 46-49, 2010.
- 14. Babiker A, Andersson O, Lund E, et al. Elimination of cholesterol in macrophages and endothelial cells by the sterol 27-hydroxylase mechanism. Comparison with high density lipoprotein-mediated reverse cholesterol transport. J Biol Chem 272: 26253-26261, 1997.
- 15. Bjorkhem I, Andersson O, Diczfalusy U, et al. Atherosclerosis and sterol 27-hydroxylase: evidence for a role of this enzyme in elimination of cholesterol from human macrophages. Proc Natl Acad

Sci U S A 91: 8592-8596, 1994.

- 16. Fu X, Menke JG, Chen Y, et al. 27-hydroxycholesterol is an endogenous ligand for liver X receptor in cholesterol-loaded cells. J Biol Chem 276: 38378-38387, 2001.
- Javitt NB. 25R,26-hydroxycholesterol revisited: synthesis, metabolism, and biologic roles. J Lipid Res 43: 665-670, 2002.
- Federico A, Dotti MT, Lore F, Nuti R. Cerebrotendinous xanthomatosis: pathophysiological study on bone metabolism. J Neurol Sci 115: 67-70, 1993.
- Zhu J, DeLuca HF. Vitamin D 25-hydroxylase four decades of searching, are we there yet? Arch Biochem Biophys 523: 30-36, 2012.
- Berginer VM, Shany S, Alkalay D, et al. Osteoporosis and increased bone fractures in cerebrotendinous xanthomatosis. Metabolism 42: 69-74, 1993.
- Kuriyama M, Fujiyama J, Kubota R, Nakagawa M, Osame M. Osteoporosis and increased bone fractures in cerebrotendinous xanthomatosis. Metabolism 42: 1497-1498, 1993.
- Martini G, Mignarri A, Ruvio M, et al. Long-term bone density evaluation in cerebrotendinous xanthomatosis: evidence of improvement after chenodeoxycholic acid treatment. Calcif Tissue Int 92: 282-286, 2013.
- 23. Berginer VM, Salen G, Shefer S. Long-term treatment of cerebrotendinous xanthomatosis with chenodeoxycholic acid. N Engl J Med 311: 1649-1652, 1984.
- 24. Clayton PT, Verrips A, Sistermans E, Mann A, Mieli-Vergani G, Wevers R. Mutations in the sterol 27-hydroxylase gene (CYP27A) cause hepatitis of infancy as well as cerebrotendinous xanthomatosis. J Inherit Metab Dis 25: 501-513, 2002.
- **25.** Cruysberg JR, Wevers RA, van Engelen BG, Pinckers A, van Spreeken A, Tolboom JJ. Ocular and systemic manifestations of cerebrotendinous xanthomatosis. Am J Ophthalmol **120**: 597-604, 1995.

The Internal Medicine is an Open Access article distributed under the Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License. To view the details of this license, please visit (https://creativecommons.org/licenses/by-nc-nd/4.0/).

© 2018 The Japanese Society of Internal Medicine Intern Med 57: 1611-1616, 2018