HETEROCYCLES, Vol. 97, No. 2, 2018, pp. 916 - 930. © 2018 The Japan Institute of Heterocyclic Chemistry Received, 15th February, 2018, Accepted, 20th March, 2018, Published online, 13th April, 2018 DOI: 10.3987/COM-18-S(T)68

SYNTHESIS OF 4,5-DISUBSTITUTED PYRANO[3,4-b]PYRROL-7(1*H*)-ONES VIA SONOGASHIRA–HAGIHARA CROSS-COUPLING OF *N*-BENZENESULFONYL-3-BROMO-1*H*-PYRROLE-2-CARBOXYLATE AND SUBSEQUENT IODINE-MEDIATED CYCLIZATION

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Abstract method 4,5-disubstituted А for the synthesis of pyrano[3,4-b]pyrrol-7-(1H)-ones has been developed in this study. The key reactions involved are the Sonogashira-Hagihara cross-coupling of methyl N-benzenesulfonyl-3-bromo-1H-pyrrole-2-carboxylate with terminal alkynes, followed by the iodine-mediated cyclization of 3-alkynylated N-benzenesulfonyl-1H-pyrrole-2-carboxylates. The thus-obtained 5-substituted 4-iodopyrano [3,4-b] pyrrol-7(1H)-ones could be converted to 4,5-disubstituted pyrano[3,4-b]pyrrol-7(1H)-ones via the Suzuki-Miyaura or Sonogashira-Hagihara cross-coupling reactions.

INTRODUCTION

Heterocyclic compounds possessing a common pyrano[3,4-*b*]pyrrol-7(1*H*)-one ring system have been isolated from natural sources such as prosobranch mollusk, ascidians, and sponges.¹ These include lamellarins (A–Z, α – χ , and A1–A6, including their acetate and sulfate derivatives),² ningalins A, B, E, and F,³ and bacliferin O⁴ (Figure 1). Many of these natural products and their derivatives exhibit unique structures and significant biological activities. For instance, lamellarin D shows potent cytotoxicity against cancer cell lines, including multi-drug-resistant phenotypes.⁵ The strong correlation observed between the cytotoxicity and topoisomerase I inhibition indicates that DNA topoisomerase I is a major molecular target of lamellarin D in cancer cells.⁶ Lamellarin D also induces apoptosis of cancer cell lines by directly inhibiting the mitochondrial function.⁷ In contrast, lamellarin N strongly inhibits

This paper is dedicated to Professor Dr. Kiyoshi Tomioka on the occasion of his 70th birthday.

several protein kinases, such as CDK1, CDK5, GSK-3, PIM1, and DYRK1A, relevant to cancer and neurodegenerative diseases,⁸ whereas lamellarin α 20-sulfate and other related lamellarin sulfates exhibit anti-HIV-1 activities at noncytotoxic concentrations by inhibiting the virus entry⁹ or integration steps.^{2j,5c} In addition, ningalin B and its hexamethyl ether display multi-drug-resistance (MDR) reversal activity.¹⁰ Due to their unique structures and significant biological activities, the synthesis of these compounds has attracted considerable amount of attention from organic and medicinal chemists in recent years. As a result, several synthetic methods have been developed hitherto.^{11,12} Although these approaches are useful for the preparation of lamellarin and ningalin, most of them involve the construction of a 4,5-benzo-fused pyrano[3,4-*b*]pyrrol-7(1*H*)-one scaffold. In order to prepare various types of lamellarin and ningalin analogues for lead discovery and/or optimization in medicinal chemistry, it is necessary to develop methods via the construction of a non-benzo-fused pyrano[3,4-*b*]pyrrol-7(1*H*)-ones **1**, starting from the readily available methyl *N*-benzenesulfonyl-3-bromo-1*H*-pyrrole-2-carboxylate (**2**).^{110,14}



Figure 1. Examples of natural products possessing a common pyrano[3,4-b]pyrrol-7(1H)-one scaffold

RESULTS AND DISCUSSION

The key step in the synthesis of 4,5-disubstituted pyrano[3,4-*b*]pyrrol-7(1*H*)-ones **1** from methyl *N*-benzenesulfonyl-3-bromo-1*H*-pyrrole-2-carboxylate (**2**) is the construction of a 2-pyrone ring at the 2and 3-positions of the preexisting pyrrole ring. Yao and Larock reported the highly efficient synthesis of various substituted isocoumarins and 2-pyrones via the electrophilic cyclization of o-(1-alkynyl)benzoates and (*Z*)-2-alken-4-ynoates using iodine as an electrophilic source; we utilized this method in our synthesis.¹⁵ The preparation of **1** from **2** is shown retrosynthetically in Scheme 1. Compound **1** was obtained from 5-substituted 4-iodopyrano[3,4-*b*]pyrrol-7(1*H*)-one **3** by Pd-catalyzed reactions such as Suzuki–Miyaura and Sonogashira–Hagihara cross-couplings.^{16,17} The 2-pyrone ring scaffold of **3** was constructed via the iodine-mediated 6-*endo-dig* electrophilic cyclization of 3-alkynylated pyrrole-2-carboxylate **4**.¹⁸ Finally, 3-alkynylated pyrrole-2-carboxylate **4** was prepared from **2** via the Sonogashira–Hagihara cross-coupling with terminal alkynes **5**.



Based on the retrosynthetic analysis, we first examined the Sonogashira–Hagihara cross-coupling of 3-bromopyrrole **2** with trimethylsilylacetylene (**5a**), the results of which are summarized in Table 1. Initially, various solvents were screened under the standard Sonogashira–Hagihara cross-coupling conditions [Pd(PPh₃)₂Cl₂ (1 mol%), CuI (2 mol%), 80 °C, 20 h] (entries 1–4).¹⁹ The 3-(trimethylsilyl)ethynylated pyrrole **4a** was obtained in moderate yields (entries 1 and 2) using secondary diethylamine or bidentate N,N,N',N'-tetramethylethylenediamine (TMEDA) as a solvent. When the reaction was performed in tertiary diisopropylethylamine and triethylamine, the yield of **4a** was drastically improved to 90 and 92%, respectively (entries 3 and 4). Next, other Pd-based catalyst systems were screened using triethylamine as a solvent but the yield of product **4a** did not improve (entries 5–7).²⁰ Thus, the conditions shown in entry 4 were considered optimal.

Having established the optimal reaction conditions for the Sonogashira–Hagihara cross-coupling of 3-bromopyrrole 2, we examined the Sonogashira–Hagihara cross-coupling of 2 with different terminal alkynes 5 (Table 2). Treatment of 2 with phenylacetylene (5b) (2.4 equiv) in the presence of $Pd(PPh_3)_2Cl_2$ (1 mol%) and CuI (2 mol%) in triethylamine at 80 °C for 20 h furnished 4b in 95% yield (entry 1). However, the other terminal alkynes 5c–e gave the 3-alkynylated products 4c–e in modest yields, along with the unreacted 2 (entries 2–4). When propargyl alcohol (5f) was used, the desired product 4f was not observed. Instead, starting material 2 was recovered in 24% yield, accompanied by the *N*-deprotected product 7 in 49% yield (entry 5). It is possible that compound 7 was produced by the nucleophilic attack of the alcohol on the sulfonyl group of 2.

N sc	Br → _{CO2} Me + ==→TMS P ₂ Ph 5a (2.4 equiv) 2	Pd catalys ligand (2 Cul (2 solv 80 °C	t (1 mol%) 2 mol%) mol%) /ent , 20 h	TMS O ₂ Me
entry	Pd catalyst	ligand	solvent	4 a (%) ^a
1	PdCl ₂ (PPh ₃) ₂	_	Et ₂ NH	63
2	$PdCl_2(PPh_3)_2$	_	TMEDA	63
3	$PdCl_2(PPh_3)_2$	_	<i>i</i> -Pr ₂ NEt	90
4	$PdCl_2(PPh_3)_2$	_	Et_3N	92
5	$Pd(PPh_3)_4$	_	Et_3N	67
6	Na ₂ [PdCl ₄]	PPh ₃	Et_3N	69
7	Na ₂ [PdCl ₄]	6 ^b	Et_3N	74

Table 1. Sonogashira–Hagihara cross-coupling of 3-bromopyrrole **2** with trimethylsilylacetylene (**5**a)

^a Isolated yield. ^b 2-(Di-*t*-butylphosphino)-1-phenylindole (**6**).

Table 2	. Sonogashira	a–Hagihara	cross-coupl	ling of	3-bromopy	rrole 2	with 1	terminal	alkv	nes f	5
	- 0	0		0					2		

	$ \begin{array}{c} & Br \\ & CO_2Me \\ & SO_2Ph \\ & 2 \end{array} $	Pc +	I(PPh ₃) ₂ Cl ₂ (1 mol%) Cul (2 mol%) Et ₃ N 80 °C, 20 h	R^1 N CO_2N SO_2Ph 4	le
entry	5	\mathbb{R}^1	4	4 (%) ^a	$2 (\%)^{a}$
1	5b	Ph	4b	95	0
2	5c	cyclohex-1-en-1-yl	4 c	33	56
3	5d	<i>n</i> -Bu	4d	47	51
4	5e	CH ₂ OTIPS	4e	56	22
5	5f	CH ₂ OH	4f	0	24 ^b

^a Isolated yield. ^b Methyl 3-bromo-1*H*-pyrrole-2-carboxylate (7) was also obtained in 49% yield.

With several types of 3-alkynylated pyrrole-2-carboxylates **4** in hand, we attempted to convert them into 5-substituted 4-iodopyrano[3,4-*b*]pyrrol-7(1*H*)-ones **3**, the results of which are summarized in Table 3. When **4b** was treated with I₂ (1.2 equiv) in CH₂Cl₂ at 30 °C for 18 h, the cyclized 4-iodopyrano[3,4-*b*]pyrrol-7(1*H*)-one **3b** was obtained in 96% yield and no 5-*exo-dig* cyclization product was observed (entry 1). Under similar conditions, compounds 4c-e gave the corresponding 4-iodopyrano[3,4-*b*]pyrrol-7(1*H*)-ones **3c**-e in good-to-excellent yields (entries 2–4). The cyclization of **4a** was slightly slower than the reaction of **4b**-e (entry 5). However, the yield of **3a** was improved to 79% on extending the reaction time to 120 h (entry 6). These results suggested that the ease of cyclization of 3-alkynylated pyrrole-2-carboxylate **4** depends on the steric hindrance around the alkynyl group.

	N SC	$ \begin{array}{c} $		$\mathcal{A}^{R^{1}}$	
entry	4	\mathbf{R}^1	3	${f 3}(\%)^{a}$	4 (%) ^a
1	4 b	Ph	3 b	96	_
2	4 c	cyclohex-1-en-1-yl	3c	93	_
3	4d	<i>n</i> -Bu	3d	92	_
4	4e	CH ₂ OTIPS	3e	74	_
5	4 a	TMS	3 a	62	27
6 ^b	4 a	TMS	3 a	79	12

Table 3. Iodine-mediated cyclization of 3-alkynylated pyrrole-2-carboxylate 4

^a Isolated yield. ^b The reaction was carried out for 120 h.

After the 4-iodopyrano[3,4-b]pyrrol-7(1*H*)-ones **3** were obtained, further conversion to 4,5-disubstituted pyrano[3,4-b]pyrrol-7(1*H*)-ones **1** was attempted (Scheme 2). For example, 4-iodo-5-phenyl-pyrano[3,4-b]pyrrol-7(1*H*)-one (**3b**) gave the corresponding C-4 substituted products **1a** and **1b** in good-to-excellent yields via the Suzuki–Miyaura and Sonogashira–Hagihara cross-coupling reactions.

In conclusion, we have developed a method for the synthesis of 4,5-disubstituted pyrano[3,4-b]pyrrol-7(1*H*)-ones (1). Key steps to this approach are the Sonogashira–Hagihara cross-coupling of methyl *N*-benzenesulfonyl-3-bromo-1*H*-pyrrole-2-carboxylate (2), followed by the iodine-mediated 6-*endo-dig*



electrophilic cyclization. This method may be utilized for the synthesis of various bioactive natural products and their analogues possessing the pyrano[3,4-b]pyrrol-7(1*H*)-one scaffold. Further studies to expand the scope of this method are in progress in our laboratory.

EXPERIMENTAL

The melting points were determined with a Yanagimoto micro melting point apparatus and were reported as obtained. IR spectra were obtained with a Thermo Nicolet Nexus 670 NT FT-IR instrument and are reported in terms of the absorption frequency (cm⁻¹). NMR spectra were recorded on a JEOL JNM-AL400 instrument (400 MHz for ¹H and 100 MHz for ¹³C NMR spectroscopies) or a Varian NMR System 500PS SN instrument (500 MHz for ¹H and 126 MHz for ¹³C NMR spectroscopies). Chemical shifts for ¹H NMR were expressed in parts per million (ppm) relative to tetramethylsilane (δ 0.0 ppm). The data from the ¹H NMR spectra are reported as follows: chemical shift (δ ppm), multiplicity (s = singlet, d = doublet, dd = double doublet, t = triplet, sep = septet, m = multiplet, br s = broad singlet), coupling constant (Hz), and integration. Chemical shifts for ¹³C NMR are expressed in ppm relative to tetramethylsilane (δ 0.0 ppm), ¹³C NMR data are reported in terms of only chemical shift. HMQC and HMBC spectra were recorded on a JEOL JMS-700N (fast atom bombardment mass spectrometry, FABMS) instrument. Column chromatography was conducted using silica gel 60N, 63–210 µm (Kanto Chemical Co., Inc.) or Chromatorex NH-DM1020 (Fuji Silysia Chemical Ltd.). Flash chromatography was conducted using silica gel 60N, 40–50 µm (Kanto Chemical Co., Inc.).

Typical procedure for Sonogashira–Hagihara cross-coupling of methyl *N*-benzenesulfonyl-3-bromo-1*H*-pyrrole-2-carboxylate (2) with trimethylsilylacetylene (5a) (Table 1). Under an argon atmosphere, a mixture of methyl *N*-benzenesulfonyl-3-bromo-1*H*-pyrrole-2-carboxylate (2) (344 mg, 1.00 mmol), trimethylsilylacetylene (5a) (339 μ L, 2.40 mmol), CuI (3.8 mg, 20 μ mol), an appropriate Pd-based catalyst (10 μ mol), and an appropriate solvent (1.0 mL) was heated in a sealed tube at 80 °C for 20 h. After cooling to rt, the mixture was diluted with CH₂Cl₂ and evaporated. The residue was diluted with CH₂Cl₂ and the mixture was washed with water and brine, dried over Na₂SO₄, and evaporated. The residue was purified by column chromatography over silica gel 60N (hexane–EtOAc = 5:1) to give **4a**. The results are summarized in Table 1.

Methyl *N*-(benzenesulfonyl)-3-[2-(trimethylsilyl)ethynyl]-1*H*-pyrrole-2-carboxylate (4a). Pale yellow granules. Mp 96–97 °C (Et₂O–hexane). IR (KBr): 2163, 1729, 1249, 1142, 851 cm⁻¹. ¹H NMR (400 MHz, CDCl₃): δ 0.22 (s, 9H), 3.78 (s, 3H), 6.41 (d, *J* = 3.4 Hz, 1H), 7.50–7.57 (m, 2H), 7.59 (d, *J* = 3.4 Hz, 1H), 7.60–7.66 (m, 1H), 7.93–7.98 (m, 2H). ¹³C NMR (100 MHz, CDCl₃): δ –0.2, 51.8, 97.0, 100.8, 114.7, 117.1, 127.1, 127.2, 128.0, 128.9, 134.0, 138.7, 159.1. Anal. Calcd for C₁₇H₁₉NO₄SSi: C, 56.48; H, 5.30; N, 3.87. Found: C, 56.48; H, 5.11; N, 3.85.

Typical procedure for Sonogashira–Hagihara cross-coupling of methyl *N*-benzenesulfonyl-3-bromo-1*H*-pyrrole-2-carboxylate (2) with terminal alkynes 5 (Table 2). Under an argon atmosphere, a mixture of methyl *N*-benzenesulfonyl-3-bromo-1*H*-pyrrole-2-carboxylate (2) (344 mg, 1.00 mmol), an appropriate terminal alkyne 5 (2.40 mmol), CuI (3.8 mg, 20 µmol), Pd(PPh₃)₂Cl₂ (7.0 mg, 10 µmol), and Et₃N (1.0 mL) was heated in a sealed tube at 80 °C for 20 h. After cooling to rt, the mixture was diluted with CH₂Cl₂ and evaporated. The residue was diluted with CH₂Cl₂ and the mixture was washed with water and brine, dried over Na₂SO₄, and evaporated. The residue was purified by column chromatography over silica gel 60N to give **4**. The results are summarized in Table 2.

Methyl *N*-(benzenesulfonyl)-3-(2-phenylethynyl)-1*H*-pyrrole-2-carboxylate (4b). According to the typical procedure, phenylacetylene (5b) (264 μL, 2.40 mmol) was reacted. After purification by flash chromatography over silica gel 60N (hexane–EtOAc = 7:1), 4b was obtained as a pale brown solid (346 mg, 95%). Recrystallization from Et₂O–hexane gave pale brown needles. Mp 110–112 °C. IR (KBr): 1720, 1365, 1246, 1140 cm⁻¹. ¹H NMR (400 MHz, CDCl₃): δ 3.83 (s, 3H), 6.48 (d, *J* = 3.4 Hz, 1H), 7.31–7.36 (m, 3H), 7.45–7.50 (m, 2H), 7.52–7.58 (m, 2H), 7.61–7.67 (m, 1H), 7.65 (d, *J* = 3.4 Hz, 1H), 7.96–8.00 (m, 2H). ¹³C NMR (100 MHz, CDCl₃): δ 52.0, 82.1, 94.9, 114.5, 117.5, 122.9, 126.3,

127.5, 128.0, 128.4, 128.7, 128.9, 131.6, 134.0, 138.7, 159.2. Anal. Calcd for $C_{20}H_{15}NO_4S$: C, 65.74; H, 4.14; N, 3.83. Found: C, 65.65; H, 3.94; N, 3.82.

Methyl *N*-(benzenesulfonyl)-3-[2-(cyclohex-1-en-1yl)ethynyl]-1*H*-pyrrole-2-carboxylate (4c). According to the typical procedure, 1-ethynylcyclohexene (5c) (282 μL, 2.40 mmol) was reacted. After chromatographic purification over silica gel 60N (hexane–toluene = 2:1 to toluene), 4c was obtained as a pale brown solid (120 mg, 33%). Recrystallization from Et₂O–hexane gave a pale brown powder. Mp 75.5–79 °C. IR (KBr): 2209, 1723, 1375, 1240, 1140 cm⁻¹. ¹H NMR (400 MHz, CDCl₃): δ 1.55–1.70 (m, 4H), 2.09–2.21 (m, 4H), 3.78 (s, 3H), 6.17–6.22 (m, 1H), 6.38 (d, *J* = 3.4 Hz, 1H), 7.50–7.56 (m, 2H), 7.60 (d, *J* = 3.4 Hz, 1H), 7.60–7.66 (m, 1H), 7.92–7.96 (m, 2H). ¹³C NMR (100 MHz, CDCl₃): δ 21.4, 22.2, 25.8, 29.0, 51.8, 79.5, 97.1, 114.5, 118.2, 120.6, 125.8, 127.5, 127.9, 128.9, 133.9, 136.2, 138.9, 159.3. Anal. Calcd for C₂₀H₁₉NO₄S: C, 65.02; H, 5.18; N, 3.79. Found: C, 64.77; H, 5.38; N, 3.55.

Methyl *N*-(benzenesulfonyl)-3-(hex-1-yn-1-yl)-1*H*-pyrrole-2-carboxylate (4d). According to the typical procedure, 1-hexyne (5d) (276 μL, 2.40 mmol) was reacted. After purification by flash chromatography over silica gel 60N (hexane–EtOAc = 4:1), 4d was obtained as a pale brown solid (161 mg, 47%). Recrystallization from Et₂O–hexane gave colorless plates. Mp 62.5–63.5 °C. IR (KBr): 2234, 1717, 1444, 1249, 1174, 1136 cm⁻¹. ¹H NMR (500 MHz, CDCl₃): δ 0.92 (t, *J* = 7.3 Hz, 3H), 1.41–1.50 (m, 2H), 1.52–1.58 (m, 2H), 2.41 (t, *J* = 7.0 Hz, 2H), 3.77 (s, 3H), 6.35 (d, *J* = 3.4 Hz, 1H), 7.51–7.56 (m, 2H), 7.58 (d, *J* = 3.4 Hz, 1H), 7.60–7.65 (m, 1H), 7.92–7.96 (m, 2H). ¹³C NMR (126 MHz, CDCl₃): δ 13.6, 19.3, 21.8, 30.6, 51.8, 73.1, 96.7, 114.8, 118.3, 126.0, 127.2, 127.9, 128.9, 133.9, 138.9, 159.4. HRMS (*m/z*) Calcd for C₁₈H₂₀NO₄S [(M+H)⁺]: 346.1113. Found: 346.1113.

Methyl *N*-(benzenesulfonyl)-3-{3-[(triisopropylsilyl)oxy]prop-1-yn-1-yl}-1*H*-pyrrole-2-carboxylate (4e). According to the typical procedure, triisopropyl(prop-2-yn-1-yloxy)silane (5e)²¹ (510 mg, 2.40 mmol) was reacted. After chromatographic purification over silica gel 60N (hexane–EtOAc = 10:1), 4e was obtained as a pale brown solid (265 mg, 56%). Recrystallization from Et₂O–hexane gave pale yellow plates. Mp 80–81 °C. IR (KBr): 1725, 1243, 1136, 1087, 1057 cm⁻¹. ¹H NMR (400 MHz, CDCl₃): δ 1.04–1.19 (m, 21H), 3.77 (s, 3H), 4.58 (s, 2H), 6.38 (d, *J* = 3.4 Hz, 1H), 7.51–7.57 (m, 2H), 7.59 (d, *J* = 3.4 Hz, 1H), 7.61–7.67 (m, 1H), 7.94–7.98 (m, 2H). ¹³C NMR (100 MHz, CDCl₃): δ 12.0, 17.9, 52.0, 52.5, 77.0, 93.7, 114.8, 117.0, 126.4, 127.1, 128.0, 128.9, 134.0, 138.8, 159.3. HRMS (*m/z*) Calcd for $C_{24}H_{34}NO_5SSi [(M+H)^+]$: 476.1927. Found: 476.1928.

Methyl 3-bromo-1*H*-pyrrole-2-carboxylate (7). ¹H NMR (500 MHz, CDCl₃): δ 3.90 (s, 3H), 6.35 (t, *J* = 2.9 Hz, 1H), 6.88 (t, *J* = 2.9 Hz, 1H), 9.31 (br s, 1H). ¹³C NMR (126 MHz, CDCl₃): δ 51.7, 103.6, 114.9, 120.1, 122.6, 160.6. These physical and spectroscopic data are in good agreement with those previously reported.²²

Typical procedure for iodine-mediated cyclization of 3-alkynylated pyrrole-2-carboxylate 4 (Table 3). Under an argon atmosphere, a mixture of 3-alkynylated pyrrole-2-carboxylate 4 (0.277 mmol), I_2 (84.2 mg, 0.332 mmol), and CH_2Cl_2 (2.0 mL) was stirred in a sealed tube for 18 h at 30 °C. After addition of 10% aqueous Na₂SO₃, the products were extracted with CH_2Cl_2 and the extract was washed with water and brine, dried over Na₂SO₄, and evaporated. The residue was purified by column chromatography over silica gel 60N to give **3**. The results are summarized in Table 3.

1-(Benzenesulfonyl)-4-iodo-5-phenylpyrano[3,4-*b***]pyrrol-7(1***H***)-one (3b).** According to the typical procedure, methyl *N*-(benzenesulfonyl)-3-(2-phenylethynyl)-1*H*-pyrrole-2-carboxylate (**4b**) (102 mg, 0.277 mmol) was reacted. After chromatographic purification over silica gel 60N (hexane–EtOAc = 3:1), **3b** was obtained as a pale brown solid (127 mg, 96%). Recrystallization from CH₂Cl₂–hexane gave colorless needles. Mp 144.5–145.5 °C. IR (KBr): 1728, 1390, 1178, 1143, 1037 cm⁻¹. ¹H NMR (500 MHz, CDCl₃): δ 6.56 (d, *J* = 3.4 Hz, 1H), 7.39–7.46 (m, 3H), 7.53–7.57 (m, 2H), 7.63–7.68 (m, 3H), 8.00 (d, *J* = 3.4 Hz, 1H), 8.16–8.20 (m, 2H). ¹³C NMR (126 MHz, CDCl₃): δ 65.2, 111.4, 114.8, 128.1, 129.1, 129.2, 129.7, 130.2, 131.8, 133.5, 134.7, 137.4, 142.3, 152.2, 155.8. HRMS (*m/z*) Calcd for C₁₉H₁₃INO₄S [(M+H)⁺]: 477.9610. Found: 477.9637.

1-(Benzenesulfonyl)-5-(cyclohex-1-en-1-yl)-4-iodopyrano[**3**,**4**-*b*]**pyrrol-7**(**1***H*)-**one** (**3c**). According to the typical procedure, methyl *N*-(benzenesulfonyl)-3-[2-(cyclohex-1-en-1yl)ethynyl]-1*H*-pyrrole-2-carboxylate (**4c**) (51.1 mg, 0.138 mmol) was reacted. After chromatographic purification over silica gel 60N (toluene), **3c** was obtained as a colorless oil (62.1 mg, 93%). IR (KBr): 1751, 1383, 1176, 1141, 1034 cm⁻¹. ¹H NMR (500 MHz, CDCl₃): δ 1.60–1.67 (m, 2H), 1.67–1.74 (m, 2H), 2.15–2.21 (m, 2H), 2.22–2.27 (m, 2H), 6.14–6.18 (m, 1H), 6.47 (d, *J* = 3.4 Hz, 1H), 7.51–7.56 (m, 2H), 7.62–7.66 (m, 1H), 7.94 (d, *J* = 3.4 Hz, 1H), 8.14–8.17 (m, 2H). ¹³C NMR (126 MHz, CDCl₃): δ 21.5, 22.2, 25.1, 26.3, 63.7, 111.3, 114.6, 129.1, 129.1, 131.5, 132.2, 134.6, 135.3, 137.5, 142.3, 152.4, 158.5. HRMS (*m/z*) Calcd for C₁₉H₁₇INO₄S [(M+H)⁺]: 481.9923. Found: 481.9907.

1-(Benzenesulfonyl)-5-butyl-4-iodopyrano[3,4-*b*]pyrrol-7(1*H*)-one (3d). According to the typical procedure, methyl *N*-(benzenesulfonyl)-3-(hex-1-yn-1-yl)-1*H*-pyrrole-2-carboxylate (4d) (66.5 mg, 0.193 mmol) was reacted. After chromatographic purification over silica gel 60N (hexane–EtOAc = 5:1), 3d was obtained as a colorless oil (81.3 mg, 92%). IR (KBr): 1745, 1382, 1175, 1141, 1034 cm⁻¹. ¹H NMR (500 MHz, CDCl₃): δ 0.92 (t, *J* = 7.4 Hz, 3H), 1.32–1.41 (m, 2H), 1.58–1.66 (m, 2H), 2.74 (t, *J* = 7.7 Hz, 2H), 6.41 (d, *J* = 3.4 Hz, 1H), 7.51–7.56 (m, 2H), 7.62–7.67 (m, 1H), 7.93 (d, *J* = 3.4 Hz, 1H), 8.13–8.17 (m, 2H). ¹³C NMR (126 MHz, CDCl₃): δ 13.7, 22.1, 29.4, 35.2, 65.5, 110.4, 114.5, 129.1, 129.1, 131.7, 134.6, 137.5, 141.8, 152.7, 159.8. HRMS (*m*/*z*) Calcd for C₁₇H₁₇INO₄S [(M+H)⁺]: 457.9923. Found: 457.9926.

1-(Benzenesulfonyl)-4-iodo-5-{[(triisopropylsilyl)oxy]methyl}pyrano[3,4-*b*]pyrrol-7(1*H*)-one (3e). According to the typical procedure, methyl *N*-(benzenesulfonyl)-3-{3-[(triisopropylsilyl)oxy]prop-1-yn-1-yl}-1*H*-pyrrole-2-carboxylate (4e) (132 mg, 0.277 mmol) was reacted. After chromatographic purification over silica gel 60N (hexane–EtOAc = 5:1), 3e was obtained as a colorless solid (120 mg, 74%). Recrystallization from Et₂O–hexane gave colorless plates. Mp 138.5–139.5 °C. IR (KBr): 1745, 1383, 1176, 1140, 1040 cm⁻¹. ¹H NMR (500 MHz, CDCl₃): δ 1.05–1.09 (m, 18H), 1.10–1.20 (m, 3H), 4.73 (s, 2H), 6.47 (d, *J* = 3.4 Hz, 1H), 7.51–7.56 (m, 2H), 7.62–7.67 (m, 1H), 7.96 (d, *J* = 3.4 Hz, 1H), 8.13–8.17 (m, 2H). ¹³C NMR (126 MHz, CDCl₃): δ 12.0, 17.9, 64.6, 66.5, 110.6, 115.4, 129.1, 129.2, 131.7, 134.7, 137.4, 141.2, 152.2, 156.0. HRMS (*m/z*) Calcd for C₂₃H₃₁INO₅SSi [(M+H)⁺]: 588.0737. Found: 588.0732.

1-(Benzenesulfonyl)-4-iodo-5-(trimethylsilyl)pyrano[**3,4-***b*]**pyrrol-7**(**1***H*)-one (**3a**). According to the typical procedure, methyl *N*-(benzenesulfonyl)-3-[2-(trimethylsilyl)ethynyl]-1*H*-pyrrole-2-carboxylate (**4a**) (100 mg, 0.277 mmol) was reacted for 120 h. After chromatographic purification over silica gel 60N (toluene to toluene–EtOAc = 10:1), **3a** was obtained as a colorless oil (104 mg, 79%). IR (KBr): 1743, 1383, 1203, 1029, 847 cm⁻¹. ¹H NMR (500 MHz, CDCl₃): δ 0.41 (s, 9H), 6.43 (d, *J* = 3.4 Hz, 1H), 7.51–7.56 (m, 2H), 7.62–7.66 (m, 1H), 7.95 (d, *J* = 3.4 Hz, 1H), 8.14–8.18 (m, 2H). ¹³C NMR (126 MHz, CDCl₃): δ –1.1, 77.4, 110.1, 116.3, 129.1, 129.2, 131.0, 134.6, 137.5, 140.2, 154.0, 166.1. HRMS (*m/z*) Calcd for C₁₆H₁₇INO₄SSi [(M+H)⁺]: 473.9692. Found: 473.9683.

1-(Benzenesulfonyl)-4-(4-methoxyphenyl)-5-phenylpyrano[3,4-*b*]**pyrrol-7**(1*H*)-one (1a). Under an argon atmosphere, a mixture of 1-(benzenesulfonyl)-4-iodo-5-phenylpyrano[3,4-*b*]**pyrrol-7**(1*H*)-one (**3b**) (40.0 mg, 83.8 µmol), 4-methoxyphenylboronic acid (**8**) (25.5 mg, 0.168 mmol), Pd(PPh₃)₄ (9.7 mg, 8.4 µmol), Na₂CO₃ (58.6 mg, 0.553 mmol), DME (3.0 mL), and degassed water (0.3 mL) was heated in a sealed tube at 85 °C for 24 h. After cooling to rt, the solvent was evaporated *in vacuo* and the residue was extracted with CH₂Cl₂. The extract was washed with water and brine, dried over Na₂SO₄, and evaporated. The residue was purified by flash chromatography over silica gel 60N (hexane–EtOAc = 3:1) to give **1a** as a colorless semisolid (20.4 mg, 53%). Recrystallization from Et₂O–hexane gave pale yellow granules. Mp 169.5–170.5 °C. IR (KBr): 1731, 1514, 1375, 1248, 1177 cm⁻¹. ¹H NMR (500 MHz, CDCl₃): δ 3.83 (s, 3H), 6.28 (d, *J* = 3.4 Hz, 1H), 6.87–6.91 (m, 2H), 7.10–7.14 (m, 2H), 7.16–7.20 (m, 2H), 7.21–7.25 (m, 1H), 7.27–7.31 (m, 2H), 7.53–7.58 (m, 2H), 7.63–7.68 (m, 1H), 7.92 (d, *J* = 3.4 Hz, 1H), 8.20–8.24 (m, 2H). ¹³C NMR (126 MHz, CDCl₃): δ 55.3, 106.9, 113.7, 114.5, 116.2, 126.1, 128.0, 129.0, 129.1, 129.1, 131.2, 132.1, 132.4, 134.5, 137.8, 141.0, 152.4, 152.7, 159.4. HRMS (*m*/₂) Calcd for C₂₆H₂₀NO₅S [(M+H)⁺]: 458.1062. Found: 458.1065.

1-(Benzenesulfonyl)-4-[2-(4-methoxyphenyl)ethynyl]-5-phenylpyrano[3,4-*b*]**pyrrol-7(1***H***)-one** (**1b**). Under an argon atmosphere, a mixture of 1-(benzenesulfonyl)-4-iodo-5-phenylpyrano[3,4-*b*]**pyrrol-**

7(1*H*)-one (**3b**) (40.0 mg, 83.8 μmol), 4-ethynylanisole (**9**) (26.1 μL, 0.201 mmol), CuI (0.32 mg, 1.7 μmol), Pd(PPh₃)₂Cl₂ (0.59 mg, 0.84 μmol), and Et₃N (1.0 mL) was heated in a sealed tube at 80 °C for 24 h. After cooling to rt, the mixture was diluted with CH₂Cl₂ and evaporated. The residue was diluted with CH₂Cl₂ and the mixture was washed with water and brine, dried over Na₂SO₄, and evaporated. The residue was purified by flash chromatography over silica gel 60N (hexane–EtOAc = 3:1) to give **1b** as a pale yellow solid (38.3 mg, 95%). Recrystallization from CH₂Cl₂–hexane gave pale yellow plates. Mp 201.5–202.5 °C. IR (KBr): 1736, 1512, 1384, 1253, 1175 cm⁻¹. ¹H NMR (500 MHz, CDCl₃): δ 3.84 (s, 3H), 6.76 (d, *J* = 3.4 Hz, 1H), 6.87–6.91 (m, 2H), 7.40–7.47 (m, 5H), 7.53–7.58 (m, 2H), 7.63–7.67 (m, 1H), 8.00 (d, *J* = 3.4 Hz, 1H), 8.13–8.17 (m, 2H), 8.17–8.21 (m, 2H). ¹³C NMR (126 MHz, CDCl₃): δ 55.4, 80.7, 96.7, 96.9, 107.1, 114.2, 114.6, 115.4, 128.1, 128.2, 129.0, 129.2, 130.3, 131.9, 132.5, 132.9, 134.6, 137.6, 140.0, 151.6, 157.4, 160.1. HRMS (*m*/*z*) Calcd for C₂₈H₂₀NO₃S [(M+H)⁺]: 482.1062. Found: 482.1060.

ACKNOWLEDGEMENTS

This work was financially supported by JSPS KAKENHI Grant Number 15K01802.

REFERENCES AND NOTES

- For reviews, see: (a) P. Cironi, F. Albericio, and M. Álvarez, *Prog. Heterocycl. Chem.*, 2005, 16, 1;
 (b) C. Bailly, *Curr. Med. Chem.-Anti-Cancer Agents*, 2004, 4, 363; (c) S. T. Handy and Y. Zhang, *Org. Prep. Proced. Int.*, 2005, 8, 411; (d) H. Fan, J. Peng, M. T. Hamann, and J.-F. Hu, *Chem. Rev.*, 2008, 108, 264; (e) D. Pla, F. Albrecio, and M. Álvarez, *Anticancer Agents in Med. Chem.*, 2008, 8, 746; (f) J. Kluza, P. Marchetti, and C. Bailly, 'Modern Alkaloids: Structure, Isolation, Synthesis and Biology,' ed. by E. Fattorusso and O. Taglialatela-Scafati, Wiley-VCH, Weinheim, 2008, pp. 171-187; (g) D. Pla, F. Albericio, and M. Álvarez, *MedChemComm*, 2011, 2, 689; (h) A.-L. Fan, W.-H. Lin, and Y.-X. Jia, *J. Chin. Pharm. Sci.*, 2011, 20, 425; (i) T. Fukuda, F. Ishibashi, and M. Iwao, *Heterocycles*, 2011, 83, 491; (j) D. Imbri, J. Tauber, and T. Opatz, *Mar. Drugs*, 2014, 12, 6142; (k) C. Bailly, *Mar. Drugs*, 2015, 13, 1105.
- (a) R. J. Anderson, D. J. Faulkner, H. Cun-heng, G. D. Van Duyne, and J. Clardy, *J. Am. Chem. Soc.*, 1985, **107**, 5492; (b) N. Lindquist, W. Fenical, G. D. Van Duyne, and J. Clardy, *J. Org. Chem.*, 1988, **53**, 4570; (c) A. R. Carroll, B. F. Bowden, and J. C. Coll, *Aust. J. Chem.*, 1993, **46**, 489; (d) S. Urban, M. S. Butler, and R. J. Capon, *Aust. J. Chem.*, 1994, **47**, 1919; (e) S. Urban, L. Hobbs, J. N. A. Hooper, and R. J. Capon, *Aust. J. Chem.*, 1995, **48**, 1491; (f) S. Urban and R. J. Capon, *Aust. J. Chem.*, 1995, **48**, 1491; (f) S. Urban and R. J. Capon, *Aust. J. Chem.*, 1996, **49**, 711; (g) M. V. R. Reddy, D. J. Faulkner, Y. Venkateswarlu, and M. R. Rao, *Tetrahedron*, 1997, **53**, 3457; (h) C. L. Cantrell, A. Groweiss, K. R. Gustafson, and M. R. Boyd, *Nat.*

Prod. Lett., 1999, 14, 39; (i) R. A. Davis, A. R. Carroll, G. K. Pierens, and R. J. Quinn, J. Nat. Prod., 1999, 62, 419; (j) M. V. R. Reddy, M. R. Rao, D. Rhodes, M. S. T. Hansen, K. Rubins, F. D. Bushman, Y. Venkateswarlu, and D. J. Faulkner, J. Med. Chem., 1999, 42, 1901; (k) J. Ham and H. Kang, Bull. Korean Chem. Soc., 2002, 23, 163; (l) P. Krishnaiah, V. L. N. Reddy, G. Venkataramana, K. Ravinder, M. Srinivasulu, T. V. Raju, K. Ravikumar, D. Chandrasekar, S. Ramakrishna, and Y. Venkateswarlu, J. Nat. Prod., 2004, 67, 1168; (m) S. M. Reddy, M. Srinivasulu, N. Satyanarayana, A. K. Kondapi, and Y. Venkateswarlu, Tetrahedron, 2005, 61, 9242; (n) F. Plisson, X.-C. Huang, H. Zhang, Z. Khalil, and R. J. Capon, Chem. Asian J., 2012, 7, 1616; (o) H. Zhang, M. M. Conte, X.-C. Huang, Z. Khalil, and R. J. Capon, Org. Biomol. Chem., 2012, 10, 2656.

- (a) H. Kang and W. Fenical, *J. Org. Chem.*, 1997, **62**, 3254; (b) F. Plisson, M. Conte, Z. Khalil, X.-C. Huang, A. M. Piggott, and R. J. Capon, *ChemMedChem.*, 2012, **7**, 983.
- G. Fan, Z. Li, S. Shen, Y. Zeng, Y. Yang, M. Xu, T. Bruhn, H. Bruhn, J. Morschhäuser, G. Bringmann, and W. Lin, *Bioorg. Med. Chem.*, 2010, 18, 5466.
- (a) A. R. Quesada, M. D. G. Grávalos, and J. L. F. Puentes, *Br. J. Cancer*, 1996, 74, 677; (b) F. Ishibashi, S. Tanabe, T. Oda, and M. Iwao, *J. Nat. Prod.*, 2002, 65, 500; (c) C. P. Ridley, M. V. R. Reddy, G. Rocha, F. D. Bushman, and D. J. Faulkner, *Bioorg. Med. Chem.*, 2002, 10, 3285; (d) C. Tardy, M. Facmpré, W. Laine, B. Baldeyrou, D. García-Gravalos, A. Francesch, C. Mateo, A. Pastor, J. A. Jiménez, I. Manzanares, C. Cuevas, and C. Bailly, *Bioorg. Med. Chem.*, 2004, 12, 1697; (e) D. Pla, A. Marchal, C. A. Olsen, A. Francesch, C. Cuevas, F. Albericio, and M. Álvarez, *J. Med. Chem.*, 2006, 49, 3257; (f) M. Chittchang, P. Batsomboon, S. Ruchirawat, and P. Ploypradith, *ChemMedChem*, 2009, 4, 457; (g) L. Shen, N. Xie, B. Yang, Y. Hu, and Y. Zhang, *Eur. J. Med. Chem.*, 2014, 85, 807; (h) K. Tangdenpaisal, R. Worayuthakarn, S. Karnkla, P. Ploypradith, P. Intachote, S. Sengsai, B. Saimanee, S. Ruchirawat, and M. Chittchang, *Chem. Asian J.*, 2015, 10, 925; (i) A. Theppawong, P. Ploypradith, P. Chuawong, S. Ruchirawat, and M. Chittchang, *Chem. Asian J.*, 2015, 10, 2631.
- (a) M. Facompré, C. Tardy, C. Bal-Mahieu, P. Colson, C. Perez, I. Manzanares, C. Cuevas, and C. Bailly, *Cancer Res.*, 2003, 63, 7392; (b) E. Marco, W. Laine, C. Tardy, A. Lansiaux, M. Iwao, F. Ishibashi, C. Bailly, and F. Gago, *J. Med. Chem.*, 2005, 48, 3796; (c) S. Khiati, Y. Seol, K. Agama, I. D. Rosa, S. Agrawal, K. Fesen, H. Zhang, K. C. Neuman, and Y. Pommier, *Mol. Pharmacol.*, 2014, 86, 193.
- (a) J. Kluza, M.-A. Gallego, A. Loyens, J.-C. Beauvillain, J.-M. F. Sousa-Faro, C. Cuevas, P. Marchetti, and C. Bailly, *Cancer Res.*, 2006, 66, 3177; (b) M.-A. Gallego, C. Ballot, J. Kluza, N. Hajji, A. Martoriati, L. Castéra, C. Cuevas, P. Formstecher, B. Joseph, G. Kroemer, C. Bailly, and P. Marchetti, *Oncogene*, 2008, 27, 1981; (c) C. Ballot, J. Kluza, A. Martoriati, U. Nyman, P.

Formstecher, B. Joseph, C. Bailly, and P. Marchetti, *Mol. Cancer Ther.*, 2009, 8, 3307; (d) C. Ballot,
J. Kluza, S. Lancel, A. Martoriati, S. M. Hassoun, L. Mortier, J.-C. Vienne, G. Briand, P.
Formstecher, C. Bailly, R. Nevière, and P. Marchetti, *Apoptosis*, 2010, 15, 769.

- (a) D. Baunbæk, N. Trinkler, Y. Ferandin, O. Lozach, P. Ploypradith, S. Ruchirawat, F. Ishibashi, M. Iwao, and L. Meijer, *Mar. Drugs*, 2008, 6, 514; (b) K. Yoshida, R. Itoyama, M. Yamahira, J. Tanaka, N. Loaëc, O. Lozach, E. Durieu, T. Fukuda, F. Ishibashi, L. Meijer, and M. Iwao, *J. Med. Chem.*, 2013, 56, 7289.
- 9. H. Kamiyama, Y. Kubo, H. Sato, N. Yamamoto, T. Fukuda, F. Ishibashi, and M. Iwao, *Bioorg. Med. Chem.*, 2011, **19**, 7541.
- 10. D. L. Boger, D. R. Soenen, C. W. Boyce, M. P. Hedrick, and Q. Jin, J. Org. Chem., 2000, 65, 2479.
- 11. For selected syntheses of lamellarins possessing the pyrano [3,4-b] pyrrol-7(1*H*)-one scaffold see: (a) A. Heim, A. Terpin, and W. Steglich, Angew. Chem., Int. Ed. Engl., 1997, 36, 155; (b) M. Banwell, B. Flynn, and D. Hockless, Chem. Commun., 1997, 2259; (c) F. Ishibashi, Y. Miyazaki, and M. Iwao, Tetrahedron, 1997, 53, 5951; (d) C. Peschko, C. Winklhofer, and W. Steglich, Chem. Eur. J., 2000, 6, 1147; (e) S. Ruchirawat and T. Mutarapat, Tetrahedron Lett., 2001, 42, 1205; (f) M. Díaz, E. Guitián, and L. Castedo, Synlett, 2001, 1164; (g) P. Ploypradith, C. Mahidol, P. Sahakitpichan, S. Wongbundit, and S. Ruchirawat, Angew. Chem. Int. Ed., 2004, 43, 866; (h) S. T. Handy, Y. Zhang, and H. Bregman, J. Org. Chem., 2004, 69, 2362; (i) C. A. Olsen, N. Parera, F. Albericio, and M. Álvarez, Tetrahedron Lett., 2005, 46, 2041; (j) D. Pla, A. Marchal, C. A. Olsen, F. Albericio, and M. Álvarez, J. Org. Chem., 2005, 70, 8231; (k) N. Fujikawa, T. Ohta, T. Yamaguchi, T. Fukuda, F. Ishibashi, and M. Iwao, Tetrahedron, 2006, 62, 594; (1) P. Ploypradith, T. Petchmanee, P. Sahakitpichan, N. D. Litvinas, and S. Ruchirawat, J. Org. Chem., 2006, 71, 9440; (m) J. C. Liermann and T. Opatz, J. Org. Chem., 2008, 73, 4526; (n) L. Chen and M.-H. Xu, Adv. Synth. Catal., 2009, 351, 2005; (o) T. Ohta, T. Fukuda, F. Ishibashi, and M. Iwao, J. Org. Chem., 2009, 74, 8143; (p) K. Hasse, A. C. Willis, and M. G. Banwell, Eur. J. Org. Chem., 2011, 88; (q) Q. Li, J. Jiang, A. Fan, Y. Cui, and Y. Jia, Org. Lett., 2011, 13, 312; (r) B. L. Flynn and M. G. Banwell, Heterocycles, 2012, 84, 1141; (s) D. Imbri, J. Tauber, and T. Opatz, Chem. Eur. J., 2013, 19, 15080; (t) M. Komatsubara, T. Umeki, T. Fukuda, and M. Iwao, J. Org. Chem., 2014, 79, 529; (u) J. T. Gupton, N. Telang, J. Patteson, K. Lescalleet, S. Yeudall, J. Sobieski, A. Harrison, and W. Curry, *Tetrahedron*, 2014, 70, 9759; (v) K. Ueda, K. Amaike, R. M. Maceiczyk, K. Itami, and J. Yamaguchi, J. Am. Chem. Soc., 2014, 136, 13226; (w) T. Fukuda, D. Sato, and M. Iwao, *Heterocycles*, 2015, 91, 782; (x) C. Dialer, D. Imbri, S. P. Hansen, and T. Opatz, J. Org. Chem., 2015, 80, 11605; (y) K. B. Manjappa, J.-R. Syu, and D.-Y. Yang, Org. Lett., 2016, 18, 332; (z) T. Fukuda, M. Anzai, and M. Iwao, Heterocycles, 2016, 93, 593; (aa) T. Fukuda, T. Katae, I. Harada, and M. Iwao, *Heterocycles*, 2017, 95, 950; (ab)

K. L. Zheng, M.-Q. You, W.-M. Shu, Y.-D. Wu, and A.-X. Wu, Org. Lett., 2017, 19, 2262; (ac) D.
M. Lade, A. B. Pawar, P. S. Mainkar, and S. Chandrasekhar, J. Org. Chem., 2017, 82, 4998; (ad) K.
B. Manjappa, J.-M. Lin, and D.-Y. Yang, J. Org. Chem., 2017, 82, 7648; (ae) R. Mei, S.-K. Zhang, and L. Ackermann, Synlett, 2017, 28, 1715; (af) T. Fukuda, T. Umeki, K. Tokushima, X. Gao, Y.
Yoshida, F. Ishibashi, Y. Oku, N. Nishiya, Y. Uehara, and M. Iwao, Bioorg. Med. Chem., 2017, 25, 6563.

- For selected syntheses of ningalins possessing the pyrano[3,4-b]pyrrol-7(1*H*)-one scaffold see: (a) D. L. Boger, C. W. Boyce, M. A. Labroli, C. A. Sehon, and Q. Jin, J. Am. Chem. Soc., 1999, **121**, 54; (b) J. L. Bullington, R. R. Wolff, and P. F. Jackson, J. Org. Chem., 2002, **67**, 9439; (c) J. T. Gupton, S. C. Clough, R. B. Miller, J. R. Lukens, C. A. Henry, R. P. F. Kanters, and J. A. Sikorski, *Tetrahedron*, 2003, **59**, 207; (d) M. Iwao, T. Takeuchi, N. Fujikawa, T. Fukuda, and F. Ishibashi, *Tetrahedron Lett.*, 2003, **44**, 4443; (e) C. Peschko, C. Winklhofer, A. Terpin, and W. Steglich, *Synthesis*, 2006, 3048; (f) J. T. Gupton, B. C. Giglio, J. E. Eaton, E. A. Rieck, K. L. Smith, M. J. Keough, P. J. Barelli, L. T. Firich, J. E. Hempel, T. M. Smith, and R. P. F. Kanters, *Tetrahedron*, 2009, **65**, 4283; (g) K. Hasse, A. C. Willis, and M. G. Banwell, *Aust. J. Chem.*, 2009, **62**, 683; (h) T. Fukuda, Y. Hayashida, and M. Iwao, *Heterocycles*, 2009, **77**, 1105.
- 13. (a) P. DeShong, D. A. Kell, and D. R. Sidler, *J. Org. Chem.*, 1985, **50**, 2309; (b) T. A. Bryson, G. A. Roth, and L. Jing-hau, *Tetrahedron Lett.*, 1986, **27**, 3685; (c) A. V. Lygin, O. V. Larionov, V. S. Korotkov, and A. de Meijere, *Chem. Eur. J.*, 2009, **15**, 227; (d) M. Shimizu, K. Hirano, T. Satoh, and M. Miura, *J. Org. Chem.*, 2009, **74**, 3478; (e) K. S. Singh, S. G. Sawant, and P. H. Dixneuf, *ChemCatChem*, 2016, **8**, 1046; (f) P.-O. Delaye, J. Petrignet, E. Thiery, and J. Thibonnet, *Org. Biomol. Chem.*, 2017, **15**, 7290; (g) S. Ruiz, C. Carrera, P. Villuendas, and E. P. Urriolabeitia, *Org. Biomol. Chem.*, 2017, **15**, 8904.
- 14. (a) T. Fukuda, T. Ohta, E. Sudo, and M. Iwao, *Org. Lett.*, 2010, **12**, 2734; (b) T. Fukuda and M. Iwao, *Heterocycles*, 2012, **86**, 1261.
- 15. T. Yao and R. C. Larock, J. Org. Chem., 2003, 68, 5936.
- (a) N. Miyaura and A. Suzuki, J. Chem. Soc., Chem. Commun., 1979, 866; (b) N. Miyaura and A. Suzuki, Chem. Rev., 1995, 95, 2457.
- (a) K. Sonogashira, Y. Tohda, and N. Hagihara, *Tetrahedron Lett.*, 1975, 16, 4467; (b) R. Chinchilla and C. Nájera, *Chem. Soc. Rev.*, 2011, 40, 5084.
- (a) J. E. Baldwin, J. Chem. Soc., Chem. Commun., 1976, 734; (b) B. Godoi, R. F. Schumacher, and G. Zeni, Chem. Rev., 2011, 111, 2937; (c) K. Gilmore and I. V. Alabugin, Chem. Rev., 2011, 111, 6513.
- (a) D. Fischer, H. Tomeba, N. K. Pahadi, N. T. Patil, Z. Huo, and Y. Yamamoto, *J. Am. Chem. Soc.*, 2008, **130**, 15720; (b) N. Ando and S. Terashima, *Bioorg. Med. Chem. Lett.*, 2009, **19**, 5461; (c) K.

Hayashi, K. Yoshida, and A. Yanagisawa, J. Org. Chem., 2013, 78, 3464.

- 20. C. Torborg, A. Zapf, and M. Beller, ChemSusChem, 2008, 1, 91.
- 21. P. Magnus and K. S. Matthews, J. Am. Chem. Soc., 2005, 127, 12476.
- 22. L. C. Axford, K. E. Holden, K. Hasse, M. G. Banwell, W. Steglich, J. Wagler, and A. C. Wills, *Aust. J. Chem.*, 2008, **61**, 80.