Review Article

Immunopathogenesis of Pelvic Endo metriosis: Role of Hepatocyte Growth Factor, Macrophages and Ovarian Steroids

Khaleque Newaz Khan^{1*}, Michio Kitajima¹, Koichi Hiraki¹, Akira Fujishita², Ichiro Sekine³, Tadayuki Ishimaru⁴, Hideaki Masuzaki¹

¹Department of Obstetrics and Gynecology, Nagasaki University Graduate School of Biomedical Sciences; ²Nagasaki Municipal Hospital, Nagasaki; ³Department of Molecular Pathology, Atomic Bomb Disease Institute, Nagasaki; ⁴Sasebo Chuo Hospital, Nagasaki, Japan.

Running Head: Role of immune system and ovarian steroids in endometriosis.

Address for Correspondence and reprint request: Khaleque Newaz Khan, MD, PhD Department of Obstetrics and Gynecology Graduate School of Biomedical Sciences, Nagasaki University 1-7-1 Sakamoto, Nagasaki 852-8501, Japan Tel: +81-95-819-7363 Fax: +81-95-819-7365 Email: nemokhan@nagasaki-u.ac.jp

Table of Contents

- 1. Abstract
- 2. Introduction
- 3. Pathogenesis and natural course of pelvic endometriosis

Common concepts in pathogenesis

Role of integrins and E-cadherins

Role of heme metabolism

4. Role of hepatocyte growth factor (HGF) in endometriosis

Biology of HGF and its receptor, c-Met

- 4.2. HGF concentrations in different body fluids
- 4.3. HGF as mitogenic and angiogenic factor in endometriosis
- 4.4. HGF as a scattering factor in endometriosis

4.5. HGF as migration factor in endometriosis

- 4.6. HGF as morphogenic factor in endometriosis
- 5. Role of macrophages (M ϕ) in pelvic inflammation and growth of endometriosis
 - 5.1. Infiltration of $M\phi$ in eutopic and ectopic endometrium
 - 5.2. Production of HGF by $M \phi$
 - 5.3. Role of endotoxin or lipopolysaccharide (LPS) in M ϕ -mediated secretion of HGF and other macromolecules
 - 5.4. TLR4-mediated cytokine production and growth of endometrial cells
- 6. Bacterial endotoxin (LPS) concentrations in different body fluids
- 7. Role of ovarian steroid hormones in endometriosis
 - 7.1. Immunological role of estrogen and progesterone
 - 7.2. Estrogen (E2) and progesterone (P) levels in different body fluids
 - 7.3. Expression of E2 receptor (ER) and P receptor (PR) in endometriosis
 - 7.4. Expression of ER and PR by macrophages
 - 7.5. Production of macromolecules by E2-, P-, and LPS-stimulated M $\pmb{\Phi}$
 - 7.6. Synergistic effect between HGF and E2 in the growth of endometriosis
- 8. Conclusions
- 9. Acknowledgement
- 10. References

1. Abstract

Endometriosis, a chronic disease characterized by endometrial tissue located outside of the uterine cavity and is associated with chronic pelvic pain and infertility. However, an in-depth understanding of the pathophysiology of endometriosis is still elusive. It is generally believed that besides ovarian steroid hormones, the growth of endometriosis can be regulated by innate immune system in pelvic microenvironment by their interaction with endometrial cells and immune cells. We conducted a series of studies in perspectives of pelvic inflammation that is triggered primarily by bacterial endotoxin (lipopolysacccharide, LPS) and is mediated by toll-like receptor 4 (TLR4) and showed their involvement in the development of pelvic endometriosis. As a cellular component of innate immune system, macrophages were found to play a central role in inducing pelvic inflammatory reaction. We further reported here that peritoneal macrophages retain receptors encoding for estrogen and progesterone and ovarian steroids also participate in producing an inflammatory response in pelvic cavity and involved in the growth of endometriosis either alone or in combination with hepatocyte growth factor (HGF). As a pleiotropic growth factor, HGF retains multifunctional role in endometriosis. We describe here the individual and step-wise role of HGF, macrophages and ovarian steroid hormones and their orchestrated involvement in the immunopathogenesis of pelvic endometriosis.

Key Words: pelvic endometriosis, hepatocyte growth factor, macrophages, toll-like receptor 4, ovarian steroids, inflammation

2. Introduction

Endometriosis, the presence of functional endometrium outside of the uterine cavity, is a common disease, causing abdominal pain, dysmenorrhea, dyspareunia and infertility in about 10% of the female population.¹ Besides metaplastic transformation of endometrial and peritoneal mesothelial cells, the transplantation, implantation and growth of exfoliated menstrual debris on the peritoneal and ovarian surfaces are the widely accepted mechanisms of endometriosis.²⁻⁵ A number of literatures have already demonstrated the potential role of ovarian steroid hormones in the regeneration of endometrium after menstruation and the growth of endometriosis.^{6,7} However, as a

non-self lesion in pelvic environment, the growth or persistence of endometriosis can also be regulated by innate immune system. The mitogenesis or angiogenesis of eutopic and ectopic endometrium possibly involves an extensive interplay between endometrial cells, inflammatory cells, ovarian hormones, soluble factors and the extracellular matrix.⁸

As a cell component of innate immune system, peritoneal fluid (PF) and intact tissue derived from women with endometriosis have been shown to contain higher numbers of activated macrophages.⁹⁻¹¹ than that found in women without endometriosis. This results in the secretion of higher concentrations of growth factors including hepatocyte growth factor (HGF) and other cytokines in PF as produced by the stimulated-M ϕ in these patients.¹²⁻¹⁴ This indicates that the growth or persistence of endometriosis is a normal inflammatory response and this was established by the accumulation of inflammatory cells in grafted endometriotic lesions in mouse endometriosis model.¹⁵

Since mesenchymal cells retain estrogen receptor, production of different cytokines by endometrial stromal cells and its modulation by estrogen has been demonstrated.¹⁶ Considering that infiltrated M ϕ is one of the cell components of

endometriotic lesion in pelvic environment, reports describing expression of steroid receptors by M ϕ and the secretion of different macromolecules in response to steroid hormones are scanty. Therefore, in this review article, we discussed the orchestrated role of HGF, innate immune system and ovarian steroid hormones in inducing pelvic inflammation and consequent development of pelvic endometriosis.

3. Pathogenesis and natural course of pelvic endometriosis (Figure 1)

3.1. Common concepts in the pathogenesis of endometriosis

In addition to transplantation and implantation theory of Sampson⁴ and coelomic metaplasia theory of Meyer¹⁷, immuno-surveillance of the refluxed endometrial cells is another attractive theory for the development of pelvic endometriosis, The immune tolerance or immune defect theory could be responsible for a deficiency in the rejection of the autologous cells derived from the eutopic endometrium in the peritoneal cavity after menstrual reflux. This rejection in the clearance of endometrial cells could be contributed by a dysfunctional immune response in the pelvic cavity.¹⁸⁻²⁰

According to the retrograde menstruation theory, endometrial fragments flow back through the fallopian tubes, reach the peritoneal cavity, attach on the pelvic mesothelium, invade the peritoneum and develop into endometriotic lesions.⁴ Limited information still exists regarding early endometrial-peritoneal attachment and invasion in the development of endometriosis and is derived mainly from in vitro studies.²¹⁻²³ After overcoming a phase of immune tolerance, a key step in the development of early endometriosis is the ability of endometrial cells to adhere to mesothelium and invade the extracellular matrix. These effects are contributed by a number of intercellular adhesion molecules (ICAM) and sub-cellular matrix degrading metalloproteinases (MMPs).²⁴⁻²⁹ The expressions of these ICAMs such as integrins and E-cadherin and MMPs are already detected in cells derived from menstrual effluent, endometrium, peritoneal fluid, peritoneum, and endometriosis.^{30,31}

3.2. Role of integrins and E-cadherins in endometriosis

Integrins and E-cadherin are proteins known to mediate adhesion of cells to neighboring cells or to extracellular matrix³² play an important role in this process. The expressions of integrins and E-cadherins in both mesothelial and in endometrial cells have been demonstrated in women with endometriosis.²³ The alphavbeta3 integrins transmit signals to the cytoskeletal structures of cells and usually mediate the expression

of fibronectin and vibronectin²², and have been localized in endometriotic lesions and endometrium in women with and without endometriosis during both follicular or secretory phases.²⁶ However, agents blocking alphavbeta3 integrin activity only minimally reduce the adhesion of menstrual endometrium to extracellular matrix in vitro.³³ On the other hand CD44, a key receptor for hvaluronic acid, has been demonstrated in endometrial cells and pretreatment of mesothelial cells by hyaluronidase diminishes the binding of endometrial cells to mesothelium.³⁴ These data suggest that the hyaluronic acid–CD44 binding may have a role in the initial attachment of endometrium to peritoneal mesothelial cells. The cellular mechanisms of ectopic endometrial growth involve invasive events similar to metastatic neoplasms that require extracellular matrix degradation.^{27,35} Invasion of endometrial cells into the mesothelium occurs after initial attachment to the peritoneal wall and is favored by the endometrial expression of matrix metalloproteinases (MMPs)²⁸ that remodel the mesothelial lining of the peritoneum. There is substantial evidence that ectopic endometrium has the capability to invade the surrounding tissue. Viable endometrial cells from human endometriotic biopsies but not from human endometrial biopsies are invasive in an *in vitro* collagen invasion assay.³⁶

3.3. Role of heme metabolism in endometriosis

A potential implication of hemoglobin in the pathogenesis of peritoneal endometriosis has been recently reported.³⁷ A simple hypothesis is that hemoglobin being released into the peritoneal cavity after red blood cell lysis may activate cell adhesion molecules, induce cytokine production, cell proliferation and the process of neovascularization. Degradation of hemoglobin yields biologically active molecules, heme and its products of oxidative cleavage by heme oxygeneses (HO) such as iron, carbon monoxide, biliverdin and bilirubin. Accumulation of heme in the peritoneal cavity might have a number of deleterious effects including induction of oxidative stress, stimulation of cell adhesion, and cytokine production by macrophages. All these biological events are finally involved in the pathophysiology of endometriosis. In fact, higher levels of hemoglobin were found in the peritoneal fluid of women with endometriosis. There was no concomitant increase in bilirubin concentrations in the peritoneal fluid and HO-1 was poorly expressed in peritoneal mesothelium and macrophages.³⁷ In contrast, HO-1 and HO-2 were strongly expressed in ectopic endometrium, especially in red lesions. These results suggest that heme may be involved in the pathogenesis of endometriosis. The HO system, although expressed, might be insufficient to detoxify heme in women with endometriosis.

The lesions of early endometriosis are either transparent or translucent because they still lack formation of vasculatures around them. We named these early lesions as non-opaque lesions³⁸ because these lesions contain either of watery, serous or mucinous secretion and there is no collection of blood in the stroma by histology (Figure 2 A, B). Once cellular attachment and invasion of endometrial cells are established, the subsequent growth or maintenance of endometriotic lesions is maintained by promotion of mitogenesis and angiogenesis with the continuation of menstrual cycle. The growth promoting effect of endometriosis is contributed by an orchestrated action of estrogen and other inflammatory or proinflammatory mediators. Over proliferation of micro-vessels in the growing endometriotic lesion causes oozing of blood in the stroma of these lesions and appear as blood-filled opaque red lesions by laparoscope (Figure 2 C, D).³⁸ With the progression of time, there is deoxygenetion process from hemoglobin to metheomoglobin or hemosiderin leading to color changes of these opaque red lesions to black lesion or related lesions. In this stage, collection of blood in the stroma disappears (Figure 2 E, F).

Black lesion again changes to white lesion due to collection of bilirubin or biliverdin and collection of fibrous tissue. In this stage, gland gradually becomes smaller and stroma sometimes disappears due to deposition of fibrous tissue. Finally old lesions disappear and there is new focus of endometriosis due to continuation of menstrual reflux.³⁸ These sequential events indicate that once exfoliated, the endometrium enters into the pelvic cavity, becomes attached to the mesothelial layer and then a process of angiogenesis, heme metabolism and fibrosis ensue to maintain the natural course of endometriosis. We still don't know why these sequential events occur to change the color appearance of peritoneal lesions even there is constant and variable degree of inflammatory reaction in pelvic cavity. We speculate that the process of initiation, progression and maintenance might be different among them. The sequential events in the natural course of pelvic endometriosis are illustrated in Figure 1.

4. Role of hepatocyte growth factor (HGF) in endometriosis

4.1. Biology of HGF and its receptor, c-Met

Hepatocyte growth factor (HGF) was first discovered as a mitogen for adult hepatocytes 39,40 and is identical to scatter factor.⁴¹ HGF is a heparin-binding

glycoprotein that consists of a 60-kDa α -chain and a 30-kDa β -chain linked by disulfide bonds.⁴² Several lines of evidence have implied that HGF behaves like a pleiotropic (multi-functional) growth factor and has mitogenic (cell proliferation), motogenic (cell migration), and morphogenic (change in cell type) functions in vitro on various epithelial cells derived from rodents and humans.^{43,44} The HGF receptor is the c-met proto-oncogene product (c-Met) and is a transmembrane tyrosine kinase, a 190-kDa glycoprotein consisting of a 145-kDa membrane-spanning β -chain and a 50-kDa α -chain.⁴⁵

The presence of c-Met on human endometrial epithelial cells and endothelial cells has been reported.⁴⁶ The synthesis of HGF by mesenchymal cells, coupled with demonstrated effects on epithelial and endothelial cells, suggests a paracrine mode of action.⁴⁷ HGF considered is principal mediator to be a of the mesenchymal-epithelial/endothelial interactions that contribute to embryogenesis, organ regeneration, wound healing, and angiogenesis.^{48,49} Furthermore, HGF has been shown to stimulate the proliferation, migration and morphogenesis of endometrial epithelial cells.² Although production of HGF by alveolar macrophages and hepatic kuffper cells has been reported.⁵⁰⁻⁵² information regarding production of HGF by peritoneal macrophages is limited. Since macrophages are the potent components of innate immune system, it is reasonable to speculate that these inflammatory cells might produce HGF in pelvic environment. Considering all these observations, we postulated that HGF and c-Met expression could be relevant to the pathophysiology of endometriosis.

4.2. HGF concentrations in different body fluids

Elevations in the peritoneal fluid (PF) concentrations of HGF, vascular endothelial growth factor (VEGF) and interleukin-6 (IL-6) in advanced endometriosis have been reported.⁵³⁻⁵⁵ Previous report demonstrated that PF concentrations of HGF in women with advanced endometriosis (stage II-IV) was significantly higher than those from women without endometriosis and found no difference from women with early endometriosis (stage I-II).⁵³ The report from our laboratory demonstrated that changes in the PF levels of HGF have an association with estradiol and both of them are elevated in the early stage of endometriosis, especially in those women with endometriosis harboring dominant distribution of highly active and blood-filled (opaque lesions) red peritoneal lesions.^{14,38}

The revised American Society of Reproductive Medicine (revised-ASRM) classification describes the morphologic appearance of endometriosis for color recognition of peritoneal lesions and a scoring protocol for evaluating the advancement of endometriosis.⁵⁶ This scoring system of revised-ASRM classification mostly includes the coexistence of fibrosis in the peritoneum or the presence of ovarian endometrioma of various sizes. However, this classification still leaves the question of debate about the importance of fibrous extension of disease or tissue activity of endometriosis and regarding their association with infertility. We demonstrated for the first time that the different pelvic endometriotic lesions that are included within the same red morphologic group of the revised-ASRM classification have different biological and tissue activities. Among these lesions, opaque red lesions displayed the highest activity in women with endometriosis. This was established by the measurement of different cytokines and chemokines in the PF and immunoreaction of HGF and proliferating cell nuclear antigen (PCNA) in intact tissue.³⁸ A recent study demonstrated that women with early or advanced endometriosis as measured by r-ASRM scoring system are not associated with an increase in either serum or PF concentrations of HGF. Rather HGF levels in serum and PF were significantly increased in women harboring blood-filled red peritoneal lesions and may be clinically useful to predict the activity of pelvic endometriosis. ⁵⁷

Since retrograde reflux of menstrual blood is one of the causes of endometriosis and a higher level of HGF was detected during menstrual phase, we recently measured HGF concentrations in PF and in corresponding menstrual fluid and serum of women with and without endometriosis. A higher level of HGF in seminal fluid has been reported which is possibly involved in the migration of sperm after sexual intercourse.^{58,59} We found a significantly higher concentration of HGF in the menstrual fluid and PF of women with endometriosis when compared with that in women without endometriosis (Figure 3). Menstrual fluid level of HGF in women with endometriosis was similar to that of seminal fluid. Among the three body fluids, HGF level was the highest in the menstrual fluid, intermediate in peritoneal fluid and lowest in the serum of women with endometriosis (Figure 3). This indicates that reflux of a portion of menstrual blood in every cycle may be responsible for a substantial accumulation of HGF in PF in addition to local production of HGF by endometriotic cells and immune cells.

4.3. HGF as mitogenic and angiogenic factor in endometriosis

In addition to an increase in the immunoexpression of HGF, c-Met and VEGF in eutopic and ectopic endometrium, a parallel increase in proliferating activity and micro-vessel density was also found in both the eutopic endometrium and red pigmented lesions.⁶⁰ A significant correlation between the quantitative histogram (Q-H) scores of HGF/c-Met and PCNA was observed in both the glandular epithelium and the stroma. Like the increased mitogenic activity, HGF also revealed marked angiogenic activity as demonstrated by the significant correlation of HGF expression with total micro-vessel counts.^{60,61} These mitogenic and angiogenic activities of HGF were observed for both the eutopic and ectopic endometrium of women with endometriosis and were found to be stronger when compared with tissues derived from women without endometriosis.

4.4. HGF as a scattering factor in endometriosis

Hepatocyte growth factor has been reported to scatter malignant cells and is possibly involved in the hematogenous or lymphogenous dissemination of cancer cells.⁶²⁻⁶⁴ We also reported that a variable dose of HGF (50-100ng/mL) was also able to scatter epithelial cells and stromal cells derived from the eutopic endometria of women with and without endometriosis. Epithelial cells were easier to scatter and stromal cells were found to scatter only in a collagen non-coating culture plate (Figure **4 A**, **B**). This scattering effect in endometrium indicates that higher levels of HGF in the menstrual blood could be responsible for the scattering of cells after endometrial break down and may cause retrograde regurgitation of endometrial cells during menstruation.

4.5. HGF as motogenic (migration) factor in endometriosis

The motogenic effect of HGF on cultured endometrial epithelial cells was determined with Boyden's chamber assay. As shown in Figure **4** (**C**, **D**), addition of 50 ng/mL and 100 ng/mL of HGF were able to migrate the epithelial cells and migrated cells were significantly higher when compared with HGF non-treated cells. No difference was observed in the migration of cells between 50ng/mL and 100ng.mL of HGF. This indicates that after initial endometrial-peritoneal mesothelial attachment, locally produced HGF may be involved in the invasion of endometriotic cells deep into the peritoneum.

4.6. HGF as morphogenic factor in endometriosis

Though coelomic metaplasia theory is attractive, a conclusive proof that peritoneal epithelium can undergo spontaneous or induced metaplasia to form endometriosis-like lesion is lacking. It has been reported that endometrial epithelial cells can transform into a gland-like structure under the influence of HGF.² Another report demonstrated that HGF could alter peritoneal mesothelial cells to hemispherical cells.⁶² Both of these studies emphasized the ability of HGF to induce change in cell morphology interchangeably between endometrial epithelial cells and peritoneal mesothelial cells. These results suggest that HGF functions as a mediator of morphological transformation between mesothelial cells and endometrial epithelial cells, both of which are derived from the same embryological origin.

We demonstrated that HGF might be involved in cellular changes to the peritoneal mesothelium adjacent to pelvic endometriosis.³ We reported an array of changes occurring with pelvic peritoneum from normal flat, through cuboidal and finally to columnar cells in the peritoneum adjacent to red lesions of pelvic endometriosis.³ HGF was clearly immunolocalized in cuboidal or flat mesothelial cells adjacent to red lesions in significantly stronger intensity when compared with corresponding cells adjacent to black peritoneal lesions. The intensity of HGF immunostaining in the cudoidal or columnar cells surrounding the red lesions was correlated well with clustered

accumulation of macrophages recognized among these cells.³ The result of this study is the direct evidence to indicate that the central feature of peritoneal endometriosis is a combined effect of different phases of inflammatory reactions which are elicited by transplantation of endometrial tissue fragments and subsequent transformation of mesothelial cells into endometrioid structure via the action of HGF. The multifunctional role of HGF in the development of endometriosis is shown in Figure **5**.

5. Role of $M\phi$ in pelvic inflammation and growth of endometriosis

5.1. Infiltration of $M\phi$ in eutopic and ectopic endometrium

The retrograde reflux of menstrual debris into the pelvic cavity can induce an inflammatory response and release different chemo-attractant proteins, which in turn recruit peripheral blood mononuclear cells (PBMCs) into the peritoneal environment. These PBMCs time-dependently mature into macrophages.^{9,12} The matured macrophages (M ϕ) could be harbored either in intact tissue or in the peritoneal fluid. These M ϕ can induce an inflammatory response by their accumulation or can produce pro-inflammatory mediators in response to any exogenous or endogenous stimuli. As a

component of the innate immune system, these activated $M\phi$ with their liberated cytokines and growth factors are suitable for the growth of endometriosis.^{9,12} Although a link between the severity of endometriosis and macrophage activation has been reported.⁶⁵ information regarding the tissue infiltration of these inflammatory cells and their relationship with the revised-ASRM staging and morphologic appearance of endometriosis is limited.

Our recent study¹¹ demonstrated that tissue infiltration of $M\phi$ in the eutopic and ectopic endometrium in women with stage I-II endometriosis was significantly higher than with stage III-IV endometriosis or in control women. Red peritoneal lesions and their adjacent peritoneum had the greatest $M\phi$ concentration, compared with black or white lesions. These inflammatory cells showed a higher distribution in the secretory phase of the menstrual cycle. These results indicate that early endometriosis with red peritoneal lesions induces a higher inflammatory response in the pelvic cavity than advanced endometriosis by the increased recruitment and accumulation of $M\phi$ in these tissues and this inflammation may be extended to the peritoneum around these lesions.

The different cytokines that regulate HGF and other glycoprotein expression

are supplied by the infiltrated $M \phi$ in PF, endometrial tissues, and adjacent peritoneum.^{11,66-68} However, the relationship between $M \phi$ infiltration and HGF expression or angiogenesis in endometrial tissues is unknown. We demonstrated a significant correlation between $M \phi$ density in the eutopic endometrium and corresponding red lesions and immunoreaction of HGF or between $M\phi$ accumulation and micro-vessel density in these tissues. No relationship was found between them in control women or in other peritoneal lesions. A substantial amount of HGF was also produced by the isolated basal $M\phi$ from women with endometriosis.

The inflammatory reactions in the eutopic and ectopic endometrium suggest that the growth of endometriosis does not depend on the fibrotic extension of disease; rather, it depends on the tissue activity of endometriosis. We presume that extension of disease could be related to pelvic pain, but higher activity of endometriosis associated with abundant recruitment and infiltration of M ϕ could be related to infertility. Our results agree with those of Donnez et al.,^{69,70} who also reported that the increased tissue activity of endometriosis is associated with an increased pelvic inflammatory response. Our findings of tissue inflammatory reaction corresponded to similar reaction in PF. In

fact, PF of women with early endometriosis and those containing red lesions in the pelvic cavity harbored abundant $M\phi$ in PF when compared with that of control women, advanced endometriosis or other pigmented lesions or chocolate cysts.⁷¹

5.2. Production of HGF by MØ

The production of HGF by peritoneal $M\phi$ was confirmed at the gene and protein level in the isolated $M\phi$ and in intact tissue derived from women with and without endometriosis. We demonstrated transcriptional activity of HGF and its receptor, c-Met from the isolated peritoneal fluid $M\phi$.⁷¹ The mRNA expression of HGF was found to be significantly higher in women with endometriosis than those without endometriosis. No difference in the expression of c-Met mRNA nor phases of menstrual cycle was seen between these two groups of women (Figure 6 A). The tissue localization of HGF was demonstrated in the same position as CD68-immunoreactive $M\phi$ in the serial section of intact tissues derived from the eutopic endometria of women with endometriosis (Figure 6 B, C). This indicates that HGF is being synthesized and secreted by the infiltrated $M\phi$ of intact tissue and PF.

5.3. Role of endotoxin or lipopolysaccharide (LPS) in $M \phi$ -mediated

production of HGF and other macromolecules

We examined the ability of PF M ϕ to synthesize HGF and other macromlecules in basal conditions and after treatment with lipopolysaccharide (LPS), a bacterial endotoxin derived from the cell wall extract of Gram-negative bacteria. Since our initial study⁷² demonstrated that PF of women with endometriosis contains a higher concentration of LPS (endotoxin) than that of those without endometriosis, we speculated that LPS could be a primary inflammatory mediator of M ϕ stimulation in pelvic microenvironment. In fact, activated M ϕ synthesize and secrete variable amount of different secondary inflammatory mediators such as IL-1, IL-6, IL-10, TNF α and other growth factors in response to LPS.^{9,12,72}

We found that the production of HGF, VEGF, IL-6, and TNFa by the LPS-treated M ϕ was more remarkable in women with stage I-II endometriosis than those with stage III-IV endometriosis or control women (Figure 7). In fact, a 3-fold and a 2-fold increase in the production of HGF by M ϕ was observed after LPS treatment in women with stage I-II endometriosis and stage III-IV endometriosis, respectively, when compared with non-treated M ϕ . However, only a 1.5-fold increase in the production of

HGF was observed in the control women after LPS treatment compared with no treatment. This was also confirmed at the gene level. A 3-, 5- and 4-fold increase in the expression of HGF mRNA in $M\phi$ was found in women with endometriosis at 1, 5, and 10 ng/mL of LPS treatment, respectively.⁷³ In contrast, a 1.5- to 2-fold increase in the relative expression of HGF mRNA was found in women without endometriosis.

5.4. TLR4-mediated cytokine production and growth of endometrial cells

The major component of the outer membrane of Gram-negative bacteria, endotoxin or lipopolysaccharide (LPS), is recognized by toll-like receptor 4 (TLR4) in association with other accessory molecules.⁷⁴⁻⁷⁹ We examined the protein and gene expression of TLR4 in M ϕ , in the glandular epithelial cells and stromal cells derived from the eutopic and ectopic endometrium of women with or without endometriosis. We detected both the gene and protein expression of TLR4 in isolated M ϕ , stromal cells and epithelial cells and also in eutopic and ectopic endometria derived from women with and without endometriosis.⁸⁰⁻⁸²

In an attempt to examine that the stimulating effect of LPS in the production of all these macromolecules is mediated by TLR, we pre-treated $M\phi$ with antibody against

TLR4, then again treated them with LPS. We found that the levels of a number of cytokines and growth factors were significantly decreased in comparison with cells without blocking TLR4.^{80,81} This was confirmed at both protein and gene levels. The pretreatment of glandular epithelial cells and stromal cells, derived from eutopic and ectopic endometria, with anti-TLR4 antibody was able to significantly suppress the growth of these cells when compared with cells without anti-TLR4 antibody treatment. These results indicated that LPS-mediated inflammatory reaction and growth of endometriotic cells is mediated by TLR4. We recommend that targeting TLR4 could be involved in decreasing pelvic inflammation and growth of endometriosis.⁸³

6. Bacterial endotoxin (LPS) concentrations in differ ent body

fluids

Since LPS or bacterial endotoxin acts as a primary inflammatory mediator and consequently stimulate immune cells for the production of secondary inflammatory mediators such as cytokines, chemokines, growth factors⁹ but we don't know exactly the presence of endotoxin in pelvic environment of women with or without endometriosis. We measured endotoxin levels in the menstrual blood and corresponding peritoneal fluid

of women with and without endometriosis. We found that the concentration of bacterial endotoxin is 2-4 fold higher in the menstrual fluid when compared with that in peritoneal fluid and serum. The endotoxin level in menstrual fluid was also significantly higher in women with endometriosis than that of women without endometriosis. When we distributed endotoxin level in the peritoneal fluid according to menstrual cycle, we found the highest endotoxin level during the menstrual phase and persistence of a small amount of endotoxin during the proliferative phase or secretory phase of the menstrual cycle. This indicates that menstrual blood of women with endometriosis is highly enriched with bacterial endotoxin followed by the presence of a modest amount in the peritoneal fluid. But we do not know the exact source of endotoxin in menstrual blood in women with and without endometriosis.

There is a possibility that the lower genital tract of women with or without endometriosis is contaminated with a number of normal bacterial florae including *Escherichia coli* (*E.coli*). Therefore, we speculated that there might be an ascending migration of *E.coli* from the vaginal lumen up into the uterine cavity that causes contamination of menstrual blood and resulting in the subsequent release of its cell wall extract, endotoxin, into menstrual blood and back to the peritoneal fluid. In fact, we found that a significantly increased colony formation of *E.coli* in the menstrual blood of women with endometriosis than that in the menstrual blood of women without endometriosis.⁸⁰

7. Role of ovarian steroid hormones in endometriosis

The specific relation between ovarian steroid hormones and the development or maintenance of endometriosis remains unclear, although endometriosis occurs almost exclusively in menstruating women. Breast cancer, endometrial cancer, endometriosis, adenomyosis, and leiomyomas are diseases that are believed to grow in an estrogen-dependent fashion.⁸⁴ They commonly contain estrogen receptors (ERs), progesterone receptors (PRs) and androgen receptors. In past decades, the presence of ERs and PRs has been documented in both human endometrium and typical endometriosis using radioligand binding assay.⁸⁵⁻⁸⁷ or more recently, using immunohistochemistry.^{88,89} Using the latter method, investigators have researched whether the levels of expression of ER and PR in typical endometriosis, as defined by glandular structures and stroma, parallel those of the endometrium and whether typical endometriosis is under the same cyclic ovarian control as endometrium. Nisolle et al.⁹⁰

reported that the contents of ERs and PRs in typical endometriosis lesions were lower than those of normal endometrium, but the menstrual cyclic changes were observed on the endometriosis in synchrony with the endometrium. On the other hand, other observers found no cyclic variation in typical endometriosis.^{91,92} In addition, a marked heterogeneity in the expression of ERs and PRs in typical endometriosis from different patients was noticed; the contents of ERs and PRs differed between typical endometriosis and eutopic endometrium from the same patients.^{91,93,94} The responsiveness of endometriosis tissue to ovarian steroid hormones also differed between samples of the same patients.^{92,93} Thus, there is no consensus among investigators on the response of typical endometriosis to ovarian steroid hormones.

We have already learned that endometrial cancer, endometriosis, adenomyosis and leiomyomas contain not only ER but also aromatase, an enzyme that catalyzes the conversion of androgens to estrogens, suggesting that local estrogen production may increase the estrogen concentration.^{84,95} Together with the circulating estrogen, this stimulates the growth of tissue mediated by the ER. Because of very low levels of aromatase activity and small tissue volume, it had been difficult to detect aromatase activity in endometriotic implants. Reverse-transcription polymerase chain reactions enabled the detection of mRNA and the protein of aromatse P450 (P450arom), the major component of aromatase in such tissues. In addition, endometriotic tissue contains 17β -hydroxysteroid dehydrogenase type 1, an enzyme that converts estrone (E1) to the more potent 17β -estradiol (E2), whereas they lack 17β -HSD type 2, an enzyme responsible for the inactivation of E2 to E1, resulting in raising the local estrogen activity level.^{84,96}

A number of literatures have already demonstrated the potential role of ovarian steroid hormones in the regeneration of endometrium after menstruation and the growth of endometriosis.^{97,98} However, as a non-self lesion in pelvic environment, the growth or persistence of endometriosis can also be regulated by innate immune system. It has also been reported that the female sex hormones estradiol and progesterone modify the risk of uterine infection.^{98,99} In fact, the association between ovarian steroid hormones and innate immune system is not well described.

7.1. Immunological role of estrogen and progesterone

In recent years increasing attention has been paid to innate immunity as the primary defense system against pathogens. *Escherichia coli* (*E.coli*) are the most commonly isolated pathogenic bacteria from clinical uterine diseases in cattle and also in human vaginal cavity.^{100,101} The ascending migration of *E.coli* towards endometrial cavity is possible that may cause contamination of endometrium. In bovine uterine lumen, there are high concentrations of the pathogenic ligand of *E.coli* known as bacterial endotoxin or LPS. The endometrium provides a barrier against infection and an opportunity to detect these bacteria by innate immune receptors.

The endometrium is regulated by changing concentrations of the female sex hormones, E2 and P during the ovarian cycle, and these steroids also have a profound effect on infections.^{82,101} For example, in humans, rodents and cattle, progesterone suppresses uterine immune function by decreasing the proliferative capacity of lymphocytes as demonstrated earlier in vitro,¹⁰² thereby increasing the susceptibility to bacterial infection.⁹⁹ Conversely, E2 may play a role in the recruitment of immune cells, as more macrophages are present in the endometrium when E2 concentrations are high in rodents.¹⁰¹ However, study regarding TLR expression and its involvement in the control of steroid hormone function is still unknown. The role of E2 in infection is dependent on the species, tissue and concentration of the sex hormone. Estrogen is generally thought to confer protection against infection.⁹⁹

Besides potential role in bacterial infections, sex steroid hormones are known as modulators of the immune system and may influence development and course of a variety of autoimmune disorders.¹⁰³⁻¹⁰⁵ A rapid decline of female sex hormone production after menopause may thus affect various immune parameters. Indeed, it has been reported that menopause may be associated with systemic and local changes in T and B cell subpopulation and function^{106,107} as well as immunoregulatory cytokine production.^{108,109} Immune responses may also be affected by exogenous hormone supplementation. Experimental in vivo administration of estrogens in mice has been reported to down-regulate NK cell cytotoxicity¹¹⁰ and to be associated with viral infection or enhanced tumor growth.^{111,112} NK cell activity in women taking oral contraceptives is significantly lower compared to non-users.^{113,114} Considering the effect of HRT on NK cell-mediated cytotoxicity, there exist only few and inconsistent reports with variable effects on NK cells.^{107,115} A recent clinical study demonstrated that HRT is associated with a significant decrease of NK cell cytotoxicity and a variable change of Th1 and Th2 cytokines. These changes in individual patients did not correlate with changes of serum sex hormone levels.¹¹⁶ These findings imply that estrogen/progesterone HRT may affect cell-mediated immunity and may be a potential factor influencing development of autoimmune disorders or neoplastic diseases.

Generally, sex steroid hormones are implicated in the immune response, with estrogens as enhancers at least of the humoral immunity and androgens and progesterone as natural immunosuppressors. The role of estrogen in humoral immunity has been supported by the detection of higher levels of E2 in synovial fluid and serum of patients with rheumatoid arthritis,^{117,118} which is most probably due to an increase in local aromatase activity. It has also been demonstrated that ovarian steroids, E2 and P, equally participate in humoral immune responses by modulating asymmetric antibody synthesis.¹¹⁹ In fact, sex steroids are potentially capable of driving the balance toward a cell-mediated (Th1) or humoral (Th2) response. It has been argued that imbalance in favor of a Th1 response is fostered by low levels of estrogen and prolactin, whereas, Th2 response is promoted by high levels of estrogen and testosterone.¹²⁰

Despite the widespread associations between immunology and steroid hormones, the study of systemic interactions remains limited. Many diverse facts are still accumulating on the interactions between endocrine and immune systems in the human endometrium. Understanding the molecular pathways in endocrine immune interactions in the human endometrium and in different endocrine diseases is crucial to better clarify events such as menstrual bleeding, tissue repair and regeneration, inflammation, angiogenesis, blastocyst implantation, and progression of pregnancy. These events require a balanced regulation of endometrial differentiation, proliferation, cell survival, immune cell recruitment, apoptosis, and angiogenesis by ovarian steroids. Recently, a novel concept of endometrial dissemination has been described as a result of a neuroendocrine-immune disequilibrium in response to high levels of perceived stress caused by cardinal clinical symptoms or tissue reaction of endometriosis.^{121,122}

7.2. Estrogen (E2) and progesterone (P) levels in different body fluids

We already reported increased immunoexpressions of estrogen and progesterone receptors in eutopic endometrium and ectopic endometrial lesions from our laboratory.⁶ However, information regarding concentrations of estrogen (E2) and progesterone (P) in the peritoneal fluid (PF) and other body fluids of women with and without endometriosis is scarce. It could be possible that ovarian steroids act directly or indirectly via the production of different macromolecules in the growth or maintenance of endometriosis. Therefore, we measured E2 and P levels in the PF, serum and menstrual fluid of a group of women with and without endometriosis by a modified immulyze-enzyme amplified luminescence system and as described previously.¹²³ Estrogen and P concentrations in the seminal fluid derived from healthy volunteers were used as a positive control. We also investigated the changes of E2 and P in the PF according to revised ASRM staging and morphologic appearances of endometriosis.

We found the highest concentration of E2 and P in the PF, intermediate levels in the menstrual fluid and the lowest levels in the serum (Figure 8). Both E2 and P levels in these body fluids were higher in women with endometriosis than that in control women, irrespective of menstrual cycle, revised-ASRM staging and color appearance. Although, no significant difference was observed in E2 and P levels between these two groups of women, when we distributed the patients in a different revised-ASRM staging, PF levels of E2 in women with stage I-II endometriosis were found to be significantly higher than in women with stage III-IV endometriosis or without endometriosis.¹⁴ An increase tendency in P concentration was observed between them. When we distributed the patients into different morphologic appearances of endometriosis, PF levels of E2 and P were found to be significantly higher in women containing dominant red peritoneal lesions when compared with women containing other peritoneal lesions or women without endometriosis.¹⁴ Estradiol (E2) level in the PF of women with endometriosis in the proliferative phase was higher than that of women without endometriosis. Women with endometriosis also displayed higher P levels in the secretory phase than those in the proliferative phase. No difference was observed in serum levels E2 or P according to revised-ASRM staging or color appearances of endometriosis.

7.3. Expression of E2 receptor (ER) and P receptor (PR) in endometriosis

The expressions of estrogen receptor (ER) and progesterone receptor in the eutopic and ectopic endometria are well described.⁸⁵⁻⁹⁴ Aside from typical endometriosis, there are atypical varieties of endometriosis characterized by a lack of glandular epithelial structures or stromal cells in non-pigmented lesions,^{124,125} inclusion cyst, endosalpingiosis, reactive mesothelium, and columnar epithelial cells in the pelvic peritonea. Reports on the expression of ERs and PRs in these other peritoneal lesions are scarce, and the effect of ovarian steroids on them is not well understood.

We re-evaluated ER and PR expression in endometrium and typical endometriosis and extended our study to include other peritoneal lesions. The highest score of ERs and PRs was observed in the epithelial and stromal cells of the normal uterine endometrium at the early proliferative phase of the menstrual cycle. The ER and PR scores declined throughout the secretory phase. In typical endometriotic lesions, The ER and PR scores were constantly high independent of the menstrual cycle. The expression pattern of ER mRNA was mostly in parallel with that of ERs. In typical endometriosis, ERs and PRs were found in both glandular epithelial cells and their surrounding stromal cells. Expression of ER mRNA was found in typical endometriotic peritonea and in pelvic peritoneum with columnar epithelial cells, but not in normal pelvic peritoneum (mesothelium). Estrogen receptor and PRs were negative in mesothelium, but were positive in the nuclei of fibroblasts in the connective tissue.

Our findings indicate that the columnar type cells in mesothelium are more similar to the epithelial cells in endometrium and endometriosis than to flat mesothelium. The relation of the columnar cells with their adjacent mesothelium and endometriotic lesions appears to be a key point that will decide whether endometriosis is an implant of the endometrium or is a metaplasia of mesothelium. In our previous study, clusters of columnar cells were found in mesothelium in cases in which endometriosis was absent.¹²⁶ The study suggested that the columnar cells did not originate from endometriosis and that those found in the periphery of endometriotic lesions may manifest eventually as endometriosis. Although there is no unequivocal evidence for this path, there are some supporting data: Murphy et al.¹²⁷ found a glandular orifice adjacent to normal mesothelium, and Nakamura et al.¹²⁸ documented columnar and ciliated cells in many peritoneal infoldings that dip into sub-peritoneal stroma. We strongly presume that pelvic peritoneum may have the potentiality for metaplastic changes under the influence of steroid hormones and/or a stimulating substance such as growth factors. In fact, we already described that HGF, as a pleiotropic growth factor, has the parallel potentiality involving in cellular changes of peritoneal mesothelium to a columnar phenotype and consequent gland-like invagination mimicking endometriosis.³

7.4. Expression of ERs and PRs in macrophages

Since mesenchymal cells retain estrogen receptor, production of different cytokines by endometrial stromal cells and its modulation by estrogen has been demonstrated.¹⁶ Considering that infiltrated M ϕ is one of the cell components of endometriotic lesion in pelvic environment, reports describing expression of steroid receptors by M ϕ and the secretion of different macromolecules in response to steroid hormones are scanty.

Several reports demonstrated that these inflammatory cells retain the mRNA encoding both ER and PR.^{71,129} Majority of CD68 immunoreactive M ϕ as isolated from women with or without endometriosis showed stronger nuclear staining for ER but were less reactive to PR. In contrast, tissue localization of ER and PR was equally demonstrated in the same position of CD68 immunoreactive $M\phi$ in the serial section of intact tissues derived from the eutopic endometrium of woman with endometriosis.⁷¹ This indicates that besides glandular epithelium and stroma, ER and PR are also being synthesized and expressed by the infiltrated $M\phi$ in intact tissue. This was further confirmed at gene level and revealed that basal $M\phi$ isolated from the PF of women with or without endometriosis contained the mRNA encoding for ER and PR. No phase of the menstrual cycle dependent variation in the expression of these receptor mRNAs was evident for either group.⁷¹

7.5. Production of macromolecules by E2-, P-, and LPS-stimulated $M\phi$

Several reports have already demonstrated the potential role of ovarian steroid hormones in the regeneration of endometrium after menstruation and the growth of endometriosis.^{6,7} However, information regarding inflammatory cell mediated growth or persistence of endometriosis by ovarian steroids is limited.

We and others found that direct stimulation of $M\phi$ in culture with E2 and P resulted in a variable increase in the secretion of HGF, VEGF, IL-6 and TNF α by peritoneal fluid $M\phi$.^{71,129} The production of HGF was significantly increased by E2 in women with endometriosis than that of control women or non-treated macrophages. A marked increase in the secretion of VEGF was observed by treatment with both E2 and P in women with endometriosis and also in women without endometriosis when compared with non-treated PF macrophages. No phase of cycle differences were seen. When we performed a blocking experiment on E2 by using ER antagonist, tamoxifen, we found that tamoxifen significantly reversed the secretion of and tended to reverse the secretion of HGF by the estrogen-treated PF macrophages towards the non-treated macrophages.⁷¹ This indicates that it is the direct effect of estrogen on the PF macrophages that was able to produce significant amount of HGF and VEGF and is being mediated by ER as located on these inflammatory cells. We also found a substantial amount of increase in the levels of IL-6 and TNFa in response to ovarian steroids without displaying any significant difference with non-treated cells.⁷¹

It has been reported that activation with LPS significantly increased the amount of a number of macromolecules secreted by inflammatory cells.^{71,73} We observed that activation of basal macrophages further enhanced the response of these cells to ovarian steroids. In fact, exogenous treatment with E2 was able to further increase the amount of both HGF and VEGF secretion by PF macrophages when these cells were activated with LPS.⁷¹ Although progesterone increased the secretion of VEGF by non-activated M ϕ , it was unable to further enhance the secretion of either HGF or VEGF by activated PF macrophages.⁷¹ These results confirmed that irrespective of activation status, PF macrophages were independently stimulated to produce HGF and VEGF by E2. This also indicates that in addition to primary and secondary inflammatory mediators, ovarian steroids equally produce a pelvic inflammatory reaction mediated by macrophages. An inflammatory response and ovarian steroid hormones may function either alone or in combination to regulate the production of a variety of macromolecules by PF macrophages in pelvic microenvironment.

It is generally believed that ovarian steroid hormones are essential for the growth or persistence of ectopic endometrium and corresponding eutopic endometrium in women with endometriosis.¹³⁰ Since the major cellular constituents of PF are macrophages, comprising between 82 and 99% of the total cell population,¹³¹ it is quite reasonable to speculate that these cells may be responsive to ovarian steroids.

7.6. Synergistic effect between HGF and E2 in the growth of endometriosis

The presence of c-Met receptor and estrogen receptor in endometrial and endometriotic cells and $M\phi$ might be expected to enable these cells to respond to endogenous or exogenous HGF and E2.^{71,73,132} When we examined the effect of HGF and E2 on the proliferation of endometrial gland cells, stromal cells and $M\phi$, we found that these cells significantly proliferated in response to HGF and E2 either alone or in combination when compared with non-treated cells.⁷¹ A synergistic effect between HGF and E2 on cell proliferation was observed. A similar pattern of cell proliferation was also found in gland cells and stromal cells derived from ectopic endometrium. These results indicate that an innate immune system in pelvic environment and ovarian steroid hormones function in an orchestrated fashion and are involved in the growth of endometriosis. Our results are summarized in Figure 9.

8. Conclusions

We now know that besides steroid hormones, innate immunity plays a pivotal role in the initiation of an array of inflammatory reactions against regurgitated endometrial cells and subsequent development of peritoneal endometriosis. Currently prevalent concepts on the genesis of pelvic endometriosis are retrograde dissemination of eutopic endometrial tissues during menstruation,⁴ coelomic metaplasia of the peritoneum as a secondary Mullerian system,^{133,134} and compromised immuno-surveillance.¹⁸⁻²⁰ However, none of these theories can explain the pathogenesis of endometriosis uniformly. Based on our serial studies on the etiological role of bacterial endotoxin (LPS), we would propose a novel concept for the genesis of pelvic endometriosis via LPS/TLR4-mediated engagement of innate immune response.

According to this concept, it would appear possible to integrate two conflicting thoughts of transplantation and metaplasia as reflecting the different phases of initiation and progression of pelvic endometriosis. Transplantation and consequent implantation of regurgitated endometrial cells during menstruation may trigger strong inflammatory reaction in early endometriosis. In addition, a variety of pro-inflammatory factors are also secreted from the infiltrated mononuclear cells of innate immune system. During progression of the affected lesion, cellular changes of juxtaposed mesothelium into endometrioid cells and gland-like structures subsequently ensues and was described as metaplasia of peritoneal mesothelium.³ As a pleiotropic growth factor, HGF being produced by macrophages and stromal cells has been shown to serve this unique role. The multi-functional role of HGF can be performed with the aid of systemic or focal hormonal environment characterized by consistent estrogen synthesis. Our presenting findings demonstrate for the first time that besides other pro-inflammatory mediators, ovarian steroids also participate in the generation of a pelvic inflammatory response by producing different macromolecules including HGF by peritoneal $M\phi$. These pro-inflammatory mediators including HGF may be involved in the growth of endometriosis either alone or in combination with estrogen (Figure 9).

A complete understanding of the mechanisms of endocrine-immune cross talk

in the mammalian species and the function of innate immunity via toll-like receptor system will be helpful for the future development of innovative therapies for manipulation of endometriosis.

9. Acknowledgments

We thank Miss Kazumi Hayashida and Miss Kyoko Ishida, Department of Obstetrics and Gynecology, Nagasaki University Graduate School of Biomedical Sciences, Nagasaki, Japan, for their excellent technical assistance. This work was supported by Grants-in-Aid for Scientific Research (Grant No. 16591671 and 18591837) from the Ministry of Education, Sports, Culture, Science and Technology of Japan (to KN Khan).

10. References

1 Strathy JH, Molgaard CA, and Coulman CB: Endometriosis and infertility:

a laparoscopic study of endometriosis among fertile and infertile women. *Fertil Steril* 1982,38,667-672.

2 Sugawara J, Fukaya T, Murakami T, Yoshida H, Yajima A: Hepatocyte growth factor stimulates proliferation, migration, and lumen formation of human endometrial epithelial cells in vitro. *Biol Reprod* 1997,57,936-942.

3 Ishimaru T, Khan KN, Fujishita A, Kitajima M, Masuzaki H: Hepatocyte growth factor may be involved in cellular changes to the peritoneal mesothelium adjacent to pelvic endometriosis. *Fertil Steril*, 2004,81(suppl 1), 810-818.

4 Sampson J: Peritoneal endometriosis due to menstrual dissemination of endometrial tissue into the peritoneal cavity. *Am J Obstet Gynecol* 1927,14,422-429.

5 Thomas EJ and Prentice A: The etiology and pathogenesis of endometriosis. Reprod.

Med. Rev. 1992,1,21-36.

6 Fujishita A, Nakane PK, Koji T, Masuzaki H. Chavez RO, Yamabe T, Ishimaru T:
Expression of estrogen and progesterone in endometrium and peritoneal endometriosis:
an immunohistochemical and in situ hybridization study. *Fertil Steril* 1997,67,856-864.
7 Nisolle M, Casanas-Rouz F, Donnez J: Immunohistochemical analysis of proliferative
activity and steroid receptor expression in peritoneal and ovarian endometriosis. *Fertil Steril* 1997,68,912-919.

8 Folkman J and Klagsbrun M: Angiogenic factors. Science 1987,235,442-446.

9 Halme J, Becker S, Haskill S: Altered maturation and function of peritoneal macrophages: possible role in pathogenesis endometriosis. *Am J Obstet Gynecol* 1987,156, 783-789.

10 Khan KN, Fujishita A, Kitajima M, Masuzaki H, Sekine I, Ishimaru T: Infiltrated macrophage activity in intact tissue of endometriosis. *Proceedings of Endometriosis* 2002,23,137-142 (in Japanese).

11 Khan KN, Masuzaki H, Fujishita A, Kitajima M, Sekine I, Ishimaru T: Differential macrophage infiltration in early and advanced endometriosis and adjacent peritoneum.

Fertil Steril 2004,81,652-661.

12 Halme J, White C, Kauma S, Estes J, Haskill S: Peritoneal macrophages from patients with endometriosis release growth factor activity in vitro. *J Clin Endocrinol Metab* 1988,66,1044-1049.

13 Halme J: Release of tumor necrosis factor-alpha by human peritoneal macrophages in vivo and in vitro. *Am J Obstet Gynecol* 1989,161,1718-1725.

14 Khan KN, Masuzaki H, Fujishita A, Hamasaki T, Kitajima M, Hasuo A, Ishimaru T: Association of interleukin-6 and estradiol with hepatocyte growth factor in peritoneal fluid of women with endometriosis. *Acta Obstet Gynecol Scand* 2002,81,764-771.

15 Kitajima M, Khan KN, Fujishita A, Masuzaki H, Ishimaru T: Histomorphometric alteration and cell-type specific modulation of arylhydrocarbon receptor and estrogen receptor expression by 2,3,7,8-tetrachlorodibemzo-p-dioxin and 17b-estradiol in mouse experimental model of endometriosis. *Reprod Toxicol.* 2004,18,793-801.

16 Tabibzadeh SS, Santhanan V, Sehgal PB, May LT: Cytokine induced production of IFN- β 2/IL-6 by freshly implanted human endometrial stromal cells: modulation by estradiol 17 β . *J Immunol* 1989,42,3134-3139.

17 Meyer R: Ueber den Stand der Frage der Adenomyositis und Adenomyom im Allgemeinen und insbesonderes ueber Adenomyositis seroepithelialis und Adeno-myometritis sarcomatosa. *Zbl Gynaekol* 1919,36,745-750.

18 Oosterlynck DJ, Cornillie FJ, Waer M, Vandeputte M, Koninckx PR: Women with endometriosis show a defect in natural killer activity resulting in a decreased cytotoxicity to autologous endometrium. *Fertil Steril* 1991,56,45-51.

19 Lebovic DI, Mueller MD, Taylor RN: Immunobiology of endometriosis. *Fertil Steril* 2001,75,1-10.

20 Giudice LC, Kao L: Endometriosis. Lancet 2004,364,1789-1799.

21 Witz CA, Montoya-Rodriguez AI, Schenken RS: Whole peritoneal explants: a novel model of the early endometriosis lesion. *Fertil Steril* 1999,71,56-60.

22 Debrock S, Vander P, Meuleman C, Moerman PH, Hill JA, D'Hooghe TM: In vitro adhesion of endometrium to autologous peritoneal membranes: effect of the cycle phase and the stage of endometriosis. *Hum Reprod* 2002,17,2523-2528.

23 Groothuis PG, Koks CA, de Goeij AF, Dunselman GA, Arends JW, Evers JL. Adhesion of human endometrial fragments to peritoneum in vitro. *Fertil Steril* 1999,71,1119-1124.

24 Lessey BA, Damjanovich L, Coutifaris C: Integrin adhesion molecules in the human endometrium. Correlation with the normal and abnormal menstrual cycle. *J Clin Invest* 1992,90,188-195.

25 Lessey BA, Castel baum AJ, Sawin SW, Buck CA, Schinnar R, Bilker W, Strom BL: Aberrant integrin expression in the endometrium of women with endometriosis. *J Clin Endocrinol Metab* 1994,79,643-649.

26 Regidor PA, Vogel C, Regidor M, Schindler AE, Winterhager E: Expression pattern of integrin and adhesion molecules in endometriosis and human endometrium. *Hum Reprod* 1998,4,710-718.

27 Chung HW, Wen Y, Chun SH, Nezhat C, Woo BH, Polan ML: Matrix metalloproteinase-9 and tissue inhibitors of metalloproteinase-3 mRNA expression in ectopic and eutopic endometrium in women with endometriosis: a rationale for endometriotic invasiveness *Fertil Steril* 2001,75,152-159.

28 Sillem M, Prifti S, Koch A, Neher M, Jauckus J, Runnebaum B: Regulation of matrix metalloproteinases and their inhibitors in uterine endometrial cells of patients with and

without endometriosis. Eur J Obstet Gynecol Reprod Biol 2001,95,167-174.

29 Osteen KG, Keller NR, Feltus FA, Melner MH: Paracrine regulation of matrix metalloproteinase expression in the normal human endometrium. *Gynecol Obstet Invest* 1999,48,2-13.

30 van der Linden PJQ, van der Linden EPM, de Goeij AFPM, Ramaekers FC, Dunselman GAJ: Expression of integrins and E-cadherin in cells from menstrual effluent, endometrium, peritoneal fluid, peritoneum, and endometriosis. *Fertil Steril* 1994,61,85-90.

31 Witz CA, Takahashi A, Montoya-Rodriguez IA, Cho S, Schenken RS: Expression of the $\alpha 2\beta 1$ and $\alpha 3\beta 1$ integrins at the surface of mesothelial cells: a potential attachment site of endometrial cells. *Fertil Steril* 2000,74,579-584.

32 Sillem M, Prifti S, Monga B, Arslic T, Runnebaum B: Integrin-mediated adhesion of uterine endometrial cells from endometriosis patients to extracellular matrix proteins is enhanced by tumor necrosis factor alpha (TNF alpha) and interleukin-1 (IL-1). *Eur J Obstet Gynecol Reprod Biol* 1999,87, 123-127.

33 Koks CAM, Groothuis PG, Dunselman GA, De Goiej AFPM & JLH Evers:

Adhesion of menstrual endometrium to extracellular matrix: the possible role of integrin $\alpha_6\beta_1$ and laminin interaction. *Mol Hum Reprod* 2000,6, 170-7.

34 Dechaud H, Witz CA, Montoya-Rodriguez IA, Degraffenreid LA,

Schenken RS: Mesothelial cell-associated hyaluronic acid promotes adhesion of endometrial cells to mesothelium. *Fertil Steril* 2001,76, 1012–8.

35 Mulayim N, Savlu A, Guzeloglu-Kayisli O, Kayisli UA & A Arici. Regulation of endometrial stromal cell matrix metalloproteinase activity and invasiveness by interleukin-8. *Fertil Steril* 2004,81, 904-911.

36 Gaetje R, Kotzian S, Herrmann G, Baumann R & A Starzinski-Powitz:

Nonmalignant epithelial cells, potentially invasive in human endometriosis, lack the tumor suppressor molecule E-cadherin. *Am J Pathol* 1997,150, 461-467.

37 Langendonckt AV, Casanas-Roux F, Dolmans M-M, Donnez J: Potential involvement of hemoglobin and heme in the pathogenesis of peritoneal endometriosis. *Fertil Steril* 2002,77,561-570.

38 Khan KN, Masuzaki H, Fujishita A, Kitajima M, Sekine I, Ishimaru T: Higher activity by opaque endometriotic lesions than non-opaque lesions in women with endometriosis. Acta Obstet Gynecol Scand 2004,83,375-382.

39 Miyazawa K, Tsubouchi H, Naka D, Takahashi K, Okigaki M, Arakaki N: Molecular cloning and sequence analysis of cDNA for human hepatocyte growth factor. *Biochem Biophys Res Commun* 1998,163,967-973.

40 Nakamura T, Nishizawa T, Hagiya M, Seki T, Shimonishi M, Sugimura A: Molecular cloning and expression of human hepatocyte growth factor. *Nature* 1989,342,440-443.

41 Wiedner KM, Arakaki N, Vandekereckhove J, Weingart S, Hartmann G, Rieder H: Evidence for the identity of human scatter factor and human hepatocyte growth factor. *Proc Natl Acad Sci USA* 1991,88,7001-7005.

42 Gherardi E, Gray J, Stocker M, Perryman M, Furlong R: Purification of scatter factor, a fibroblast-derived basic protein which modulates epithelial interactions and movement. *Proc Natl Acad Sci USA* 1989,86,5844-5848.

43 Nakamura T, Teramoto H, Ichihara A: Purification and characterization of a growth factor from rat platelets from mature parenchymal hepatocytes in primary culture. *Proc Natl Acad Sci USA* 1986,83,6489-6493.

44 Tajima H, Matsumoto K, Nakamura T: Regulation of cell growth and motility by

hepatocyte growth factor and receptor expression in various cell species. *Exp Cell Res* 1992,202,423-431.

45 Bottaro DP, rubin JS, Falleto DL, Chan AML, Kmeicik TE, Vande Woude GF: Identification of the hepatocyte growth factor receptor as the c-met proto-oncogene product. *Science* 1991,251,802-804.

46 Wagatsuma S, Konno R, Sato S, Yajima A: Tumor angiogenesis, hepatocyte growth factor, and c-Met expression in endometrial carcinoma. *Cancer* 1998,82,520-530.

47 Sonnenberg E, Meyer D, Weidner KM, Birchmeier C: Scatter factor/ hepatocyte growth factor and its receptor, c-met tyrosine kinase, can mediate a signal exchange between mesenchyme and epithelia during mouse development. *J Cell Biol* 1993,123,223-235.

48 Grant SD, Kleinman HK, Goldberg ID, Bhargava MM, Nickoloff BJ, Kinsella JL: Scatter factor induces blood vessel formation in vivo. *Proc Natl Acad Sci USA* 1993,90,1937-1941.

49 Rosen EM, Nigam SK, Goldberg ID: Scatter factor and the c-Met receptor: a paradigm for mesenchymal/epithelial interaction. *J Cell Biol* 1994,127,1783-1787.

50 Skrtic S, Wallenius V, Ekberg S, Brenzel A, Gressner AM, Jansson JO: Hepatocyte-stimulated expression of hepatocyte growth factor (HGF) in cultured rat hepatic stellate cells. *J Hepatol.*, 1999,30(1), 115-124.

51 Morimoto K, Amano H, Sonoda F, Baba M, Senba M, Yoshimine H, Yamamoto H, Ii T, Oishi K, Nagatake T: Alveolar macrophages that phagocytose apoptotic neutrophils produce hepatocyte growth factor during bacterial pneumonia in mice. *Am J Respir Cell Mol Biol.*, 2001,24(5), 608-615.

52 Crestani B, Dehoux M, Hayem G, Lecon V, Hochedez F, Marchal J, Jaffre S, Stern JB, Durand G, Valeyre D, Fournier M, Aubier M: Differential role of neutrophils and alveolar macrophages in hepatocyte growth factor production in pulmonary fibrosis. *Lab Invest*. 2002,82(8), 1015-1022.

53 Osuga Y, Tsutsumi O, Okagaki R, Takai Y, Fujimoto A, Suenaga A, Maruyama M, Momoeda M, Yano T, Taketani Y: Hepatocyte growth factor concentrations are elevated in peritoneal fluid of women with endometriosis. *Hum. Reprod.* 1999,14,1611-613.

54 Mahnke JL, Dawood Y, Huang JH: Vascular endothelial growth factor and interleukin-6 in peritoneal fluid of women with endometriosis. *Fertil Steril*.

2000,73,166-170.

55 Sugawara J, Fukaya T, Murakami T, Yoshida H, Yajima A: Increased secretion of hepatocyte growth factor by eutopic endometrial stromal cells in women with endometriosis. *Fertil Steril* 1997,68,468-472.

56 The American Society for Reproductive Medicine: Revised American Society for Reproductive Medicine classification of endometriosis: 1996. *Fertil. Steril.* 1997,67,817-821.

57 Khan KN, Masuzaki H, Fujishita A, Kitajima M, Hiraki K, Miura S, Sekine I, Ishimaru T: Peritoneal fluid and serum levels of hepatocyte growth factor may predict the activity of endometriosis. *Acta Obstet Gynecol Scand* 2006,85,458-466.

58 Kitamura M, Matsumiya K, Yamanaka M, Matsumoto K, Okuyama A: Effect of hepatocyte growth factor on sperm motility. *Am J Reprod Immunol* 2000,44(4),193-196. 59 Wiltshire EJ, Flaherty SP, Couper RT. Hepatocyte growth factor in human semen and its association with semen parameters. *Hum Reprod* 2000,15(7),1525-1528.

60 Khan KN, Masuzaki H, Fujishita A, Kitajima M, Sekine I, Ishimaru T: Immunoexpression of hepatocyte growth factor and c-Met receptor in eutopic endometrium predicts the activity of ectopic endometrium. *Fertil Steril*, 2003,79,173-181.

61 Yoshida S, Harada T, Mitsunari M, Iwabe T, Sakamoto Y, Tsukihara S, Iba Y, Horie S, Terakawa N: Hepatocyte growth factor/Met system promotes endometrial and endometriotic stromal cell invasion via autocrine and paracrine pathways. *J Clin Endocrinol Metab* 2004,89(2),823-832.

62 Yashiro M, Chung YS, Inoue T, Nishimura S, Matsuoka T, Fujihara T,: Hepatocyte growth factor (HGF) produced by peritoneal fibroblasts may effect mesothelial cell morphology and promote peritoneal dissemination. *Int J Cancer* 1996,67,289-293.

63 Bae-Jump V, Segreti EM, Vandermolen D, Kauma S: Hepatocyte growth factor (HGF) induced invasion of endometrial carcinoma cell line in vitro. *Gynecologic Oncology* 1999,7,265-272.

64 Kataoka H, Hamasuna R, Itoh H, Kitamura N, Koono M: Activation of hepatocyte growth factor/scatter factor in colorectal carcinoma. *Cancer Research* 2000,60,6148-6159.

65 Becker IL, Widen RH, Mahan CS, Yeko TR, Parsons AK, Spellacy WN: Human

peritoneal macrophage and T lymphocyte populations in mild and severe endmetriosis. *Am J Reprod immunol* 1995,34,179-187.

66 Khan KN, Fujishita A, Kitajima M, Hasuo A, Miyamura Y, Masuzaki H, Ishimaru T: Immunoexpression of hepatocyte growth factor and c-met receptor in eutopic endometrium predicts the activity of ectopic endometrium. In: Genazzani AR, Artini PG, Petraglia F eds. *Recent Research in Gynecological Endocrinology*. New York: Parthenon, 2000,111-114.

67 Khan KN, Fujishita A, Kitajima M, Masuzaki H, Sekine I, Ishimaru T: Infiltrated macrophage activity in intact tissue of endometriosis. *Proc Endometriosis* 2002,23,137-142 (in Japanese).

68 Ishimaru T, Fujishita A, Khan KN, Kitajima M, Masuzaki H, Matsuyama T: Morphological appearance and growth of pelvic endometriosis. *Acta Obst Gynae Jpn* 2002,54,1158-1163 (in Japanese).

69 Donnez J, Nisolle M, Smoes P, Gillet N, Beguin S, Casanas-Roux F: Peritoneal endometriosis and 'endometriotic' nodule of rectovaginal septum are two different entities. *Fertil Steril* 1996,66,362-368.

70 Donnez J, Smoes P, Gillerot S, Casanas-Roux F, Nisolle M: Vascular endothelial growth factor in endometriosis. *Hum Reprod* 1998,13,1686-1690.

71 Khan KN, Masuzaki H, Fujishita A, Kitajima M, Sekine I, Matsuyama T, Ishimaru T: Estrogen and progesterone receptor expression in macrophages and regulation of hepatocyte growth factor by ovarian steroids in women with endometriosis. *Hum Reprod* 2005,20,2004-2013.

72 Khan KN, Fujishita A, Kitajima M, Masuzaki H, Matsuyama T, Sekine I, Ishimaru T: Detection of *Escherichia coli* in menstrual blood and its possible involvement in the growth of endometriosis. *Endo2003, The 85th Annual Meeting of American Endocrine Society*, 2003,383:564.

73 Khan KN, Masuzaki H, Fujishita A, Kitajima M, Kohno T, Sekine I, Matsuyama T, Ishimaru T: Regulation of hepatocyte growth factor by basal and stimulated macrophages in women with endometriosis. *Hum Reprod*, 2005,20,49-60.

74 Akira S and Takeda K: Toll-like receptor signaling. *Nat Rev Immunol* 2004,4,499-511.
75 Check W: Innate immunity depends on Toll-like receptors. *ASM News* 2004,70,317-22.

76 Takeda K and Akira S: Toll-like receptors in innate immunity. *Int. Immunol* 2005, 7,1-14.

77 Wira CR, Fahey JV, Sentman CL, Pioli PA, Shen L: Innate and adaptive immunity in female genital tract: cellular responses and interactions. *Immunol Reviews* 2005,206,306-35.

78 Aflatoonian R, Tuckerman E, Elliot SL, Bruce C, Aflatoonian A, Li TC, Fazeli A: Menstrual cycle-dependent changes of toll-like receptors in endometrium. *Hum Reprod* 2007,22,586-93.

79 Fazeli A, Bruce C, Anumba DO: Characterization of Toll-like receptors in the female reproductive tract in humans. *Hum Reprod* 2005,20(5),1372-78.

80 Khan KN, Fujishita A, Kitajima M, Hiraki K, Masuzaki H, Sekine I, Matsuyama T, Ishimaru T: Detection of *Escherichia coli* in menstrual blood and endotoxin in peritoneal fluid: an implication in pelvic inflammation and toll-like receptor 4 (TLR4)-mediated growth of endometriosis. *Eur J Obstet Gynecol Reprod Biol* 2005,123,S15-S16.

81 Khan KN, Kitajima M, Imamura T, Hiraki K, Fujishita A, Matsuyama T, Sekine I, Ishimaru T, Masuzaki H: Toll-like receptor 4 (TLR4)-mediated growth of endometriosis by endogenous heat shock protein 70. *Proceedings of the Endocrine Society's* 88th Annual *Meeting*. 2006,256-257.

82 Hirata T, Osuga Y, Hirota Y, Koga K, Yoshino O, Harada M, Morimoto C, Yano T, Nishii O, Tsutsumi O, Taketani Y: Evidence for the presence of Toll-like receptor 4 system in the human endometrium. *J Clin Endocrinol Metab* 2005,90,548-556.

83 Kiechl S, Lorenz E, Reindl M, Wiedermann CJ, Oberhollenzer F, Bonora E, Willeit J,

Schwartz DA: Toll-like receptor 4 polymorphisms and atherogenesis. *N Engl J Med* 2002,347,185-192.

84 Kitawaki J, Kado N, Ishihara H, Koshiba H, Kitaoka Y, Honjo H: Endometriosis: the pathophysiology as an estrogen-dependent disease. *J Steroid Biochem Mol Biol* 2003,83,149-155.

85 Tamaya T, Motoyama T, Ohono Y, Ide N, Tsurusaki T, Okada H: Steroid receptor
levels and histology of endometriosis and adenomyosis. *Fertil Steril* 1979,31,396-400.
86 Jane O, Kauppila A, Kokko E, Lantto T, Ronberg L, Vihko R: Estrogen and
progesterone receptors in endometriosis lesions: comparison with endometrial tissue. *Am J Obstet Gynecol* 1981,141,562-566.

87 Gould SF, Shannon JM, Cunha GR: Nuclear receptor binding sites in human endometriosis. *Fertil Steril* 1983,39,520-524.

88 McClellan MC, West NB, Tacha DF, Green GL, Brenner RM: Immunocytochemical localization of estrogen receptors in the macaque reproductive tract with monoclonal antiestrophilins. *Endocrinology* 1984,114,2002-2014.

89 Okulicz WC, Savasta AM, Hoberg LM, Longcope C: Biochemical and immunohistochemical analysis of estrogen and progesterone receptors in the rhesus monkey uterus during the proliferative and secretory phases of artificial menstrual cycles. *Fertil Steril* 1990,53,913-920.

90 Nisolle M, Menten Y, Casanas-Roux F, Mathieu PE, Wyns C, Donnez J: Immunohistochemical analysis of estrogen and progesterone receptors in endometrium and peritoneal endometriosis: a new quantitative method. *Fertil Steril* 1994,62,751-759. 91 Lessey BA, Metzgeer DA, Haney AF, McCarty KS: Immunohistochemical analysis of estrogen and progesterone receptors in endometriosis: comparison with normal endometrium during the menstrual cycle and the effect of medical therapy. *Fertil Steril* 1989,51,409-415. 92 Bergqvist A, Ljunberg O, Skoog L: Immunohistochemical analysis of estrogen and progesterone receptors in endometriotic tissue and endometrium. *Hum Reprod* 1993,8,1915-1922.

93 Prentice A, Randall BJ, Weddell A, McGill A, Herny L, Horne CHW: Ovarian steroid receptor expression in endometriosis and in two potential parent parent epithelia: endometrium and peritoneal mesothelium. *Hum Reprod* 1992,9,1318-1325.

94 Howell RJ, Dowsett M, Edmonds DK: Oestrogen and progesterone receptors in endometriosis: heterogeneity of different sites. *Hum Reprod* 1994,9,1752-1758.

95 Attar E and Bulun SE: Aromatase and other steroidogenic genes in endometriosis: translational aspects. *Hum Reprod Update* 2006,12,49-56.

96 Bulun SE, Imir G, Utsunomiya H, Thung S, Gurates B, Tamura M, Lin Z: Aromatase in endometriosis and uterine leiomyomata. *J Steroid Biochem Mol Biol* 2005,95,57-62, 97 Nisolle M, Casanas-Rouz F, Donnez J: Immunohistochemical analysis of proliferative activity and steroid receptor expression in peritoneal and ovarian endometriosis. *Fertil Steril* 1997,68,912-919.

98 Lewis GS: Steroidal regulation of uterine immune defenses. Anim. Reprod. Sci.

2004,82-83,281-294.

99 Beagley and KW, Gockel CM: Regulation of innate and adaptive immunity by the female sex hormones estradiol and progesterone. *FEMS Immunol Med Microbiol* 2003,38,13-22.

100 Larsen S and Galask RP: Vaginal microbial flora: practical and theoretic relevance. *Obstet Gynecol* 1980,55,100S-113S.

101 Herath S, Fischer FD, Werling D, Williams EJ, Lilly ST, Dobson H, Bryant CE, Sheldon IM: Expression and Function of Toll-like receptor 4 in the endometrial cells of the uterus. *Endocrinology* 2006,147,562-570.

102 Mori T, Kobayashi H, Nishimoto H, Suzuki A, Nishimura T, Mori T: Inhibitory effect of progesterone and 20 alpha-hydroxypregn-4-en-3-one on the phytohemagglutinin-induced transformation of human lymphocytes. *Am J Obstet Gynecol* 1977,127,151-157.

103 Grossman CJ. Regulation of the immune system by sex steroids. *Endocrinol Rev.* 1984,5,435-455.

104 Lahita RG: The role of sex hormones in systemic lupus erythematosus. Curr. Opi.

Rheumatoid 1999,11,352-356.

105 Tanriverdi F, Silveira IFG< MacColl GS, Bouloux PMG: The hypothalamic-pituitary-gonadal axis: immune function and anutoimmunity. *J Endocrinol* 2003,176,293-304.

106 Giglio T, Imro MA, Filaci G, Scudeletti M, Puppo F, Cecco D, Indiveri F, Costantini S: Immune cell circulating subsets are affected y affected by gonadal function. *Life Sci*. 1994,54,1305-1312.

107 Yang JH, Chen CD, Wu MY, Chao KH, Yang YS, Ho HN: Hormone replacement therapy reverses the decrease in natural killer cytotoxicity but does not reverse the decreases in the T-cell subpopulation or interferon-gamma production in postmenopausal women. *Fertil Steril* 2000,74,261-266.

108 Kamada M, Irahara M, Maegawa M, Ohmoto Y, Takeji T, Yasui T, Aono T: Postmenopausal changes in serum cytokine levels and hormone replacement therapy. *Am J Obstet Gynecol* 2001,184,309-313.

109 Deguchi K, Kamada M, Irahara M, Maegawa M, Yamamoto S, Ohmoto Y, Murata K, Yasui T, Yamano S, Aono T: Postmenopausal changes in production of type 1 and type 2 cytokines and the effects of hormone replacement therapy. *Menopause* 2001,8,266-272.

110 Seaman WE, Blackman MA, Ginhart TD, Roubinian JR, Loeb JM, Talal N: β -estradiol reduces natural killer cells in mice. *J Immunol* 1978,121,2193-2198.

111 Hanna N: Role of natural killer cells in control of cancer metastasis. *Cancer Metastasis Rev.* 1882,1,45-64.

112 Hanna N, and Schneider M: Enhancement of tumor metastasis and suppression of natural killer cell activity by β -estradiol treatment. *J Immunol* 1883,130,974-980.

113 Shakhar K, Shakhar G, Rosenne E, Ben-Eliyahu S: Timing within the menstrual cycle, sex and the use of oral contraceptives determine adrenergic suppression of NK cell activity. *Br. J Cancer* 2002,83,1630-1636.

114 Yovel G, Shakhar K, Ben-Eliyahu S: The effect of sex, menstrual cycle, and oral contraceptives in the number and activity of natural killer cells. *Gynecol Oncol* 2001,81,254-262.

115 Albrecht AE, Hartmann BW, Scholten CH, Huber JC, Kalinowska W, Zielinski CC: Effects of estrogen replacement therapy on natural killer cell activity in postmenopausal women. *Maturitas* 1996,25,217-222. 116 Stopinska-Gluszak U, Waligora J, Grzela T, Gluszak M, Jozwiak J, Radomski R, Roszkowski PI, Malejczyk J: Effect of estrogen/progesterone hormone replacement therapy on natural killer cell cytotoxicity and immunoregulatory cytokine relese by peripheral blood mononuclear cells of postmenopausal women. *J Reprod Immunol* 2006,69,65-75.

117 Cutolo M, Villaggio B, Craviotto C, Pizzzorni C, Seriolo B, Sulli A: Sex hormones and rheumatoid arthritis. *Autoimmun Rev.* 2002,1,284-289.

118 Cutolo M, Sulli A, Capellino S, Villaggio B, Montagna P, Seriolo B, Straub RH: Sex hormones influence on the immune system: basic and clinic aspects in autoimmunity. *Lupus* 2004,13,635-638.

119 Canellada A, Blios S, Gentile T, Margni Idehu RA: In vitro modulation of protective antibody responses by estrogen, progesterone and interleukin-6. *Am J Reprod Immunol* 2002,48,334-343.

120 Whitacre CA, Reingold SC, O'Looney PA: A gender gap in autoimmunity. *Science* 1999,283,1277-1278.

121 Tariverdian N, Theoharides TC, Sindentopf F, Gutierrez G, Jeschke U, Rabinovich

GA, Blois SM, Arck PC: Neuro-endocrine-immune disequilibrium and endometriosis: an interdisciplinary approach. *Semin Immunopathol* 2007,29,193-210.

122 Khan KN, Kitajima M, Imamura T, Hiraki K, Fujishita A, Sekine I, Ishimaru T, Masuzaki H: Toll-like receptor 4 (TLR4)-mediated growth of endometriosis by human heat shock protein 70 (Hsp70). *Hum Reprod* 2008, (in press).

123 Babson AL: The DPC Cirrus IMMULITE automated immunoassay system. *J Clin Immunol* 1991,14,817-821.

124 Jansen RPS and Russell P: Non-pigmented endometriosis: clinical, laparoscopic, and pathological definition. *Am J Obstet Gynecol* 1986,155,1154-1159.

125 Martin DC, Hubert GD, Vander-Zwaag R, EL-Zeky F: Laparoscopic appearances of peritoneal endometriosis. *Fertil Steril* 1989,51,63-67.

126 Ishimaru T and Masuzaki H: Peritoneal endometriosis: endometrial tissue implantation as its primary etiologic mechanism. *Am J Obstet Gynecol* 1991,165,210-214.

127 Murphy AA, Green WR, Bobbie D, dela Cruz ZC, Rock JA: Unsuspected endometriosis documented by scanning electron microscopy. *Fertil Steril* 1986,46,522-524.

128 Nakamura M, Katabuchi H, Tohya T, Fukumatsu Y, Matsuura K, Okamura H: Scanning electron microscopic and immunohistochemical studies of pelvic endometriosis. *Hum Reprod* 1993,8,2218-2226.

129 McLaren J, Prentice A, Charnock-Jones DS, Millican SA, Muller KH, Sharkey AM, Smith SK: Vascular endothelial growth factor is produced by peritoneal fluid macrophages in endometriosis and is regulated by ovarian steroids. *J Clin Invest* 1996,98,482-489.

130 Bergqvist A: Steroid receptors in endometriosis. In Thomas E and Rock J (eds). Modern Approaches to Endometriosis. Kluwer, Dordrecht, The Netherlands 1992,257-274.

131 Eischen A, Duclos B, Schmitt-Gognel M, Rouyer N, Bergerat JP, Hummel M, OskamR, Oberling Fr: Human resident peritoneal macrophages: phenotype and histology. *Br. J.Haematol* 1994,88,712-722.

132 Khan KN, Masuzaki H, Fujishita A, Kitajima M, Hiraki K, Sekine I, Matsuyama T,Ishimaru T: Interleukin-6- and tumor necrosis factor *a*-mediated expression of

hepatocyte growth factor by stromal cells and its involvement in the growth of

endometriosis. Hum Reprod 2005,20,2715-2723.

133 Lauchlan SC: The secondary Mullerian system. Obstet Gynecol Survey 1992,27,133-146

134 Witz CA and Schenken RS: Pathogenesis. *Semin Reprod Endocrinol* 1997,15(3),199-208.

Figure Legends

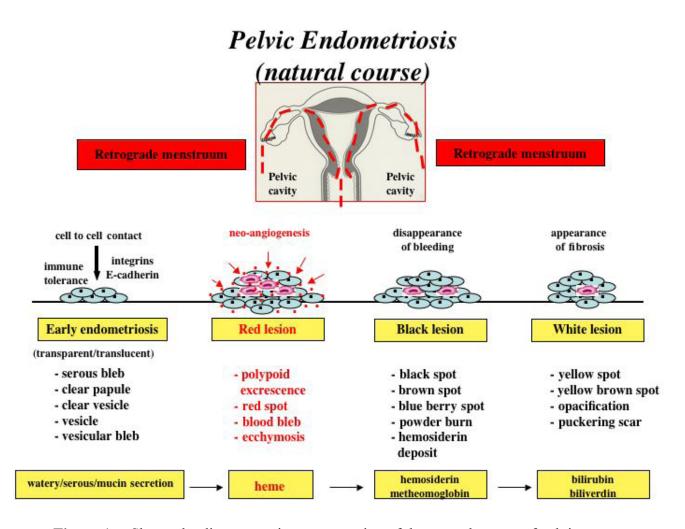


Figure 1. Shows the diagrammatic representation of the natural course of pelvic endometriosis in pelvic cavity. After initial attachment of refluxed endometrial cells with peritoneal cells producing early endometriotic lesions, the consequent events of mitosis, angiogenesis, metabolic degradation of heme and appearances of fibrosis result in the generation of different morphological appearances of pelvic endometriosis as shown in this figure.

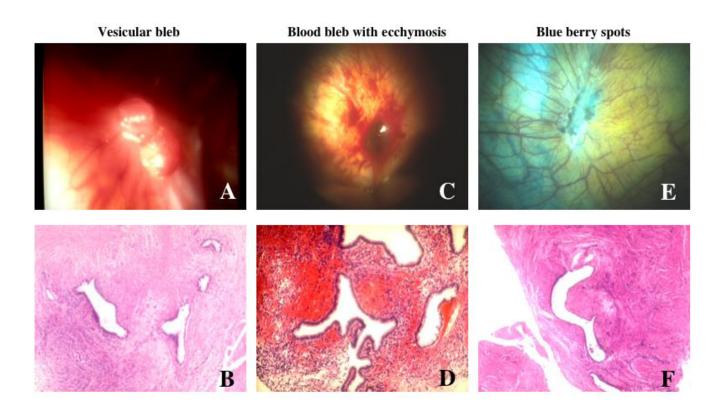


Figure 2. Shows the laparoscopic and histologic appearance of blood-filled opaque red lesions and non-opaque red lesions (A-D) and blue berry spots (E, F) in pelvic cavity. It can be noted here that non-opaque endometriotic lesions such as vesicular bleb (A) lack oozing of blood in stroma (B) and opaque red lesions such as blood bleb with ecchymosis (C) are accompanied by oozing of blood in the stroma of these lesions (D). In contrast, black lesions such as blue berry spots (E) are manifested by color change and disappearance of blood from the stroma of these lesions (F).

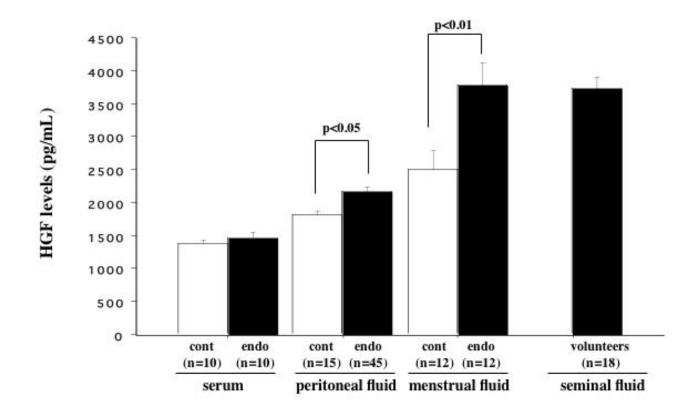


Figure 3. Shows the levels of hepatocyte growth factor (HGF) in the different body fluids of women with and without endometriosis. The level of HGF in seminal fluid was used as a positive control. Although not significant in serum, HGF levels were found to be significantly higher in the menstrual fluid (p<0.01) and peritoneal fluid (p<0.05) of women with endometriosis than that of control women (without endometriosis).

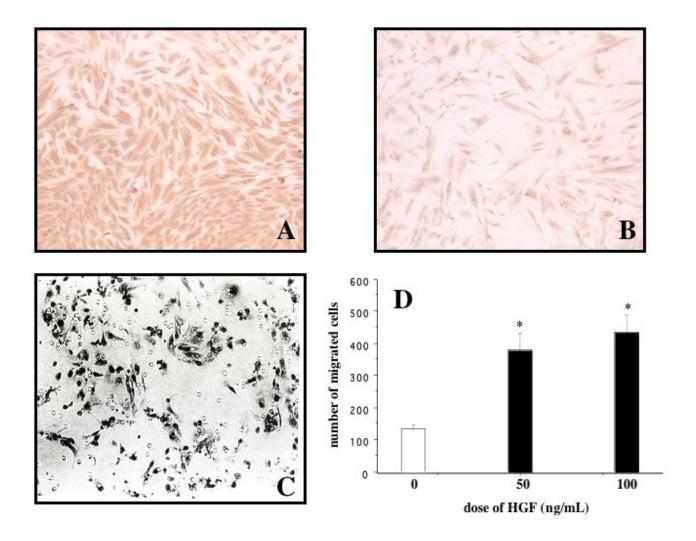


Figure 4. Shows the effect of HGF on the scattering (A, B) and migration (C, D) of endometrial cells derived from women with endometriosis. After application of 100ng/ml, HGF was found to scatter endometrial stromal cells (B) in a collagen non-coating culture plate when compared with non-treated cells (A). Boyden's chamber assay showed that addition of 50ng/ml and 100ng/ml of HGF were able to migrate epithelial cells (C) and migrated cells were significantly higher (p<0.001) when compared with HGF non-treated cells (D).

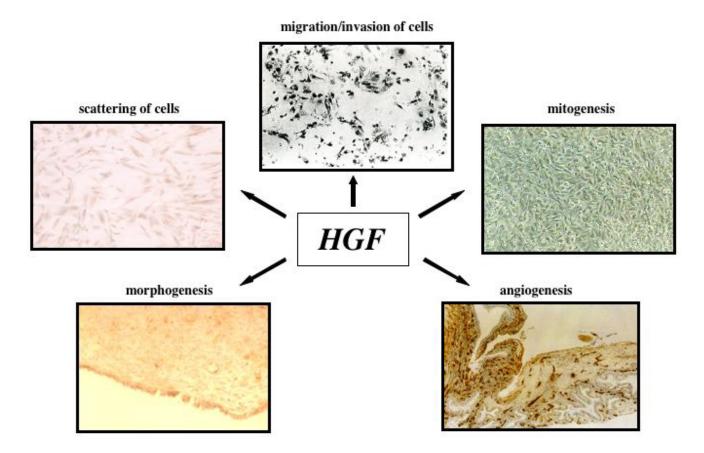
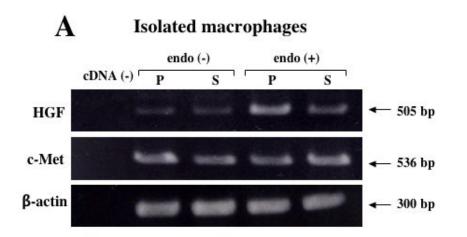
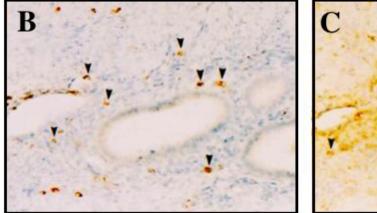


Figure 5. Shows the multifunctional role of HGF in the development of endometriosis such as migration/invasion of cells, scattering of cells, cell proliferation effect (mitogenesis) on cells of eutopic and ectopic endometrium, immunoreaction of HGF to transforming cells of pelvic peritoneum (morphogenesis) and strong immunoreaction of HGF to vascular endothelial cells (angiogenesis).



Serial section of intact tissue



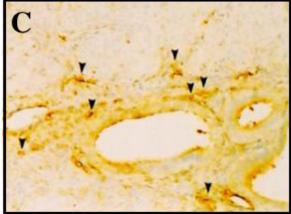


Figure 6. Shows production of HGF by macrophages. Upper panel (A) shows expression of mRNA encoding for HGF and its receptor, c-Met in isolated macrophages derived from the peritoneal fluid of women with endometriosis (endo+) and without endometriosis (endo-). The mRNA expression of HGF was found be to be higher in women with endometriosis than those without endometriosis. P, proliferative phase; S, secretory phase of menstrual cycle. Lower column (B and C) shows immunoreaction of HGF in CD68-positive macrophages in a serial section of tissue derived from the eutopic endometrium of women with endometriosis. Arrowheads show the localization

of HGF (C) in the same position as CD68-stained macrophages (B) in the stroma of

eutopic endometrium.

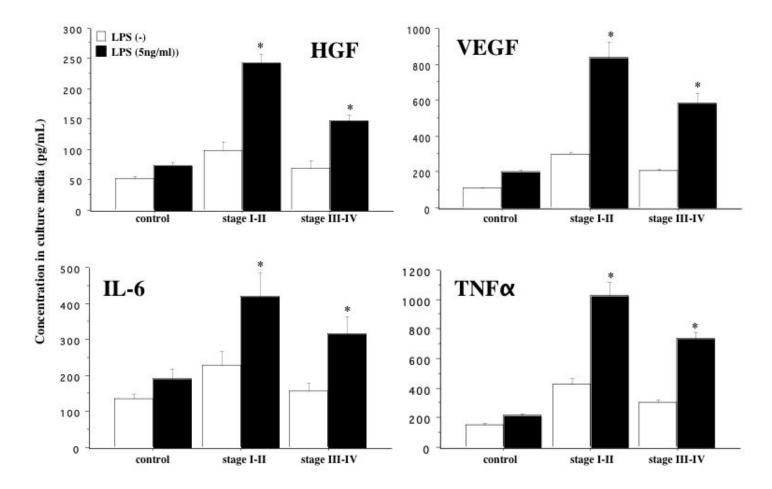


Figure 7. Shows concentrations of HGF, VEGF, IL-6 and TNF α in the culture media of basal macrophages (non-LPS-treated) and LPS-treated (5ng/ml) macrophages and according to revised-ASRM staging of endometriosis. The asterisks denote the significant difference between stage I-II and stag III-IV endometriosis. The levels of HGF, VEGF, IL-6 and TNF α in the LPS-treated culture media were significantly higher (p<0.05 for all) in women with early endometriosis (stage I-II) than that of women with advanced endometriosis (stage III-IV) or control women. Although a substantial increase was observed, basal macrophages did not show any significant difference in all macromolecules levels between these two groups of women.

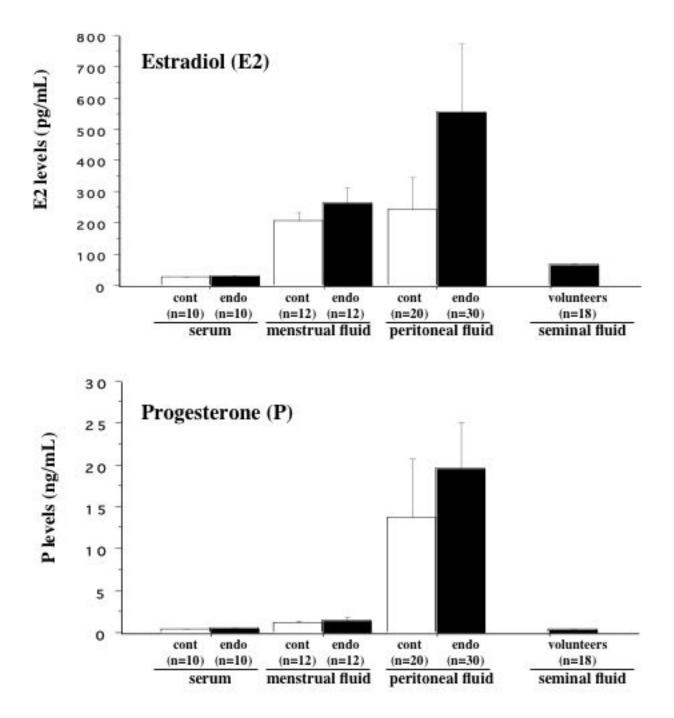


Figure 8. Shows levels of estradiol (E2) and progesterone (P) in different body fluids collected from women with and without endometriosis. The level of ovarian steroids in seminal fluid was used as a positive control. A higher increase of E2 and P in the peritoneal fluid and menstrual fluid derived from women with endometriosis was

observed when compared with corresponding levels in control women. cont, control

women; endo, women with endometriosis.

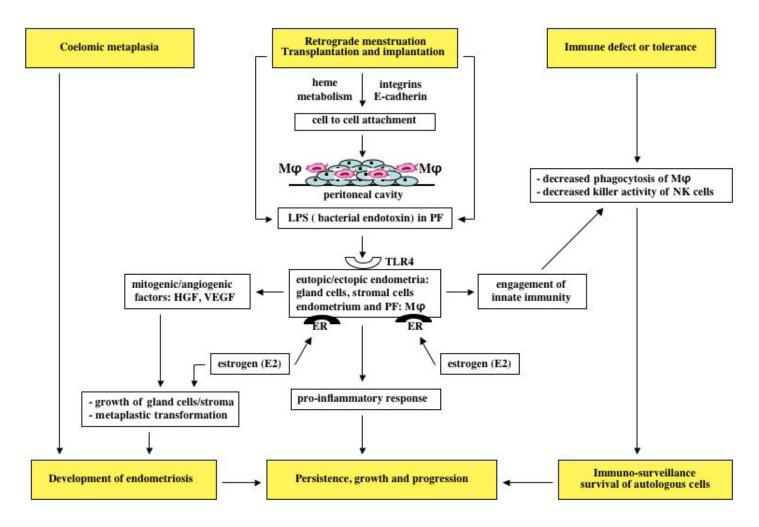


Figure 9. Shows our proposed concepts on the immunopathogenesis of pelvic

endometriosis by hepatocyte growth factor, bacterial endotoxin (LPS) via toll-like receptor 4, macrophages and ovarian steroid hormone (estrogen) and their cross-talk in the development, persistence and progression of endometriosis. The details of the development of early endometriosis by initial cell to cell contact and tissue invasion, the immuno-endocrine relationship and engagement of innate immune system in endometriosis are described in the text.