

1 **Use of sterols to monitor surface water quality change and nitrate pollution source**

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25 **Abstract**

26 Coprostanol was tested as ecological indicator to trace domestic and manure effluents and to investigate
27 possible pollution sources in surface water. Pollution assessment was performed by analysing NO_3^- , NO_2^- ,
28 coprostanol ($5\beta(\text{H})$ -Cholestan- 3β -ol), and cholesterol ($5\alpha(\text{H})$ -Cholestan- 3β -ol) in water samples from 42
29 sites along rivers in Shimabara and Unzen City, Japan. NO_2 -N concentration exceeded 0.04 mg L^{-1} at two
30 sampling sites during winter and six sampling sites during summer. $(\text{NO}_3+\text{NO}_2)$ -N concentration
31 exceeded 10 mg L^{-1} at 19 sampling sites during winter and 7 sampling sites during in summer. The
32 highest concentration was 82.4 mg L^{-1} in summer. Detectable NO_3 -N concentration was observed in
33 northern parts of the study area. Coprostanol concentration exceeded 700 ng L^{-1} (Australian Drinking
34 Water Standard) at 8 sampling points during winter and 6 sampling sites during summer. At 10 and 5% of
35 the sampling sites, both nitrate and coprostanol concentration exceeded drinking water standard during
36 winter and summer, respectively. The percentage of sampling sites where either concentration was above
37 drinking water standard was 45% during winter and 22% during summer season. However, depending on
38 sampling site, the relationships between nitrate and coprostanol concentrations showed different patterns.
39 The sterol ratio exceeded 0.5 at 17 sampling sites during winter and 14 sampling sites during summer.
40 Thus, it was confirmed that fecal pollution is present in the studied surface water. A method to distinguish

41 between principal pollution sources was developed by separating four areas in a nitrate concentration and
42 sterol ratio plot. Results show that sampled data could be reasonably classified into appropriate
43 polluted/non-polluted groups. Thus, coprostanol and sterol ratio can be used as indicators to distinguish
44 between different nitrate pollution sources in surface water.

45 **Keywords**

46 Surface water, Nitrate pollution, Coprostanol, Sterol ratio

47

48 **1. Introduction**

49 Many areas in the world experience nitrate pollution in surface and groundwater supplies (e.g.,
50 Nakagawa et al., 2016; Amano et al., 2018; Górsk et al., 2019; Chitsazan et al., 2017; Sorensen et al.,
51 2015; Hansen et al., 2012; Chandna et al., 2011; Ribbe et al., 2008; Liu et al., 2005). Nitrate pollution is
52 related to significant health threats known as cause of blue baby syndrome for infants and cancer
53 occurrence for adults. Thus, World Health Organization (WHO, 2011) has set a maximum nitrate level in
54 drinking water at 50 mg L⁻¹. Eckhardt and Stackeberg (1995) considered nitrate concentration above 13
55 mg L⁻¹ in groundwater as indicative of pollution by human activity.

56 A common problem is to establish sources for the nitrate pollution. A common method for this
57 is to use isotopes of nitrogen and oxygen in the nitrate (Kendall, 1998). However, when many sources of
58 nitrate overlap, it may still be difficult to separate between them. For this reason, we propose to use
59 coprostanol to improve the source separation (Nakagawa et al., 2017). Coprostanol is a sterol, which is
60 produced by bacterial reduction of cholesterol in gut of higher animals such as humans and livestock.

61 Coprostanol and related sterols can thus, be used as a biomarker of pollution from domestic and manure
62 effluent discharge (Reeves and Patton 2005). Coprostanol has been widely used as an indicator of fecal
63 pollution in the water environment such as river, lagoons, and estuaries (He., 2018; Costa et al., 2018;
64 Rada et al., 2016; Adnan et al., 2010; Froehner et al., 2009; Martins et al., 2007; Reeves and Patton, 2005).
65 This, has been confirmed by comparison with biological indicators such as *E. coli*. (Albuquerque de Assis
66 Costa et al., 2018). Consequently, coprostanol may be used as a proxy for ecological indicators. Although,
67 problems remain to establish relationships in different environments.

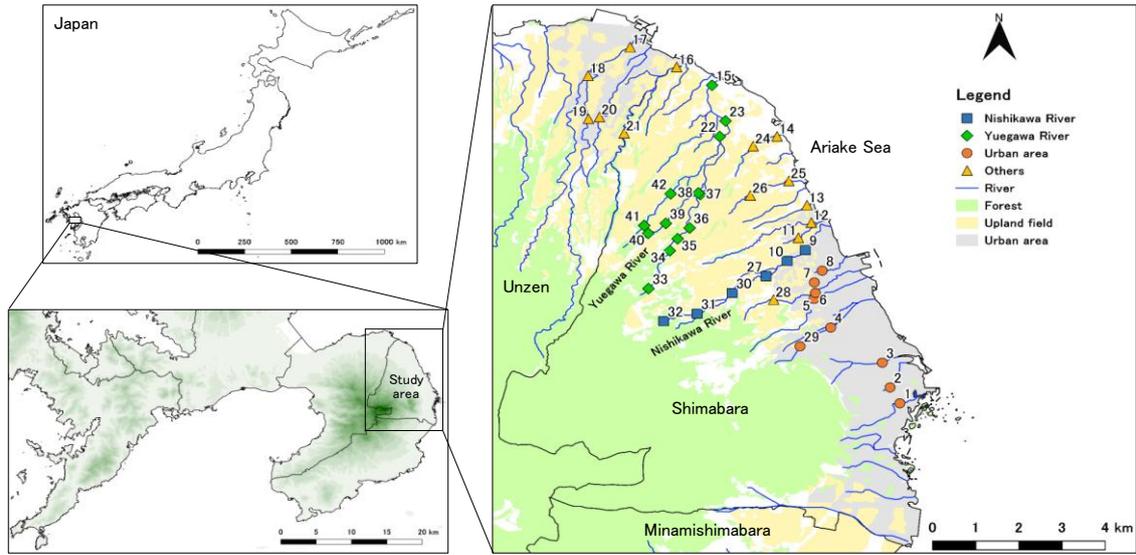
68 For groundwater, the concentration of coprostanol is often quite low. A possible reason for this
69 is adsorption to soil and rock material. Sterols are hydrophobic and thus, coprostanol may be assumed to
70 be associated with particles (Froehner et al., 2010). Thus, it may be expected that surface water displays
71 higher concentrations and more clear relationships between coprostanol and nitrate concentration.
72 However, to improve the pollution source classification, the sterol ratio between cholestanol and
73 coprostanol can be used. Cholestanol is also an isomer sterol that is formed from cholesterol reduction to
74 cholestanol. This occurs preferentially in natural environments. In view of this, we firstly investigated the
75 spatial variation of nitrate, coprostanol, and sterol ratio for the study area. Secondly, we plotted
76 relationships between $\delta^{15}\text{N}$ and $\delta^{18}\text{O}$ from nitrate as suggested by Kendall (1998). Thirdly, we analyzed
77 relationships between $\delta^{15}\text{N}$ from nitrate and sterol ratio to validate initial results. Finally, we propose a
78 general methodology to distinguish between main sources of nitrate pollution in surface water by use of
79 sterol ratio and nitrate. In other words, this study assessed the nitrate pollution in rivers and possibilities
80 to separate between different sources of pollution using several biochemical indicators.

81

82 **2. Experimental study area**

83 Shimabara is one of the cities on Shimabara Peninsula (**Fig. 1**) in Nagasaki prefecture, Japan. It
84 is adjacent to the Unzen and Minamishimabara Cities. The city area is 82.8 km², which occupies 18% of
85 the peninsula. Land use is concentrated to forest, upland fields, and urban areas. In the southern part of
86 the city, urban areas are situated on the sloping flat land between Mt. Mayuyama and the Ariake Sea. The
87 sloping flat land is constituted by debris avalanche deposits from the part of Mt. Mayuyama that collapsed
88 in 1792. The northern and central parts of the study area are constituted by agricultural areas.

89 The city is well-known for volcanic activities. The most recent eruptive activity of Mt.
90 Fugendake in the center of the peninsula occurred from 1990 to 1995. Due to the frequent volcanic
91 activities, the geology is mainly constituted by volcanic rock such as Pre-Unzen and Unzen volcanic
92 rocks (Sugimoto, 2006). Pre-Unzen volcanic rock is covered by Unzen volcanic rock. The former is
93 composed of olivine basalt and two pyroxene andesite. The latter is mainly constituted by hornblende
94 andesite to dacite.



95

96 **Fig. 1** Location of water sampling sites.

97

98 The climate is humid temperate with a mean annual temperature of 17.1°C and annual

99 precipitation of 1989 mm. The precipitation in January, February, and August for each sampling campaign

100 was 56.5, 125.5, and 196.5 mm, respectively. About 41% of the population of 46,437 people are

101 connected to waste water treatment (Nagasaki Prefecture, 2017). Most of the treatment (97.6%) is

102 constituted by septic tanks due to lacking sewage system connections in the city. It is estimated that about

103 224 kg T-N/day are released to the river system through discharge of treated waste water (treated septic

104 tank effluent). Nitrate load to the groundwater from treated waste water is, however, only about 5% of the

105 load from fertilizers and livestock waste (Committee on Nitrate Reduction on Shimabara Peninsula,

106 2016).

107

108 **3. Materials and methods**

109 **3.1. Water samples**

110 Water samples were taken at 42 locations in Shimabara City including a part of Unzen City
111 during winter (January 17 and 24 and February 6) and summer (August 21 and 22) 2017. However,
112 during the summer campaign, sampling at site 27 (**Fig. 1**) was not possible due to dried up river
113 conditions. The reason for sampling during both summer and winter was to better understand the seasonal
114 variation of the biochemical indicators. Water samples of 500 mL and 1000 mL were collected directly
115 from the center section of the river (except for sampling sites 6 and 27) in pre-washed bottles and then
116 stored in refrigerator for analysis of NO_3^- , NO_2^- , NH_4^+ , coprostanol, and cholestanol. At sites 6 and 27, we
117 used a bailer sampler due to the difficulty to access the water surface. Samples of 30 mL for nitrate
118 isotope ratios were filtered through 0.22 μm membrane filter and kept frozen until analysis.

119

120 **3.2. Analysis procedure**

121 Dissolved oxygen (DO) was determined in situ by use of a luminescence based sensor (HACH
122 HQ30d). NO_3^- , NO_2^- , and NH_4^+ were analyzed by ion chromatography (Metrohm 861 Advanced Compact
123 IC). Nitrate $\delta^{15}\text{N}$ and $\delta^{18}\text{O}$ were determined by the denitrifier method (Casciotti et al., 2002; Sigman et al.,
124 2001; Hosono et al., 2014; Hosono et al., 2013). Analysis of $\delta^{15}\text{N}$ and $\delta^{18}\text{O}$ of induced N_2O was
125 conducted simultaneously. The nitrate isotopes were analyzed in deionized water extracted samples from
126 the cattle manure and excrement of milk and beef cattle. Cattle manure is composed by cattle excrements
127 and sawdust. The cattle manure comes from both milk cattle (cow milking raw milk for dairy products)
128 and beef cattle (cow raised for meat production). Animal waste was collected at Nagasaki Agricultural

129 and Forestry Technical Development Center on September 17 2015. In the extraction, 30 mL of deionized
130 water was added to 3.0 g of dried samples (manure and excrement) in a centrifuge tube and shaken during
131 30 min. The extract was separated using centrifugal separation at 3000 rpm x 10 min. Supernatant liquid
132 was added to a 50 ml volumetric flask through a filter paper. For further details, see Oyanagi et al. (2004).

133 As mentioned earlier, we focused on two sterols: coprostanol and cholestenol. These sterols are
134 isomer and originate from different processes. Coprostanol is produced by bacterial reduction of
135 cholesterol in gut of higher animals (Martins et al., 2007). On the other hand, cholesterol reduction to
136 cholestanol occurs preferentially in natural environment. Coprostanol ($5\beta(\text{H})\text{-Cholestan-3}\beta\text{-ol}$) and
137 cholestenol ($5\alpha(\text{H})\text{-Cholestan-3}\beta\text{-ol}$) were extracted from the water samples according to Hussain et al.
138 (2010) with some modification (Nakagawa et al., 2017). At start, a surrogate was added to 800 mL of
139 water samples to monitor the performance of preprocessing. 1 M HCl was added to samples to modify pH
140 from 2 to 3. After suction filtration of water samples using 0.7 mm and 2.7 mm borosilicate glass fiber
141 filters, the sterols on the filters were extracted with methanol using an ultrasonic bath during three
142 successive times. The extracts with methanol were mixed with the water sample after filtration with 0.20
143 μm membrane filter. The sterols were extracted from water samples by liquid-liquid extraction with 60
144 mL dichloromethane in room temperature during three successive times. The extracts were concentrated
145 to near dryness (<1.0 mL) under pure nitrogen gas flow and dehydrated with anhydrous sodium sulfate.
146 The extract was formed to trimethylsilyl using BSTFA (bis-trimethylsilyl trifluoroacetamide) at 80°C for
147 60 min after concentration and dehydration, then quantified by use of 7000A Triple Quadrupole GC/MS
148 (Agilent Technologies). The detection limit for sterols ranged between 0.2 and 3.2 ng L⁻¹. The mean

149 recovery of surrogates was 85.9% with a standard deviation $\pm 15.3\%$.

150

151 **3.3. Spatial variation of nitrate, coprostanol, and sterol ratio**

152 To accomplish the first aim of the study, to improve the understanding of the spatial variation
153 of nitrate, coprostanol, and sterol ratio, geographical information system (GIS) was applied. Thus, QGIS
154 2.18.21 'Las Palmas' (QGIS development team 2019) was used to map the results. The QGIS is a Free
155 and Open Source Geographic Information System licensed under the GNU General Public License.

156 Maximum permissible nitrate concentration in Japan for drinking water is 10 mg L^{-1} . Therefore,
157 this concentration was used as a criteria for nitrate pollution. To evaluate fecal pollution, several
158 coprostanol concentrations have been proposed. In Australia, a maximum of 700 ng L^{-1} has been proposed
159 for drinking water (Hussain et al., 2010). In the natural water bodies, according to the relationship
160 between BOD and coprostanol concentration, 500 ng L^{-1} has been proposed for indicating fecal pollution
161 (Itoh and Tatsukawa, 1978). In this study, we evaluated fecal pollution based on drinking water standard
162 (proposed in Australia) of coprostanol concentration as same as nitrate concentration.

163 Another method to evaluate fecal pollution is to use the sterol ratio (Matić et al., 2016):

$$164 \quad S_r = \frac{5\beta}{5\alpha + 5\beta}$$

165 where S_r is sterol ratio, 5β is concentration of coprostanol (ng L^{-1}), and 5α is concentration of cholestanol
166 (ng L^{-1}). Depending on sterol ratio, samples are often divided into three pollution classes "certain (> 0.5)",
167 "uncertain ($0.3 - 0.5$)", and "no pollution (< 0.3)" (Matić et al., 2016). Consequently, these thresholds
168 were used to indicate fecal pollution in the current study.

169

170 **3.4. Relationship between nitrate $\delta^{15}\text{N}$ and $\delta^{18}\text{O}$**

171 Kendall (1998) suggested a method to investigate pollution sources that can be used together
172 with groundwater sampling (Nakagawa et al., 2017). It builds on a scatter plot using $\delta^{15}\text{N}$ and $\delta^{18}\text{O}$
173 concentrations. The scatter plot will display distinctive ranges depending on nitrate source (e.g.,
174 ammonium fertilizer, soil N, and manure and septic waste). Thus, several studies have used this method to
175 elucidate nitrate pollution source (Baily et al., 2011; Hosono et al., 2011; Kaown et al., 2009).

176

177 **3.5. Relationship between nitrate $\delta^{15}\text{N}$ and sterol ratio**

178 The relationship between nitrate $\delta^{15}\text{N}$ and sterol ratio is plotted in a scatter diagram. The nitrate
179 $\delta^{15}\text{N}$ indicates the nitrate source while the sterol ratio displays the risk for fecal pollution. Nitrate
180 originating from manure or septic waste would have a heavier isotopes range compared to ammonium
181 fertilizer and nitrate from soil with a high sterol ratio (> 0.5).

182

183 **3.6. Relationship between $\text{NO}_3+\text{NO}_2\text{-N}$ and sterol ratio**

184 We suggest a general methodology to distinguish between main pollution sources of nitrate by
185 using a scatter plot between $\text{NO}_3+\text{NO}_2\text{-N}$ (x-axis) and sterol ratio (y-axis). In such a plot, four main
186 groups may be distinguished. These groups correspond to drinking water standard regarding $\text{NO}_3+\text{NO}_2\text{-N}$
187 and fecal pollution criteria (group I, $\text{NO}_3+\text{NO}_2\text{-N}$: $< 10 \text{ mg L}^{-1}$, sterol ratio: < 0.3 ; group II, $\text{NO}_3+\text{NO}_2\text{-N}$:
188 $< 10 \text{ mg L}^{-1}$, sterol ratio: > 0.3 ; group III, $\text{NO}_3+\text{NO}_2\text{-N}$: $> 10 \text{ mg L}^{-1}$, sterol ratio: < 0.3 ; group IV,

189 $\text{NO}_3+\text{NO}_2\text{-N} > 10 \text{ mg L}^{-1}$, sterol ratio: > 0.3). Group I indicates no pollution or close to no pollution
 190 while other groups indicate pollution. However, pollution sources might be different, as domestic
 191 wastewater in group II, chemical fertilizer in group III, and livestock waste in group IV.

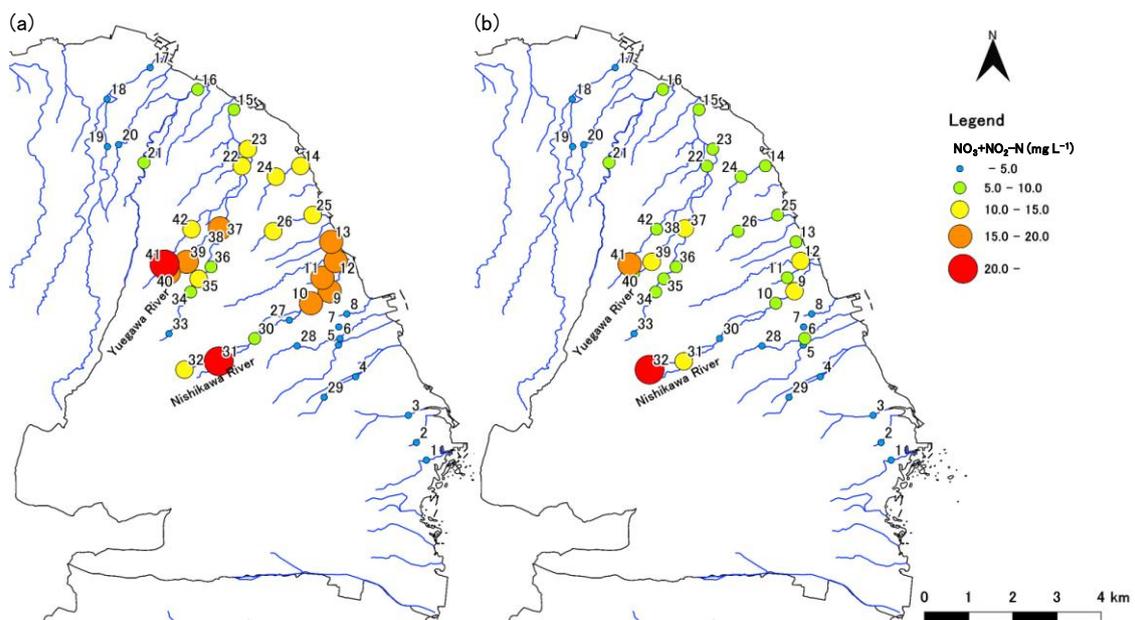
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193 4. Results

194 4.1. Spatial variation of nitrate, coprostanol, and sterol ratio

195 The spatial variation of nitrate plus nitrite as nitrogen ($\text{NO}_3+\text{NO}_2\text{-N}$, hereafter referred to as
 196 nitrate) during winter and summer 2017 is shown in **Fig. 2**. Descriptive statistics of chemical components
 197 are summarized in **Table 1**. Nitrate concentration exceeded 10 mg L^{-1} at 19 sampling locations during
 198 winter and 7 sampling locations during summer. Highest concentrations were 27.5 mg L^{-1} (site 31) in
 199 winter and 82.4 mg L^{-1} (site 32) in summer. High nitrate concentration was observed in northern parts of
 200 the study area, especially for Yuegawa and Nishikawa River.

201



202 **Fig. 2** Spatial variation of $\text{NO}_3+\text{NO}_2\text{-N}$ concentration (a) Winter (Jan.-Feb. 2017), (b) Summer

203 (August 2017).

204 **Table 1** Descriptive statistics of analysed chemical components

| | | winter season | | | | summer season | | | |
|-------------------------------------|--------------------|---------------|---------|---------|---------|---------------|-------|---------|---------|
| | | Average | SD | Maximum | Minimum | Average | SD | Maximum | Minimum |
| NO ₃ -N | mg L ⁻¹ | 9.7 | 6.9 | 27.5 | 1.0 | 7.9 | 11.3 | 74.8 | 1.3 |
| NO ₂ -N | mg L ⁻¹ | 0.2 | 0.7 | 3.5 | 0.0 | 0.2 | 1.2 | 7.6 | 0.0 |
| NO ₃ +NO ₂ -N | mg L ⁻¹ | 9.9 | 6.8 | 27.5 | 1.0 | 7.9 | 12.4 | 82.4 | 1.3 |
| NH ₄ -N | mg L ⁻¹ | 2.1 | 7.3 | 37.0 | 0.0 | 0.0 | 0.1 | 0.6 | 0.0 |
| Coprostanol | ng L ⁻¹ | 2064.7 | 10509.2 | 68340.3 | 17.9 | 330.9 | 438.7 | 1826.1 | 2.5 |
| Cholestanol | ng L ⁻¹ | 883.3 | 3267.4 | 21196.6 | 30.6 | 299.0 | 346.2 | 1654.7 | 37.5 |
| Sterol ratio | | 0.47 | 0.17 | 0.77 | 0.14 | 0.41 | 0.20 | 0.76 | 0.06 |
| DO | mg L ⁻¹ | 10.14 | 2.49 | 11.58 | 0.70 | 8.08 | 0.85 | 10.42 | 5.26 |

205

206

207 Nitrite (NO₂-N) was detected at only two locations during winter (site 30 and 32). For summer,
208 nitrite was detected at six locations (site 1, 4, 7, 8, 11, and 32). Although nitrate concentrations meet
209 drinking water standard (10 mg L⁻¹) except for site 32 (**Fig. 2(b)**), nitrite exceeds Japanese drinking water
210 standard at these sites (0.04 mg L⁻¹). Very high nitrite and nitrate concentration (7.6 mg L⁻¹ and 82.4 mg
211 L⁻¹, respectively) was detected at site 32 in summer.

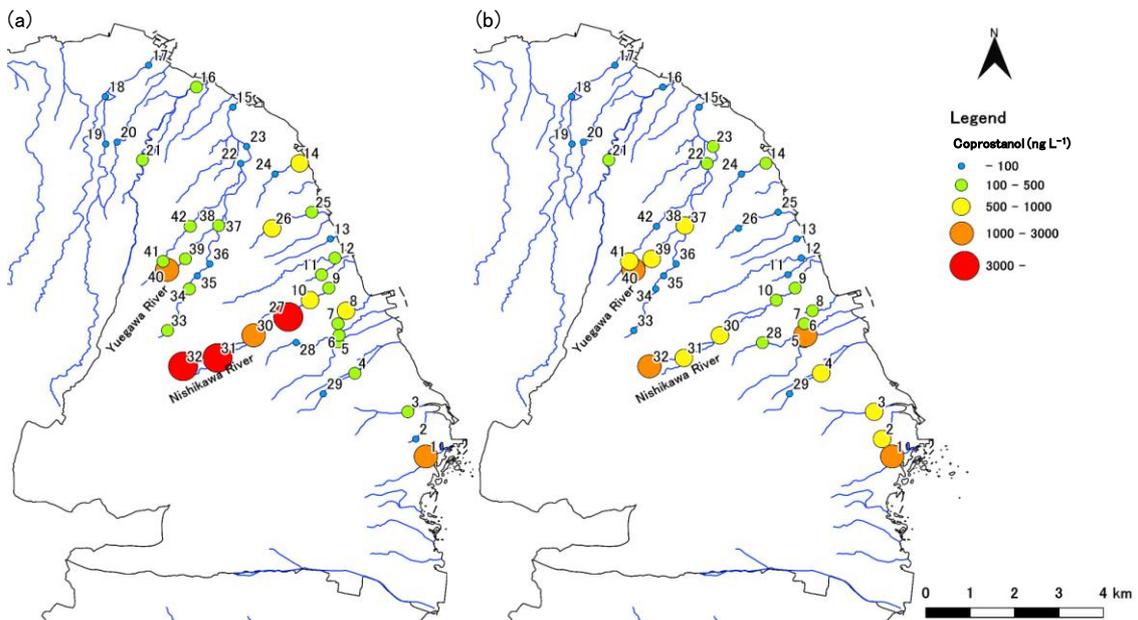
212 Ammonium (NH₄-N) was detected in water samples collected at locations 26, 27, 31 and 32
213 (winter season) and 30 (winter and summer season). The ammonium concentrations ranged between <9.5
214 to 37.0 mg L⁻¹ for most of the sampling locations, within Nishikawa River Basin (locations 27 and 30-32)
215 and <1.8 mg L⁻¹ for location 26 in winter. The maximum concentration (37.0 mg L⁻¹) was detected at site
216 32 in winter. In summer (location 30), ammonium was detected with concentration of 0.6 mg L⁻¹, which is
217 significantly lower than in winter season.

218 The spatial variation of coprostanol concentration in winter and summer 2017 is shown in **Fig.**

219 3. A total number of 8 and 6 sampling points exceeded the Australian standard of 700 ng L⁻¹ during winter
220 and summer, respectively. Site 32 (upstream Nishikawa River) displayed the highest coprostanol
221 concentration, 68340 and 1247 ng L⁻¹ during winter and summer, respectively. The Nishikawa River is
222 highly affected by fecal pollution from livestock waste. Similarly, as for the nitrate concentration,
223 coprostanol concentration decreased during summer. However, some sites like 2, 3, 4, and 6 (urban area)
224 and 22, 23, 37, 39, and 41 (Yuegawa River) displayed increasing coprostanol concentration during
225 summer.

226 According to the above, sterol ratios > 0.5 represent “certain” fecal pollution. Values between
227 0.3 and 0.5 suggest “uncertain” sewage and natural sterol inputs. Values < 0.3 indicate “no pollution”.
228 Spatial variation of sterol ratio during winter and summer in 2017 is shown in **Fig. 4**. Based on these
229 criteria, 17 sampling points in winter and 14 sampling points in summer exceeded 0.5, indicating certain
230 fecal pollution. In total, 10 sampling points in winter and 15 sampling points in summer showed a ratio of
231 less than 0.3.

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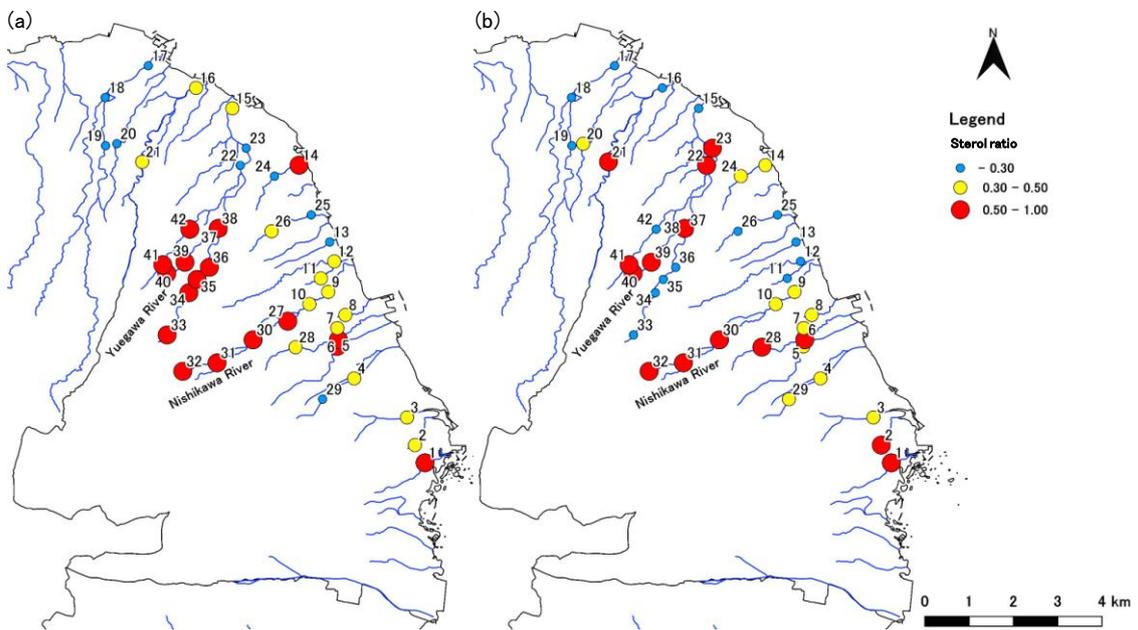


233

234 **Fig. 3** Spatial variation of coprostanol concentration (a) Winter (Jan.-Feb. 2017), (b) Summer (August

235

2017).



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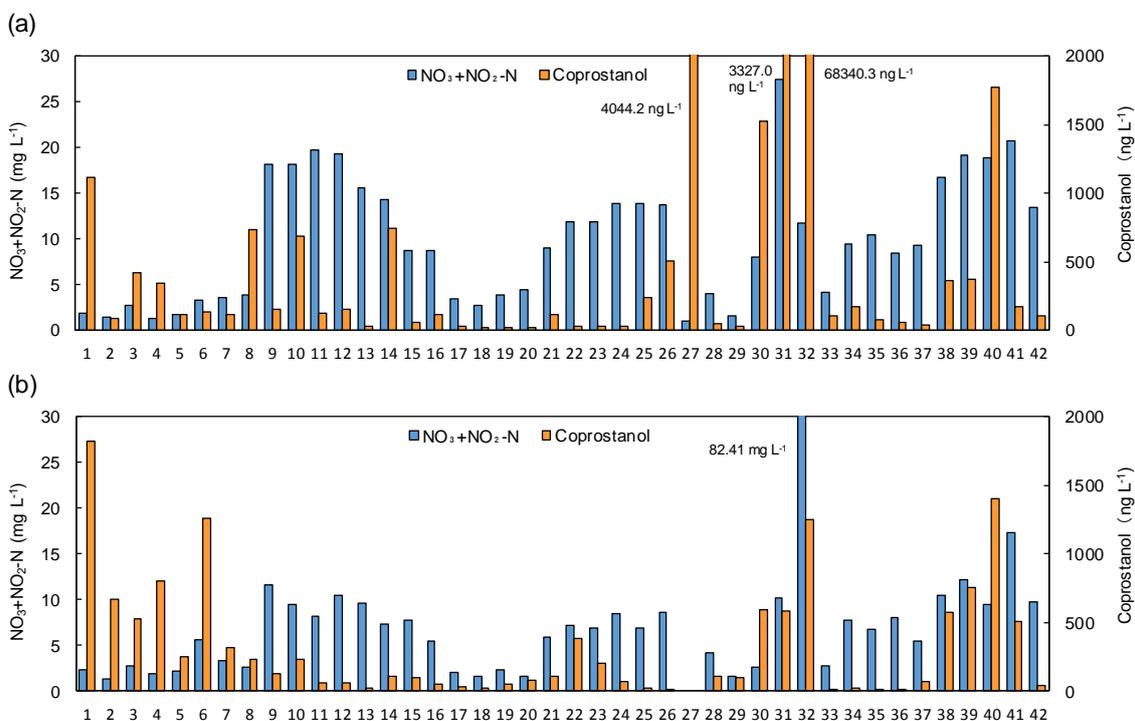
237 **Fig. 4** Spatial variation of sterol ratio (a) Winter (Jan.-Feb. 2017), (b) Summer (August 2017).

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239

Concentrations of $\text{NO}_3+\text{NO}_2\text{-N}$ and coprostanol are shown in **Fig. 5** to give a direct comparison

240 of concentrations of these components. 19 sampling sites displayed higher nitrate concentration than
 241 drinking water standard during winter season. Within these sites, 14, 31, 32, and 40 showed high
 242 coprostanol concentration, above the Australian drinking water standard. Thus, 10 % of all sampling sites
 243 showed high concentrations for winter season. The percentage that displayed high concentration for
 244 nitrate or coprostanol was 45 % for winter season. During summer season, both nitrate and/or coprostanol
 245 concentration decreased at sites 14, 31, and 40, and only site 32 displayed high concentrations for both
 246 chemicals during both seasons. In addition, site 39, where only nitrate was high during winter season,
 247 showed high concentrations of nitrate and coprostanol during summer season. The percentage of sites
 248 where both chemical concentrations were high was only 5 % for summer season. The percentage of sites
 249 sampling sites where either concentration was above drinking water standard was 45% during winter and
 250 22% during summer season.



251

252 **Fig. 5** $\text{NO}_3+\text{NO}_2\text{-N}$ and coprostanol concentration depending on sampling site (a) Winter (Jan.-Feb.
253 2017), (b) Summer (Aug. 2017).

254

255 **4.2. Relationship between nitrate $\delta^{15}\text{N}$ and $\delta^{18}\text{O}$**

256 The average concentration of nitrate $\delta^{15}\text{N}$ and $\delta^{18}\text{O}$ was plotted in a scatter diagram as shown in

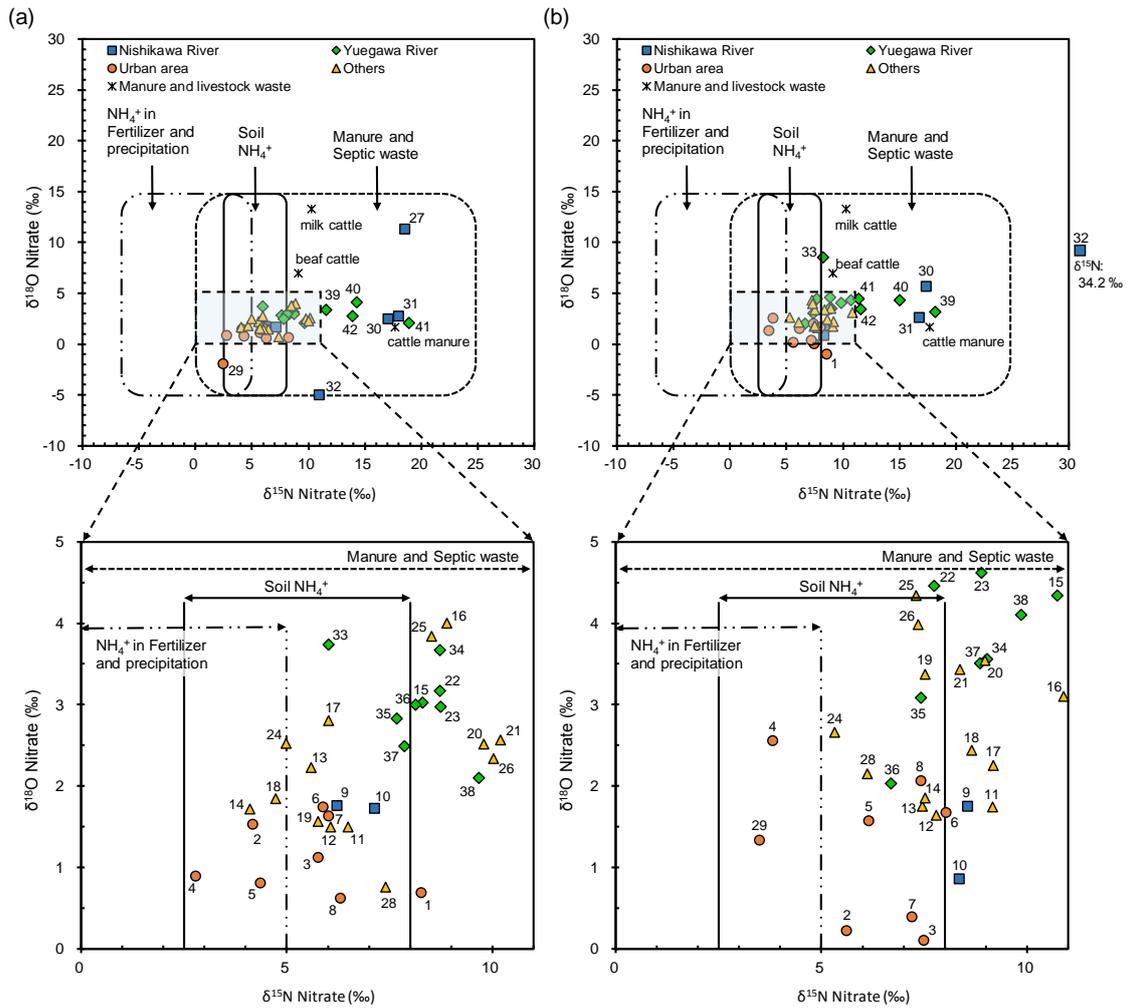
257 **Fig. 6.** Corresponding analyses from livestock waste and manure are also plotted in the figure. From

258 winter to summer, most of the samples somewhat moved towards higher concentrations for both isotopes.

259 It is seen that summer samples are more affected by livestock waste. In other words, winter samples seem

260 to be more affected by nitrate from chemical fertilizers. Yet, most of the samples and especially samples

261 from Nishikawa and Yuegawa River are located in the vicinity of manure and septic waste (**Fig. 6**).



262

263 **Fig. 6** Relationship between nitrate $\delta^{15}\text{N}$ and $\delta^{18}\text{O}$ concentrations (a) Winter (Jan.-Feb.2017), (b)

264

Summer (August 2017). The isotopic range identifying the source was organized according to

265

Kendall (1998).

266

267 4.3. Relationship between nitrate $\delta^{15}\text{N}$ and sterol ratio

268

In order to confirm the increasing tendency for both nitrate $\delta^{15}\text{N}$ and sterol ratio, the

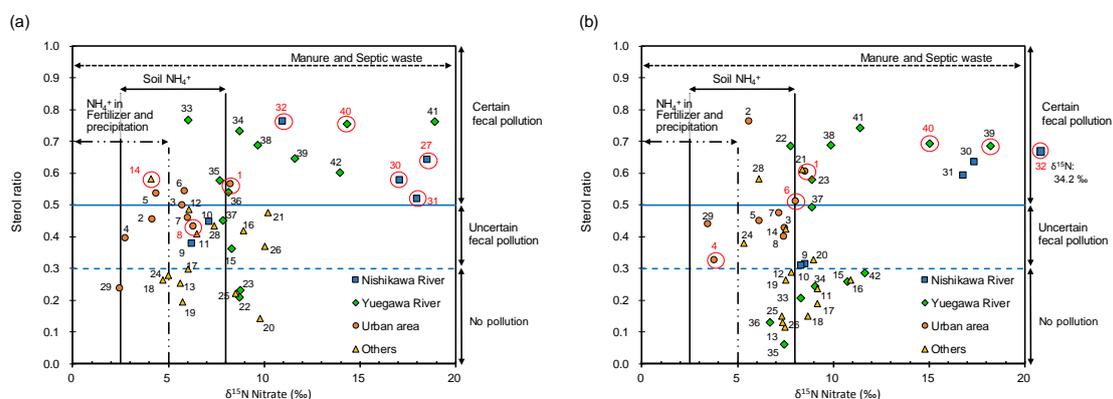
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relationship between them was plotted in **Fig. 7**. Symbols that are encircled with a red line indicate that

270

coprostanol concentration exceeded 700 ng L^{-1} . In the case of Nishikawa River, at upstream site 30, 31,

271 32, and 27 (winter only), the sterol ratio was higher than 0.5 and nitrate $\delta^{15}\text{N}$ was higher than 10‰. On
 272 the other hand, downstream sites 9 and 10 were below 0.5 for the sterol ratio and below 10‰ for the
 273 nitrate $\delta^{15}\text{N}$. In the case of Yuegawa River, most of the sampling sites, except for 22, 23 (winter) and
 274 33-36 (summer), displayed a relatively high sterol ratio (> 0.3) and nitrate $\delta^{15}\text{N}$ (> 5 ‰). Also, the urban
 275 area displayed an increasing tendency for both nitrate $\delta^{15}\text{N}$ and sterol ratio.



276

277 **Fig. 7** Relationship between nitrate $\delta^{15}\text{N}$ and sterol ratio (a) Winter (Jan.-Feb. 2017), (b) Summer
 278 (August 2017).

279

280 4.4. Relationship between $\text{NO}_3+\text{NO}_2\text{-N}$ and sterol ratio

281

The relationship between nitrate concentration ($\text{NO}_3+\text{NO}_2\text{-N}$) and sterol ratio is plotted in **Fig.**

282

8. According to the above, a methodology to distinguish principal pollution sources by separating four

283

principal fields in the plot is proposed. For example, the first field with (group I) indicates small

284

amounts of pollution because nitrate concentration meets drinking water standard and sterol ratio is

285

below 0.3. Other groups are classified as polluted, but the pollution source may be different. Coprostanol

286

concentration of samples marked in red exceeds 700 ng L^{-1} . According to this classification, most samples

287 display reduced nitrate concentration from winter to summer, and tend to end up in group I or II. Sites

288 located in the upstream of Yuegawa and Nishikawa River such as 31, 32, 38, 39, 40, and 41 are classified

289 into group IV both for winter and summer. All samples from urban areas were classified into group I or II

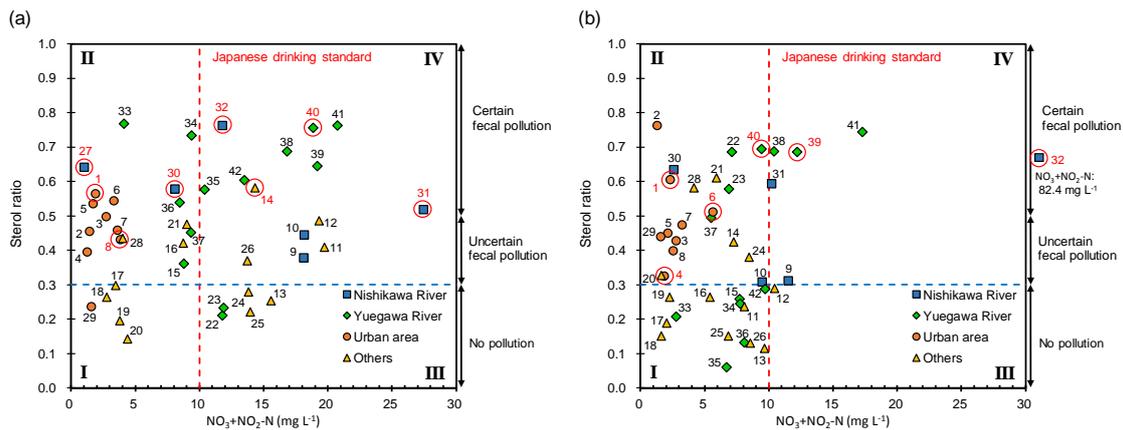
290 with lower nitrate concentration in both seasons. As shown in **Fig. 3**, nitrate concentrations in urban areas

291 were less than that of other areas, which is obvious in the winter season. Group III obtained a small

292 number of samples for both seasons. In winter season, only five samples (site 13, 22, 23, 24, and 25) were

293 classified into group III. Only site 12 belonged to group III for the summer season.

294



295

296 **Fig. 8** Relationship between $\text{NO}_3+\text{NO}_2\text{-N}$ concentration and sterol ratio (a) Winter (Jan.- Feb. 2017),

297 (b) Summer (August 2017).

298

299 5. Discussion

300 5.1. Spatial variation of nitrate, coprostanol, and sterol ratio

301 The Yuegawa and Nishikawa Rivers are part of an area with intense agriculture. In general,

302 spatial variation of nitrate in surface water displayed similarity to that of the groundwater (see Nakagawa

303 et al., 2016). Surface and groundwater were generally not polluted in forest and urban areas. Most of the
304 summer samples, except for sites 6 and 32, displayed smaller concentration as compared to samples from
305 the winter period. A reason for this is dilution of river water by precipitation during the rainy season.

306 Site 32 is located at the upstream of Nishikawa River. Here, a pig farm is located close to the
307 sampling site. In winter, although nitrate concentration met drinking water standard ($8.61 < 10 \text{ mg L}^{-1}$) at
308 this location, nitrite greatly exceeded the standard ($3.19 > 0.04 \text{ mg L}^{-1}$). Probably, nitrite was produced by
309 denitrification during the biological treatment process for wastewater, because low DO (1.51 mg L^{-1}) was
310 observed as nitrification bacteria need more oxygen as compared to heterotrophic bacteria. Ammonium
311 was detected at sites 27, 30, 31, and 32. These sites are located at the upstream of Nishikawa River and
312 probably affected by effluents from the pig farm.

313 A livestock waste water treatment plant is located upstream of the Yuegawa River. Probably,
314 the effluent of this plant affects pollution levels in the river. High coprostanol concentrations were
315 detected at site 1 in the urban area and this is most probably due to domestic waste water. The coprostanol
316 level is usually correlated with occurrence of *E. coli*. According to this, a coprostanol concentration of 60
317 ng L^{-1} has been proposed for as threshold of fluvial water quality (Albuquerque de Assis Costa et al.,
318 2018). In this sense, most of the investigated sites in this study relate to a significant health threat.
319 Measurement of coprostanol concentration makes it possible to detect serious river pollution, even if
320 nitrate concentrations are not high. Thus, observations of coprostanol concentration have an important
321 role to fill in terms of pollution source investigations.

322 The sterol ratio indicates that relatively many sampling points are affected by fecal pollution in

323 the study area. In summer, sites 2, 20, 21, 22, 23, 28, and 29 showed increased sterol ratio. In the urban
324 area (site 2 and 29), domestic waste water caused increased ratio during summer similar to the
325 coprostanol concentration. Downstream Yuegawa River (site 22 and 23) as well displayed increased ratio.
326 On the contrary, southern upstream area of Yuegawa River (site 33, 34, 35, and 36) displayed a decreased
327 ratio. This is probably due to the fact that fecal pollutants are transported from upstream to downstream
328 along the river during flood season. With sterol ratio of 0.7, the coprostanol concentration even higher
329 than 700 ng L⁻¹ can be classified as “uncertain” or close to no pollution indicators. Consequently, as
330 described by Matić et al. (2016), the ratio of 0.5 should be more secure for fecal pollution detection.

331

332 **5.2. Relationships between nitrate $\delta^{15}\text{N}$ and $\delta^{18}\text{O}$**

333 According to documented cultivation procedures of major crops in the study area (Amano et al.,
334 2016), fertilizers are applied on seven out of nine crops from September to January. This is an essential
335 reason why winter samples displayed less isotopic content of $\delta^{15}\text{N}$. However, the general tendency for
336 both plots was a small difference between winter and summer. Thus, it appears that it is difficult to
337 separate between different pollution sources using these isotopes. The same difficulty has been
338 experienced for groundwater samples (Nakagawa et al., 2017).

339 The results for Nishikawa and Yuegawa Rivers indicate that samples are clearly affected by
340 livestock waste according to the isotopic analysis using the Kendall (1998) plot. Additionally, three kinds
341 of isotopic observations from livestock waste and manure correspond to samples from Nishikawa and
342 Yuegawa Rivers. These facts are consistent with the assumption that the main nitrate source in the study

343 area is the livestock waste. A large number of livestock is raised in this area (Nakagawa et al., 2015). It is
344 estimated that the potential nitrate load from livestock waste is much higher as compared to that from
345 chemical fertilizers.

346

347 **5.3. Relationship between nitrate $\delta^{15}\text{N}$ and sterol ratio**

348 According to the above discussion, both Yuegawa and Nishikawa Rivers are affected by livestock
349 waste. In these rivers, results for both nitrate $\delta^{15}\text{N}$ and sterol ratio are quite consistent with fecal pollution.
350 For the urban areas, the results indicate pollution originating domestic sewage water.

351

352 **5.4. Relationship between $\text{NO}_3+\text{NO}_2\text{-N}$ and sterol ratio**

353 As mentioned above, group I does not indicate pollution. In group II, although nitrate
354 concentration meets the drinking water standard, sterol ratio is larger than 0.3. Thus, this group is
355 considered to be polluted mainly by livestock waste or domestic waste water discharge with a low T-N.
356 Site 1-8 and 29 are located in the urban area, indicating that some of these are affected by domestic waste
357 water discharge. In group III, although nitrate concentration exceeded drinking water standard, sterol ratio
358 is below 0.3. In this case, chemical fertilizers are considered to be the main source of pollution. As
359 mentioned above, fertilizer application on main crops in the study area is conducted from September to
360 January. This is one of the reason that larger number of samples were classified into group III for the
361 winter season. Finally, for group IV, samples exceeded both criteria and livestock waste is considered to
362 be a main source of nitrate pollution. Some samples from the Yuegawa and Nishikawa River indicate that

363 group IV samples are affected by livestock waste. This is confirmed by the close location of livestock
364 waste water treatment plant and pig farm.

365

366 **Conclusions**

367 In this study, we examined nitrate pollution in two rivers and possibilities to distinguish
368 between different sources of pollution by the use of several biochemical indicators. As a case study, and
369 to clarify the surface water pollution in the Shimabara study area, we firstly investigated the distribution
370 of nitrate, coprostanol, and sterol ratio as biochemical indicators. Secondly, we plotted the relationship
371 between nitrate $\delta^{15}\text{N}$ and $\delta^{18}\text{O}$ according to Kendall (1998) to elucidate the general tendency of the
372 samples. Then, we plotted the relationship between nitrate $\delta^{15}\text{N}$ and sterol ratio to confirm identified
373 pollution sources. Finally, we proposed a methodology to indicate the main sources of pollution in the
374 surface water of the study area. As a conclusion, we can summarize major results of our analysis
375 according to the following: 1) Nitrite ($\text{NO}_2\text{-N}$) concentration exceeded 0.04 mg L^{-1} at 2 sampling
376 locations during winter and 6 sampling points during summer. 2) Nitrate ($\text{NO}_3+\text{NO}_2\text{-N}$) concentration
377 exceeded 10 mg L^{-1} at 19 sampling points during winter and 7 sampling points during summer. The
378 highest concentration was 82.4 mg L^{-1} in summer. 3). Coprostanol concentration exceeded 700 ng L^{-1} at 8
379 sampling points in winter and 6 sampling points in summer. Thus, the study confirmed that fecal pollution
380 of the water occurs. 4) The sterol ratio exceeded 0.5 at 17 sampling points in winter and 14 sampling
381 points in summer. According to the above analysis, coprostanol and sterol ratio have a potential to be used
382 to distinguish between different nitrate pollution sources in surface water.

383 To summarize this study, we showed a potential use of coprostanol and sterol ratio to evaluate
384 pollution sources as biochemical indicators. Also, we proposed a concrete methodology to separate
385 between pollution sources based on these biochemical indicators. Additionally, the use of these indicators,
386 can not only separate between source of nitrate pollution but also display hidden health threats caused by
387 fecal pollution.

388

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391

392 **References**

393 Adnan, N.H., Zakaria, M.P., Juahir, H., Ali M.M. 2012. Faecal sterols as sewage markers in the Langat

394 River, Malaysia: Integration of biomarker and multivariate statistical approaches. *J. Environ.*

395 *Sci.* 24(9), 1600-1608. [https://doi.org/10.1016/S1001-0742\(11\)60979-0](https://doi.org/10.1016/S1001-0742(11)60979-0).

396 Amano, H., Nakagawa, K., Berndtsson, R. 2018. Surface water chemistry and nitrate pollution in

397 Shimabara, Nagasaki, Japan. *Environ. Earth Sci.* 77, 354.

398 <http://dx.doi.org/10.1007/s12665-018-7529-9>.

399 Amano, H., Nakagawa, K., Berndtsson, R. 2016. Groundwater geochemistry of a nitrate-contaminated

400 agricultural site. *Environ. Earth Sci.* 75, 1145. [http://dx.doi.org/ 10.1007/s12665-016-5968-8](http://dx.doi.org/10.1007/s12665-016-5968-8).

401 Albuquerque de Assis Costa, L., Maria Mano Pessoa, D., da Silva Carreira, R. 2018. Chemical and
402 biological indicators of sewage river input to an urban tropical estuary (Guanabara Bay, Brazil),
403 *Ecol. Indic.* 90, 513-518. <http://doi.org/10.1016/j.ecolind.2018.03.046>.

404 Baily, A., Rock, L., Watson, C.J., Fenton, O. 2011. Spatial and temporal variations in groundwater nitrate
405 at an intensive dairy farm in south-east Ireland: Insights from stable isotope data. *Agri. Ecosyst.*
406 *Environ.* 144(1), 308-318. <https://doi.org/10.1016/j.agee.2011.09.007>.

407 Casciotti, K.L., Sigman, D.M., Galanter Hastings, M., Böhlke, J.K., Hilkert, A. 2002. Measurement of the
408 oxygen isotopic composition of nitrate in seawater and freshwater using the denitrifier method.
409 *Anal. Chem.* 74(19), 4905-4912. <http://dx.doi.org/10.1021/ac020113w>.

410 Chandna, P., Khurana, M.L., Ladha, J.K., Punia, M., Mehla, R.S., Gupta, R. 2011. Spatial and seasonal
411 distribution of nitrate-N in groundwater beneath the rice–wheat cropping system of India: a
412 geospatial analysis. *Environ. Monit. Assess.* 178(1-4), 545-562.
413 <https://dx.doi.org/10.1007/s10661-010-1712-0>.

414 Chitsazan, M., Mohammad Rezapour Tabari, M., Eilbeigi, M. 2017. Analysis of temporal and spatial

415 variations in groundwater nitrate and development of its pollution plume: a case study in Karaj
416 aquifer. *Environ. Earth Sci.* 76, 391. <https://dx.doi.org/10.1007/s12665-017-6677-7>.

417 Committee on Nitrate Reduction in Shimabara Peninsula, 2016. The second term of Shimabara
418 Peninsula nitrate load reduction project, revised edn., Environmental Policy Division of
419 Nagasaki Prefectural Government, Nagasaki.
420 [http://www.pref.nagasaki.jp/bunrui/kurashi-kankyo/kankyohozen-ondankataisaku/shimaba](http://www.pref.nagasaki.jp/bunrui/kurashi-kankyo/kankyohozen-ondankataisaku/shimabara-chissofuka/286205.html)
421 [ra-chissofuka/286205.html](http://www.pref.nagasaki.jp/bunrui/kurashi-kankyo/kankyohozen-ondankataisaku/shimabara-chissofuka/286205.html), Accessed 6 November 2018 (in Japanese).

422 Costa, L.A.A., Pessoa, D.M.M., Carreira, R.S. 2018. Chemical and biological indicators of sewage
423 river input to an urban tropical estuary (Guanabara Bay, Brazil), *Ecol. Indic.* 90, 513-518.
424 <https://doi.org/10.1016/j.ecolind.2018.03.046>.

425 Eckhardt, D.A.V., Stacklberg, P.E. 1995. Relation of ground - water quality to land use on Long
426 Island, New York. *Groundwater.* 33(6), 1019-1033.
427 <https://doi.org/10.1111/j.1745-6584.1995.tb00047.x>

428 Froehner, S., Maceno, M., Martins, R.F. 2010. Sediments as a potential tool for assessment of sewage
429 pollution in Barigüi River, Brazil. *Environ. Monit. Assess.* 170(1), 261-272.
430 <http://dx.doi.org/10.1007/s10661-009-1230-0>.

431 Froehner, S., Martins, R.F., Errera, M.R. 2009. Assessment of fecal sterols in Barigui River sediments in
432 Curitiba, Brazil. *Environ. Monit. Assess.* 157(1-4), 591-600.
433 <http://dx.doi.org/10.1007/s10661-008-0559-0>.

434 Górski, J., Dragon, K., Kaczmarek, P.M.J. 2019. Nitrate pollution in the Warta River (Poland) between
435 1958 and 2016: trends and causes. *Environ. Sci. Pollut. Res.* 26, 2018.
436 <http://dx.doi.org/10.1007/s11356-017-9798-3>.

437 Hansen, B., Dalgaard, T., Thorling, L., Sørensen, B., Erlandsen, M. 2012. Regional analysis of
438 groundwater nitrate concentrations and trends in Denmark in regard to agricultural influence.
439 *Biogeosciences*, 9, 3277-3286. <http://dx.doi.org/10.5194/bg-9-3277-2012>.

440 He, D., Zhang, K., Tang, J., Cui, X., Sun, Y. 2018. Using fecal sterols to assess dynamics of sewage input
441 in sediments along a human-impacted river-estuary system in eastern China. *Sci. Total*
442 *Environ.* 636, 787-797. <https://doi.org/10.1016/j.scitotenv.2018.04.314>.

443 Hosono, T., Wang, C.H., Umezawa, Y., Nakano, T., Onodera, S., Nagata, T., Yoshimizu, C., Tayasu, I.,
444 Taniguchi, M. 2011. Multiple isotope (H, O, N, S and Sr) approach elucidates complex
445 pollution causes in the shallow groundwaters of the Taipei urban area. *J. Hydrol.* 397(1-2),

446 23-36. <https://doi.org/10.1016/j.jhydrol.2010.11.025>.

447 Hosono, T., Tokunaga, T., Kagabu, M., Nakata, H., Orishikida, T., Lin, I-T., Shimada, J. 2013. The use of
448 $\delta^{15}\text{N}$ and $\delta^{18}\text{O}$ tracers with an understanding of groundwater flow dynamics for evaluating the
449 origins and attenuation mechanisms of nitrate pollution. *Water Research*, 47(8), 2661-2675.
450 <https://dx.doi.org/10.1016/j.watres.2013.02.020>.

451 Hosono, T., Tokunaga, T., Tsushima, A., Shimada, J. 2014. Combined use of $\delta^{13}\text{C}$, $\delta^{15}\text{N}$, and $\delta^{34}\text{S}$ tracers
452 to study anaerobic bacterial processes in groundwater flow systems. *Water Res.*, 54, 284-296.
453 <https://dx.doi.org/10.1016/j.watres.2014.02.005>

454 Hussain, M. A., Ford, R., Hill, J. 2010. Determination of fecal contamination indicator sterols in an
455 Australian water supply system. *Environ. Monit. Assess.* 165(1), 147-157.
456 <http://dx.doi.org/10.1007/s10661-009-0934-5>.

457 Itoh, J., Tatsukawa, R. 1978. Fecal pollution of the river waters in the Matsuyama Plain, using
458 coprostanol as an indicator. *Jap. J. Limnol*, 39(3), 123-129.

459 Kaown, D., Koh, D.C., Maer, B., Lee, K.K. 2009. Identification of nitrate and sulfate sources in
460 groundwater using dual stable isotope approaches for an agricultural area with different land

461 use (Chuncheon, mid-eastern Korea). *Agri. Ecosyst. Environ.* 132(3-4), 223-231.

462 <https://doi.org/10.1016/j.agee.2009.04.004>.

463 Kendall, C. 1998. Tracing Nitrogen Sources and Cycling in Catchments, in *Isotope Traces in Catchment*

464 *Hydrology* (C. Kendall and J.J. McDonnell Eds.), Elsevier Science B.V., Amsterdam, 519-576.

465 Liu, G.D., Wu, W.L., Zhang, J. 2005. Regional differentiation of non-point source pollution of

466 agriculture-derived nitrate nitrogen in groundwater in northern China. *Agri. Ecosyst. Environ.*

467 107(2-3), 211-220. <https://dx.doi.org/10.1016/j.agee.2004.11.010>.

468 Martins, C.C., Fillmann, G., Montone, R.C. 2007. Natural and anthropogenic sterols input in surface

469 sediments of Patos Lagoon, Brazil. *J. Braz. Chem. Soc.* 18(1), 106-115.

470 <http://dx.doi.org/10.1590/S0103-50532007000100012>.

471 Matic B.I., Grujić, S., Jauković, Z., Laušević, M. 2016. Sterol ratios as a tool for sewage pollution

472 assessment of river sediments in Serbia. *Environ Pollut.* 213, 76-83.

473 Nakagawa, K., Watanabe, T., Amano, H. 2015. Potential map accuracy of groundwater nitrate load from

474 agriculture activities in Shimabara City, Nagasaki Prefecture, Japan. *J. Groundw. Hydrol.* 57(4),

475 483-493. (in Japanese with English Abstract). <https://doi.org/10.5917/jagh.57.483>.

476 Nagasaki Prefecture (2017) Nagasaki Prefecture waste water treatment conception in 2017,
477 <http://www.pref.nagasaki.jp/bunrui/kurashi-kankyo/mizukankyo/osuishori/osuishorizenpa>
478 [n/283418.html](http://www.pref.nagasaki.jp/bunrui/kurashi-kankyo/mizukankyo/osuishori/osuishorizenpan/283418.html) (in Japanese)

479 Nakagawa, K., Amano, H., Asakura, H., Berndtsson, R. 2016. Spatial trends of nitrate pollution and
480 groundwater chemistry in Shimabara, Nagasaki, Japan. *Environ. Earth Sci.* 75, 234.
481 <http://dx.doi.org/10.1007/s12665-015-4971-9>.

482 Nakagawa, K., Amano, H., Takao, Y., Hosono, T., Berndtsson, R. 2017. On the use of coprostanol to
483 identify sources of nitrate pollution in groundwater, *J. Hydrol.* 550, 663-668.
484 <http://dx.doi.org/10.1016/j.jhydrol.2017.05.038>.

485 Oyanagi, W., Ando, Y., Mizusawa, S., Moriyama, N. 2005. Salt composition characteristic of animal
486 waste composts. *Jpn. J. Sci. Plant Nutr.* 75(1), 91-93. (in Japanese)

487 QGIS Development Team. 2019. QGIS geographic information system. Open Source Geospatial
488 Foundation Project. <http://qgis.osgeo.org> (accessed May 17, 2019).

489 Rada, J.P.A., Duarte, A.C., Pato, P., Cachada, A., Carreira, R.S. 2016. Sewage contamination of sediments
490 from two Portuguese Atlantic coastal systems, revealed by fecal sterols. *Marine Pollut. Bull.*
491 103(1-2), 319-324. <https://doi.org/10.1016/j.marpolbul.2016.01.010>.

492 Reeves, A. D., Patton, D. 2005. Faecal sterols as indicator of sewage contamination in estuarine sediments
493 of the Tay Estuary, Scotland: an extended baseline survey. *Hydrol. Earth Syst. Sci.* 9, 81-94.
494 <http://dx.doi.org/10.5194/hess-9-81-2005>.

495 Ribbe, L., Delgado, P., Salgado, E., Flügel, W.A. 2008. Nitrate pollution of surface water induced by
496 agricultural non-point pollution in the Pochay watershed, Chile. *Desalination*, 226(1-3),
497 13-20. <http://dx.doi.org/10.1016/j.desal.2007.01.232>

498 Sigman, D. M., Casciotti, K. L., Andreani, M., Barford, C., Galanter, M., Böhlke, J. K. 2001. A bacterial
499 method for the nitrogen isotopic analysis of nitrate in seawater and freshwater. *Anal. Chem.*
500 73(17), 4145-4153. <http://dx.doi.org/10.1021/ac010088e>.

501 Sorensen, J.P.R., Butcher, A.S., Stuart, M.E., Townsend, B.R. 2015. Nitrate fluctuations at the water table:
502 implications for recharge processes and solute transport in the Chalk aquifer. *Hydrol. Process.*
503 29(15), 3355-3367. <http://dx.doi.org/10.1002/hyp.10447>.

504 Sugimoto, T. 2006. Geology and petrology at Shimabara Peninsula, Kyushu, SW Japan - from recent
505 results. *J. Geotherm. Res. Soc. Jpn.* 28(4), 347-360.

506 WHO (World Health Organization). (2011). *Guidelines for drinking water quality*, 4th edn. WHO
507 Press, Geneva