

1 **Collection and aging of greater amberjack *Seriola dumerili* larvae and juveniles around the**  
2 **Penghu Islands, Taiwan**

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17  
18 **Running head:**

19 Age of *Seriola dumerili* larvae

20  
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23

## 24 **Abstract**

25 In order to investigate the early life history of *Seriola dumerili*, we first validated otolith daily increments  
26 using reared-fish (11-51 days after hatching). Four larval and early-juvenile *S. dumerili* were collected in May and July  
27 2015 around the Penghu Islands, Taiwan (23.45-23.70 °N, 119.40-119.70 °E) by surface larval net towing but not from  
28 drifting seaweeds. *Seriola dumerili* were caught at thermal front, and total lengths and ages ranged 7.4-42.5 mm and  
29 18-56 days, respectively. Our results indicate that the hatching dates of *S. dumerili* were April to June and larvae may  
30 have been accumulated in frontal zone before juvenile phase.

31 **Key words:** Greater amberjack, *Seriola dumerili*, aging, otolith, early life history

## 33 **Introduction**

34 The greater amberjack *Seriola dumerili* (family Carangidae) is distributed widely from temperate to tropical  
35 waters around the world (Taki et al. 2005), and is an important species both for fishery and aquaculture in Japan  
36 (Nakada 2002). Spawning and morphological development of artificially-raised *S. dumerili* were described in Japan  
37 (Masuma et al. 1990; Tachihara et al. 1993; Kawabe et al. 1996; Kawabe et al. 1998), and its spawning season in the  
38 wild was estimated to be from winter to summer in western Atlantic (Fahay 1975; Wells and Rooker 2004b; Sedberry et  
39 al. 2006; Harris et al. 2007) and June and July in Mediterranean Sea (Raya and Sabatés 2015). Juveniles of *S. dumerili*  
40 in 25-297 mm standard length (SL) associate with floating objects such as drifting seaweeds (Nakata et al. 1988;  
41 Badalamenti et al. 1995; Massutí et al. 1999; Wells and Rooker 2004a, b). Based on an alizarin complexone marking  
42 experiment, it was confirmed that *S. dumerili* (136-193 mm SL) deposits otolith increments on a daily basis (Wells and  
43 Rooker 2004b). However, there is no information regarding deposition of otolith increments in larval and early-juvenile  
44 stages of this species in the wild, which is necessary to determine spawning season from hatching date and early life  
45 history of this species. Information of larval and early-juvenile *S. dumerili* is limited in the wild, with at most 15  
46 individuals (less than 0.81 ind./1000 m<sup>3</sup>) in Mediterranean Sea (Raya and Sabatés 2015), 11 individuals in the South  
47 Atlantic Bight (Fahay 1975) and less than 0.01 ind./1000 m<sup>3</sup> in the northern Gulf of Mexico (Ditty et al. 2004) have  
48 been collected. As for Asian waters, information of larval and early-juvenile *S. dumerili* is also limited, with few  
49 collection records from the basin of Tsushima Warm Current (TWC) and off southern Korea from July to August  
50 (Uchida et al. 1958), in the Pacific coast of Japan from March to September (Okuyama 2014), and in the coastal waters  
51 of Taiwan (Liu 2001) and in the northeast of Taiwan (Chen et al. 2012).

52 In order to facilitate understanding of the early life history of *S. dumerili* in Asian waters, firstly, we validated  
53 the otolith daily increments of artificially-raised *S. dumerili* larvae and juveniles. Next, we collected larval and  
54 early-juvenile *S. dumerili* around the Penghu Islands, Taiwan (Fig. 1), and investigated hydrographic conditions, larval  
55 fish densities of collection sites, and the age and spawning season of this species.

## 57 **Materials and methods**

### 58 *Validation for Otolith Daily Increments*

59 Artificially-raised *S. dumerili* were obtained from a private hatchery (Tawaki Suisan, Ltd.), in Kumamoto  
60 prefecture, Japan. Fish were reared at 24 °C and rotifers were fed until day 23 after hatching, followed by *Artemia*  
61 between day 20-35, frozen copepods between day 31-41 and dry pellets from day 33. Fish were randomly sampled from  
62 the rearing tank on day 11 ( $n=5$ ), 32 ( $n=5$ ) and 51 ( $n=4$ ), and all samples were kept frozen until analysis. Total lengths  
63 (TL, mm) of fish from each age group were measured and pairs of sagittal otoliths were extracted under a dissecting  
64 microscope. According to the method of Sakakura and Tsukamoto (1997), otoliths were embedded in epoxy resin lying

65 on their sides on a glass slide, except samples on day 11 which were embedded in transparent nail polish. The otoliths  
66 were observed after grinding using sandpaper (#1000) and lapping film (9  $\mu\text{m}$  and 3  $\mu\text{m}$ ). Growth increments were  
67 counted under a light microscope at a magnification of  $\times 1000$  using an oil immersion lens. Counting of growth  
68 increments started from a conspicuous dark mark which delimited the core of the otolith. The largest radius of sagittal  
69 otolith was also measured using a digital microscope (Keyence, VH6300).

70

#### 71 *Field collection*

72 Cruises around the Penghu Islands, Taiwan (Fig. 1, Fig. 2, 23.45-23.70 °N, 119.40-119.70 °E) were made during  
73 13-15 May, 2-4 June, 28-29 July and 25-27 August 2015 by R/V Hai-an (42 tonnes) of Taiwan Fishery Research  
74 Institute. During 09:00-14:00 (at National Standard Time), drifting seaweed and frontal zone were visually observed  
75 and drifting seaweeds were scooped together with associated fishes by a hand net ( $\Phi 45$  cm, 3 mm mesh) from the side  
76 of stationary ship, since sea surface was close to scoop drifting seaweeds from the deck. In this paper, we defined  
77 “frontal zone” as the area of surface water convergence including oceanic front created by the gradient of water  
78 temperature and/or salinity, and slick created by Langmuir circulation or internal wave. Surface tows of a larval net  
79 ( $\Phi 1.3$  m, 0.33 mm mesh) from stern were conducted (10 min. with towing speed at 2 knot) in frontal zones and other  
80 areas (Fig. 2a). In order to keep towing at surface layer, a small spherical float was attached to the outer part of the  
81 opening of a larval net. A HYDRO-BIOS flow meter was placed at the opening of a larval net to measure the volume of  
82 water filtered. Larval and juvenile fishes were preserved in 95% ethanol solution. At each sampling station, vertical  
83 profile of water temperature was measured by a conductivity-temperature-depth profiler (CTD; SBE-19 plus, Sea-Bird  
84 Electronics, Bellevue, WA) from the sea surface to a depth at 5 m above the bottom.

85

#### 86 *Sample analysis*

87 Larval and juvenile fishes were counted and *S. dumerili* was identified according to Okiyama (1988; 2014).  
88 Then, total length ( $\pm 0.1$  mm TL) and wet weight (mg) were measured with a caliper and an electronic balance,  
89 respectively. In the same manner of reared-fish, otoliths of each wild fish were examined and hatching dates were  
90 back-calculated. Horizontal distribution of sea surface temperature (SST, °C) from *in situ* CTD data was summarized  
91 and plotted by Ocean Data View (version 4.6.2).

92

#### 93 *Data analysis*

94 Comparison of regression lines for otolith increments related to the days after hatching in reared-fish and the  
95 line of  $Y=X$  was conducted with the analysis of covariance (ANCOVA). Exact Wilcoxon rank sum test was used for  
96 comparison of larval and juvenile fishes density between frontal zones and other stations. Statistical analysis was  
97 carried out using R. version 3.1.3 (R Development Core Team 2015) supplied with the exactRankTests package  
98 (Hothorn and Hornik 2015) and  $p$ -values  $< 0.05$  were considered significant in all analyses.

99

## 100 **Results**

### 101 **Validation of otolith daily increments**

102 Total lengths of artificially-raised *S. dumerili* at each age group were  $4.5 \pm 0.5$  mm,  $16.7 \pm 1.8$  mm and  $54.8$   
103  $\pm 3.6$  mm (mean  $\pm$  standard deviation) in day 11 ( $n=5$ ), 32 ( $n=5$ ) and 51 ( $n=4$ ), respectively. The relationships between  
104 age ( $x_{\text{day}}$ , days after hatching) and otolith radius ( $y_{\text{rad}}$ ,  $\mu\text{m}$ ), and between TL ( $x_{\text{TL}}$ , mm) and otolith radius ( $y_{\text{rad}}$ ,  $\mu\text{m}$ ) of

105 reared-fish, were described by the following equations:  $y_{\text{rad}} = 13.49 \cdot e^{(0.08x_{\text{day}})}$  ( $n=14, r=0.99$ , Fig. 3a), and  
106  $y_{\text{rad}} = 11.71 \cdot x_{\text{TL}} - 18.49$  ( $n=14, r=0.98$ , Fig. 3b), respectively. The linear regression between age ( $x_{\text{day}}$ , days after  
107 hatching) and number of otolith increments ( $y_{\text{inc}}$ ) was equated as  $y_{\text{inc}} = 0.99 \cdot x_{\text{day}} + 0.23$  ( $n=14, r=0.99$ , Fig. 3c). The  
108 regression line for otolith increments related to age in reared-fish was not significantly different from the line of  $Y=X$   
109 (ANCOVA,  $n=14, df=1, F=1.04, p=0.32$ ).

110

### 111 Collections of larvae

112 Frontal zones marked with the accumulation of bubbles and/or a distinct sea surface line were visually  
113 observed around the Penghu Islands and they were found to be created by gradient of water temperature but not by the  
114 difference of salinity, especially in May and July (Fig. 2). Some of frontal zones were not correlated with water  
115 temperature gradient (e.g. A11, C2, Fig. 2). We caught a total of 898 of larval and juvenile fishes from 30 hauls of  
116 surface towing of a larval net. During our study period, dominant families (monthly mean ind./100 m<sup>3</sup>) from surface  
117 towing were Carangidae (0.54-0.17; mostly doublespotted queenfish *Scomberoides lysan*), Exocoetidae (4.82-0.01) and  
118 Coryphaenidae (0.37-0.05). Density of larval and juvenile fishes collected by surface towing was not different between  
119 frontal zones and other stations in May (exact Wilcoxon rank sum test,  $n=10, df=1, W=7, p=0.71$ ). Drifting seaweeds  
120 were found in four stations only in May (Fig. 2). A total of 144 fish juveniles associated with drifting seaweeds were  
121 collected, and *Siganus* spp. (54.2%) and threadsail filefish *Stephanolepis cirrhifer* (27.8%) were dominant. A total of  
122 four *S. dumerili* were caught by surface towing of a larval net, but not from drifting seaweeds. All *S. dumerili* were  
123 caught at frontal zones (Fig. 2a, c) and SST of collection sites ranged from 24.9 to 27.4 °C (Table 1). In the station A11,  
124 two *S. dumerili* were collected in May. Total lengths and ages of *S. dumerili* were 7.4, 9.8, 13.7 and 42.5 mm, and 18,  
125 26, 36 and 56 days, respectively (Table 1). The relationship between age ( $x_{\text{day}}$ , days after hatching) and TL ( $y_{\text{TL}}$ , mm) of  
126 reared-fish was described by the following equation:  $y_{\text{TL}} = 2.26 \cdot e^{(0.06x_{\text{day}})}$ , and growth rate of wild fish was lower than  
127 that of reared-fish (Fig. 3d).

128

### 129 Discussion

130 We confirmed that deposition of otolith increments of *S. dumerili* is daily basis in larval and early-juvenile  
131 stages. Wells and Rooker (2004b) reported that *S. dumerili* deposit otolith increments on a daily basis in juvenile stage  
132 (136-193 mm SL) by a series of alizarin complexone marking experiment. Thus, age determination using sagittal otolith  
133 is valid for larval and early-juvenile stages of this species. Growth rate of wild fish was lower than that of reared-fish.  
134 Sakakura and Tsukamoto (1997) reported that growth rate of wild early-juvenile yellowtail *S. quinqueriata* was lower  
135 than that of reared-fish, which coincided with our results of *S. dumerili*.

136 We could collect larval and juvenile *S. dumerili* only around the Penghu Islands, Taiwan in the present study.  
137 Larval and juvenile *S. dumerili* had been caught in the coastal waters of Taiwan (Liu 2001, 15 ind.) and in the northeast  
138 of Taiwan in June (Chen et al. 2012, 0.13 ind./m<sup>3</sup>), indicating that *S. dumerili* in the early life stages distribute around  
139 Taiwan. However, *S. dumerili* was not collected by surface towing of a larval net from our two preliminary surveys in  
140 2015. The first was the cruise to cover the north-eastern part of Taiwan (a total of 15 hauls, 2-5 May; 25.0-26.5 °N,  
141 120.5-123.0 °E) by the R/V Fishery Researcher 1 and the second was the cruise to cover the south-western part of  
142 Taiwan (a total of 11 hauls, 25-27 July; 22.5-23.5 °N, 118.6-119.8 °E) by the R/V Fishery Researcher 2, Taiwan  
143 Fishery Research Institute, Council of Agriculture. In the southern U.S., the spawning area of *S. dumerili* is estimated

144 around the shelf-edge reef sites of 50-100 m depth (Sedberry et al. 2006). Our sampling stations around the Penghu  
145 Islands are mostly shallower than 50 m depth (Fig. 1b, Fig. 2), and the spawning area of *S. dumerili* is presumably  
146 located in the open water area off the Penghu Islands deeper than 50 m depth in our study period. In May and July 2015,  
147 frontal zones were created by the gradient of water temperature around the Penghu Islands. These frontal zones may be  
148 formed between coastal water around the Penghu Islands and the water mass intruded from the open sea. It is possible  
149 that some of frontal zones were created by Langmuir circulation or internal wave, because the number of CTD data was  
150 small to fully explain the formative factors of frontal zones. Further study is needed to understand formative factors of  
151 frontal zones around the Penghu Islands. Since larval and juvenile *S. dumerili* were collected only in frontal zones in  
152 our study, they may be spawned in the open sea side and eggs or larvae were accumulated in the frontal zones around  
153 the Penghu Islands, where total density of larval and juvenile fishes was not different between frontal zones and other  
154 stations. Raya and Sabatés (2015) reported that distribution of larval *S. dumerili* was limited by the position of thermal  
155 front in the northwestern Mediterranean Sea, which is similar to our study. SST of frontal zones in May ranged from  
156 24.9 to 25.5°C (Table 1), and frontal zones were suitable temperature for larval and juvenile *S. dumerili* because Raya  
157 and Sabatés (2015) mentioned that temperature preference of *S. dumerili* larvae is in between 24 and 25 °C. *Seriola*  
158 *dumerili* was not found with drifting seaweeds in this study. Juveniles of *S. dumerili* have been reported to associate  
159 with floating objects at 25-297 mm SL (Nakata et al. 1988; Badalamenti et al. 1995; Massuti et al. 1999; Wells and  
160 Rooker 2004a, b). Since our specimens were mainly in larval or early-juvenile stage, they may stay in frontal zone until  
161 they reach the body size at associating with drifting seaweeds.

162 The time from fertilization to hatching is only 36 to 45 h in *S. dumerili* at 23.1-23.7 °C (Masuma et al.  
163 1990), so our back-calculated hatching dates indicated that spawning of this species occurred in April and June 2015  
164 around the Penghu Islands. In this study, individuals with similar age were caught in May 2015, suggesting that  
165 concurrent spawning event occurred in April around the Penghu Islands. Taking into account the hatching date in 2015  
166 (April and early June), *S. dumerili* may spawn from spring to early summer around the Penghu Islands. In Asian waters,  
167 the spawning season of *S. dumerili* is estimated from November to March in South China Sea (off Hainan Island, China  
168 to Viet Nam) and from May to June around Japan (Hamada and Soyano 2009). Spawning season of *S. dumerili* around  
169 Taiwan seems to be between that of South China Sea and Japan. Previous studies in other waters also showed that the  
170 majority of spawning of *S. dumerili* occur between winter to summer: February to April by hatching-date analysis off  
171 Galveston, Texas (Wells and Rooker 2004b), January to June with peak spawning in April and May in South Atlantic  
172 (Sedberry et al. 2006) and off the Southeastern U.S. Atlantic (Harris et al. 2007), and in winter in the western Atlantic  
173 (Fahay 1975) and in June and July in Mediterranean Sea (Raya and Sabatés 2015). Synthesizing these evidences,  
174 spawning season of this species may range from winter to summer in Asian waters.

175 It is reported that densities of larval *S. dumerili* were higher by surface towing than oblique towing and no  
176 significant difference in day/night was detected in Mediterranean Sea (Raya and Sabatés 2015). Since number of *S.*  
177 *dumerili* in our study is not enough to reveal its early life history such as distribution and growth rate, in the future study  
178 it is needed to increase number of samples including verification of the distributional layer due to diel vertical migration  
179 by modifying sampling methods.

180

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189

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242  
243 台湾澎湖島周辺で採集されたカンパチ*Seriola dumerili*仔稚魚の日齢

244  
245 長谷川隆真・葉信明・陳君如・郭慶老・河邊玲・阪倉良孝  
246 短報ICRE-D-16-00069R1

247  
248 カンパチ*Seriola dumerili*の初期生態を調べるために、まず人工種苗（11-51日齢）を用いて  
249 耳石日周輪のバリレーションを行った。次に、2015年5月および7月に台湾澎湖島周辺  
250 （23.45-23.70 °N, 119.40-119.70 °E）で稚魚ネットの表層曳きにより計4尾のカンパチ仔稚魚  
251 を採集した。これらは流れ藻には附随していなかった。カンパチ仔稚魚は水温フロントで  
252 採集され、その全長は7.4-42.5 mm, 18-56日齢であった。以上の結果からカンパチの孵化日  
253 は4月から6月で、カンパチの仔稚魚はフロントに集積されると推測された。

254  
255 （長谷川・阪倉：〒852-8521 長崎県長崎市文教町 1-14 長崎大学大学院水産・環境科学  
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259

260 **Figure captions**

261 **Fig. 1** Map showing (a) geographical location of study area and bathymetric chart around Taiwan, and (b) study area  
262 and the bathymetry in meters (*thin solid* contours with numbers) around the Penghu Islands. ECS is the East China Sea  
263 and SCS is the South China Sea

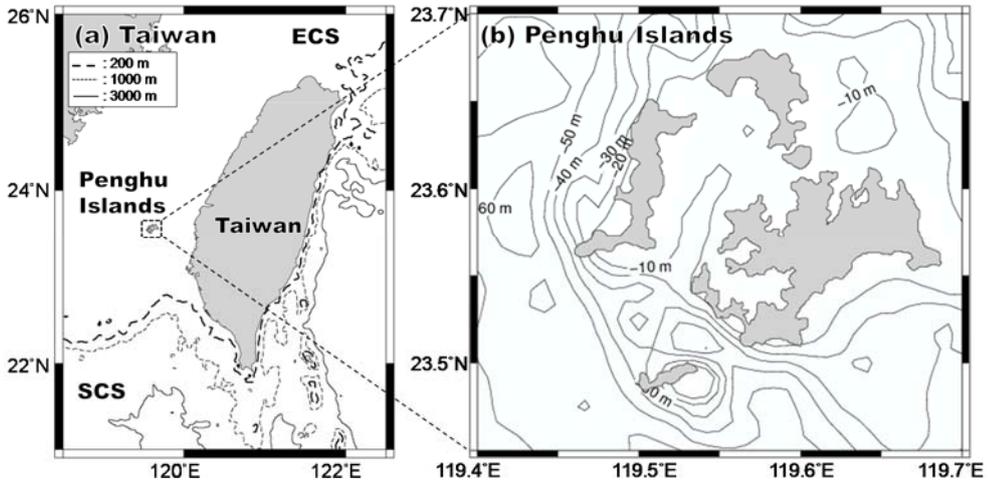
264

265 **Fig. 2** The estimated sea surface temperature (SST) in (a) 13-15 May, (b) 2-4 June, (c) 28-29 July and (d) 25-27  
266 August 2015 around the Penghu Islands, Taiwan. *Open triangle, square* and *circles* are the stations where frontal zone,  
267 drifting seaweeds were found and CTD casting, respectively. *Filled triangles* denote the stations where *S. dumerili* were  
268 collected

269

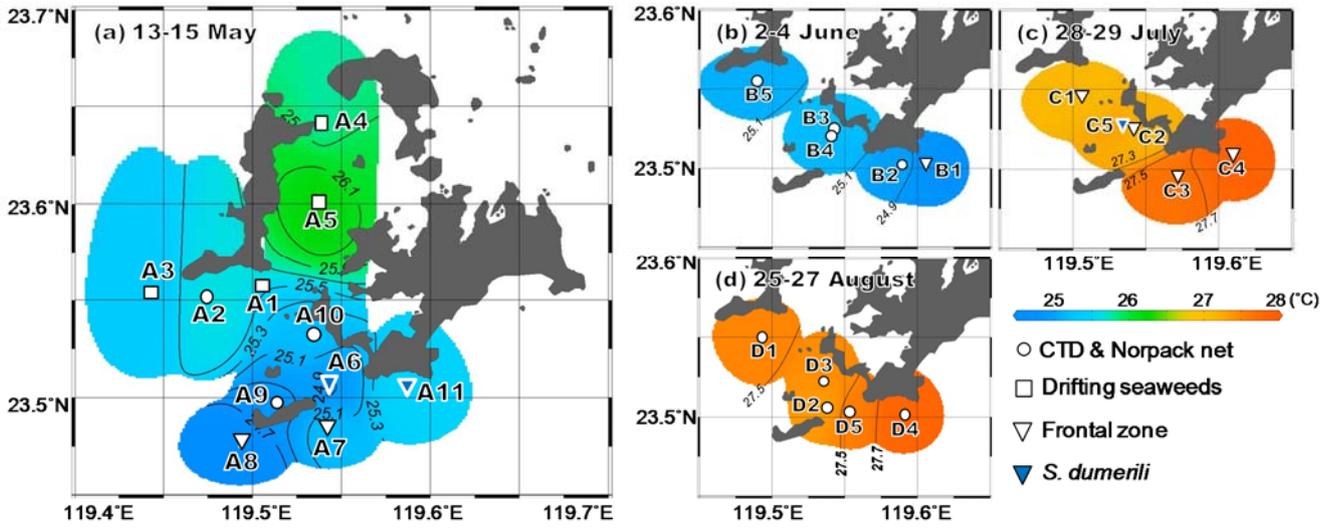
270 **Fig. 3** Relationship between (a) age (days) and otolith radius ( $\mu\text{m}$ ), (b) total length (mm) and otolith radius, (c) age and  
271 otolith increments and (d) age and total length. Panel (a)-(b) show reared-fish and panel (d) shows reared- and wild-fish.  
272 *Break line* in the panel (c) shows the line of  $Y=X$ . In the panel (d), *open circles, filled circles* and *solid line* indicate  
273 reared-fish, wild-fish and growth curve of reared-fish, respectively. The ages of reared-fish are days after hatching, and  
274 the ages of wild fish are determined by otolith analysis

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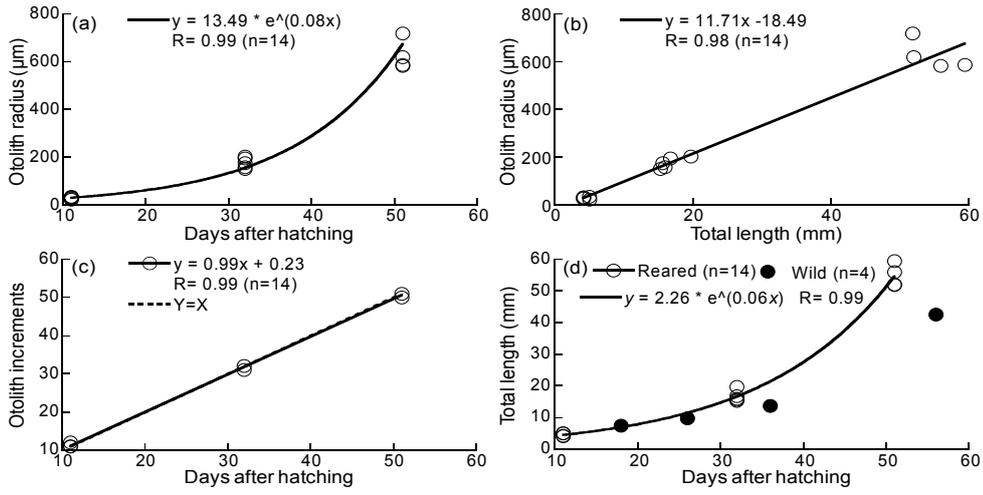
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