Short Communication

Evaluation for effect of hypothermia on the disposition of 4-nitrophenol in rats by *in vitro* metabolism study and rat liver perfusion system

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RUNNING HEAD: Effect of hypothermia on the disposition of 4-nitrophenol

16 Abstract:

17 Objectives. The aim of this study was to evaluate the effect of hypothermia on the *in*

18 *vivo* pharmacokinetics of 4-nitrophenol (4NP) using rat liver homogenate and rat liver

19 perfusion system.

20 Methods. Rat liver homogenate was incubated with 4NP, which is mainly metabolized

21 by CYP2E1, at 37, 34, 32, or 28ºC. The Michaelis constant (K_m) and maximum

22 elimination velocity (V_{max}) of 4NP were calculated by a Hanes-Woolf plot. The hepatic

23 extraction ratio (E_h) of 4NP was evaluated in a rat liver perfusion study at 37, 34, 32, or

24 28ºC. Moreover, the plasma concentration profiles of 4NP after its i.v. administration

25 to rats were analyzed by the moment theory and were compared to *in vitro* parameters.

26 Key findings. While the K_m of 4NP was not changed, the V_{max} and E_h were reduced at

27 low temperatures. The plasma concentrations of 4NP after its i.v. administration to rats

28 were significantly increased at 28ºC.

29 Conclusion. Changes in the pharmacokinetics of 4NP under hypothermic conditions

30 were caused by alterations in V_{max} and E_h. We may be able to predict the disposition of a

31 drug by *in vitro* studies.

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33 KEY WORDS: 4-nitrophenol, hypothermia, CYP2E1, liver perfusion

34

Introduction

Therapeutic hypothermia is beneficial for patients with acute myocardial infarction or post cardiac arrest syndrome [1-5]. Several drugs such as propofol, midazolam, or dexmedetomidine have been used during hypothermia to cause a sedative action or negate the complications of hypothermia [5, 6]. However, the pharmacokinetics of drugs used during therapeutic hypothermia have been shown to be altered [7-9]. We need to identify the factors affecting the disposition of a drug to optimize medication. We previously reported that the pharmacokinetics of phenolsulfonphthalein (PSP), indocyanine green (ICG), and fluorescein isothiocyanate-dextran (FD-4, MW 4400) as marker compounds and their hepatic disposition under hypothermic conditions in rats could differ with the disposition route and intrinsic clearance of these drugs [10, 11]. Despite of necessity to determine the individual factors affecting drug disposition for prediction the pharmacokinetics during hypothermia, it has not been clarified. In this study, we tried to evaluate the effect of temperature on drug disposition by focusing on the hepatic disposition.

In this study, we chose 4-nitrophenol (4NP) as a model compound metabolized in the liver by CYP2E1 [12, 13]. We thought we could evaluate the effect of hypothermia on a drug disposition metabolized by CYP2E1 which plays a major role in the metabolism

53   of several drugs used during hypothermia such as acetaminophen, isoflurane, isoniazid,

54   and theophylline [14-18].

55   In the present study, we examined the effect of low temperature on the CYP2E1

56   activity and hepatic extraction ratio of 4NP.   Moreover, we evaluated the relationship

57   between the *in vitro* and *in vivo* pharmacokinetic parameters of 4NP to evaluate the

58   possibility of predicting changes in the pharmacokinetics of drugs under hypothermic

59   conditions by an *in vitro* study.

60

61   Materials and Methods

62   Materials

63   4NP was purchased from Nacalai Tesque, Inc. (Kyoto, Japan).   All chemicals

64   were of the highest purity available.

65

66   Animals

67   Male Wistar rats (180-210 g or 240-270 g) were housed in a cage in an air-

68   conditioned room and maintained on a standard laboratory diet (MF, Oriental Yeast, Co.,

69   Ltd., Tokyo, Japan) and water *ad libitum*.   All animal experiments in the present study

70   conformed to the Guidelines for Animal Experimentation of Nagasaki University and

71 were approved by the Committee of Animal Experimentation of Nagasaki University

72 (Approval number: 0506280443).

73

74 Liver homogenate study

75   The liver was removed from male Wistar rats (180-210 g) and homogenized in

76 cold Tris/HCl buffer containing 5 mM $MgSO_4$ (pH 7.4).   Rat liver homogenate was

77 incubated at 37, 34, 32, or 28°C for 15 min after the addition of 4NP (50, 100, 200, 400,

78 800 µg/mL) and several lots of the liver homogenate were used in this study.   We have

79 preliminary examined the elimination of 4NP from liver homogenate until 15 min

80 followed the first elimination manner (data not shown).   After incubation, the

81 incubation mixture was mixed with acetone to stop the metabolism reaction and

82 centrifuged for 5 min at 15,000 rpm.   The remaining concentration of 4NP was

83 determined by spectrophotometer and then the eliminate velocity of 4NP was calculated

84 by Eq.1

85 $$v = \frac{(C_0 - C_{15}) \times V}{15} \div mg\ protein \qquad\qquad (1)$$

86   where v is the eliminate velocity, C_0 and C_{15} represent the concentration of 4NP

87 at 0 and 15 min, respectively, and V is the incubation volume (1 mL).

88   The Michaelis constant (Km) and maximum eliminate velocity (Vmax) were

89   calculated by a Hanes-Woolf plot (Eq.2).

90    $$\frac{C}{v} = \frac{1}{V_{max}} \times C + \frac{K_m}{V_{max}}$$ (2)

91   where v is the eliminate velocity normalized by protein content of the

92   homogenate, C is the concentration of 4NP, K_m is the Michaelis constant, and V_{max} is the

93   maximum eliminate velocity.

94

95   Liver perfusion study

96   Male Wistar rat liver was perfused *in situ* as described previously [11].   After a

97   stabilization period of 30 min, the 4NP solution (20 mg/mL x 0.1 mL) was injected into

98   the perfusion route.   After administration of the 4NP solution, venous outflow samples

99   were collected into tubes for 5 min.   The hepatic extraction ratio (E_h) was calculated as

100   follows Eq 3 with the assumption that the hepatic disposition of 4NP was allowed to well-

101   stirred model.

102   $$E_h = \frac{D - C_{out} \times V_{out}}{D}$$ (3)

103   where D is administration dose of 4NP, C_{out} is concentration of 4NP in outflow

104   effluent and V_{out} is the volume of outflow effluent.

105   *In vivo* study

106   Male Wistar rats (240-270 g) were anesthetized with sodium pentobarbital (50

107   mg/kg, i.p.) and the left femoral artery was cannulated with a polyethylene tube (i.d. 0.25

108   mm, o.d. 0.61 mm, Dual Plastics, Dural, Australia).

109   Rats were divided into four groups: a control group in which rectal temperature

110   was maintained at 37ºC by a heat lamp throughout the procedure; a hypothermic group

111   kept at 34ºC, 32ºC, or 28ºC, in which hypothermia was induced by external cooling with

112   icepack on their body before the administration of the drug, and rectal temperature was

113   maintained at 34, 32, or 28ºC.

114   The drug solution (20 mg/mL x 0.1 mL) was injected into the right femoral vein.

115   After administration of the drug solution, blood was collected at the selected times from

116   the heparinized cannula inserted into the femoral artery until 50 min.   Blood was

117   centrifuged at 15000 rpm for 5 min.

118   Moment parameters (AUC_p and MRT_p) were calculated by numerical integration

119   using a linear trapezoidal formula and extrapolation to infinite time based on a

120   monoexponential equation [19].

121   Assay

122   The concentration of 4NP was determined spectrophotometrically at 410 nm

123   after dilution with 1 M NaOH [20].

124

125   Statistical analysis

126   Statistical comparisons were performed by Dunnett's test after examining with

127   an analysis of variance (ANOVA) or repeated measured ANOVA. $P < 0.05$ was

128   considered to be indicative of significance compared to control group (37°C).   The

129   results were expressed the mean ± S.E.

130   Results

131   Effect of temperature on 4NP metabolism in rat liver homogenate

132   The Michaelis constant (K_m) and maximum eliminate velocity (V_{max}) of 4NP at

133   37, 34, 32, and 28°C were obtained from the Hanes-Woolf plot (fig.1A).   The V_{max} of

134   4NP was decreased to about 30% at 32°C and 70% at 28°C compared to 37°C and there

135   was a significant difference between 37°C and 28°C. (Fig 1B)   The Km of 4NP was not

136   altered at 34°C and 32°C compared to 37oC. (Fig 1B)   The Km at 28°C was decreased

137   by the half from 37°C although not significant.

138

139   Change in the hepatic extraction ratio at low temperatures in the rat liver perfusion system

140   Fig.2 illustrates the E_h of 4NP obtained by the rat liver perfusion study at each

141   temperature.   The E_h at 28°C was decreased about 15% compared to 37°C although the

142   difference was not significant.   The E_h of 4NP was linearly decreased according to the

143   reduction of temperature (r^2=0.931, p=0.034).

144

145   Pharmacokinetics of 4NP *in vivo* in rats under hypothermic conditions

146   Fig.3 shows the plasma concentration – time profile of 4NP after its i.v.

147   administration to rats under different body temperatures.   The plasma concentration of

148   4NP at 28°C was significantly higher than that at 37°C.   The AUC_p, MRT_p, and CL_{tot} of

9

149   4NP at each temperature are listed in Table I.   The AUC_p of 4NP was 1.7 (34°C), 2.9

150   (32°C), and 5.5 (28°C) times greater than that at 37°C, and the MRT_p of 4NP was

151   significantly prolonged at 32 and 28°C.   In addition, the CL_{tot} of 4NP was significantly

152   lower at 32 and 28°C than that at 37°C.

153

154   Discussion

155   We performed an *in vitro* metabolism study using rat liver homogenate and

156   isolated liver perfusion study to evaluate the effect of temperature on the elimination in

157   the liver homogenate and E_h of 4NP.   We evaluated the effect of temperature on 4NP

158   metabolism activity in rat liver homogenate since liver homogenate containing the

159   metabolic enzymes or co-enzymes necessary to metabolize drugs and easy to handling

160   compared to another method.   The V_{max} of 4NP decreased according to the temperature,

161   while no significant difference was observed in K_m.   This suggests that the affinity of

162   4NP with CYP2E1 was not affected by temperature, whereas the eliminate velocity could

163   have been altered under hypothermic conditions.   Similar to our result, it has been

164   reported that the Vmax of midazolam metabolized by CYP3A4 was decreased at 33°C

165   compared to 37°C while the Km was not altered [21].   The previous study [18] has shown

166   that NADPH, NADPH-cytochrome P-450 reductase, and lipids are required for

10

167   metabolism by CYP.   These factors also produced by enzymatic reaction and the activity

168   of these enzymes could also be decreased under hypothermia.   Further study is needed

169   to clarify the mechanisms the change in Vmax under hypothermic condition.

170   Moreover, we performed an isolated liver perfusion study to analyze changes in

171   the E_h of 4NP at low temperatures.   The isolated liver perfusion study is a useful method

172   to evaluate the effect of temperature on the hepatic uptake of 4NP because we can easily

173   control the flow rate and perfusion temperature.   Since E_h is affected by these factors,

174   we ran the liver perfusion study under a constant flow rate and protein-free conditions.

175   The E_h of 4NP at 37°C was approximately-same as the reported value obtained

176   under the steady-state condition [22] and it was decreased according to temperature. E_h

177   is influenced by the liver blood flow rate, protein binding ratio, and hepatic intrinsic

178   clearance ($CL_{int,h}$).   The reduction in E_h could have been caused by alterations in $CL_{int,h}$,

179   owing to the constant flow rate and protein-free conditions. $CL_{int,h}$ is divided into

180   several processes including influx into the cell or efflux from the cell and metabolism by

181   the enzymes in the cell.   The uptake process of 4NP into the liver has not been fully

182   identified.   Quebbeman has reported that the organic anion transporter (Oat) is related

183   to 4NP uptake into the liver [23].

184   Concerning Oat activity under hypothermic conditions, a decrease in the uptake

185   of phenolsulfonphthalein into the liver via Oat was suggested in our previous study [11].

186   Furthermore, we showed a reduction in CYP2E1 activity at low temperatures in this study,

187   and this alteration may have also had an effect on the E_h of 4NP.   Thus, the reduction in

188   CYP2E1 and Oat activity could cause changes in the E_h of 4NP under hypothermic

189   conditions.

190   As the next step, we evaluated the pharmacokinetics of 4NP in rats to identify

191   how alterations in the hepatic disposition affected the pharmacokinetics of 4NP in rats.

192   As illustrated in Fig.2, the plasma concentration of 4NP was significantly increased at

193   28°C and the CL_{tot} of 4NP was decreased according to a reduction in the body temperature

194   (Table I).   In general, the hepatic clearance of a drug depends on hepatic blood flow and

195   E_h.   It has been reported that the blood flow was reduced under hypothermia[24].

196   Moreover, we determined the protein binding ratio of 4NP with BSA by equilibrium

197   dialysis method and it was slightly increased at 28°C compared to 37°C (data not shown).

198   These results suggest that the reduction of CL_{tot} might be caused by reduction of E_h,

199   hepatic blood flow or unbound fraction of 4NP.

200   Conclusion

201   We showed that the elimination velocity from homogenate and E_h of 4NP were

202   decreased under hypothermic conditions and that these alterations could affect the

203   pharmacokinetics of a drug under hypothermic conditions in rats.   These results may be

204   helpful in predicting the pharmacokinetics of a drug during hypothermia.

205

206   Acknowledgement

208

209 Table

210 Table I Pharmacokinetic parameters for the plasma concentration time profiles of 4NP

211 after its i.v. administration to rats at a dose of 2 mg under different temperatures.

Body temperature (^{o}C)	37	34	32	28
AUC$_p$ (μg·mL/min)	106.4	207.5	372.7	727.0**
	±16.9	±111.1	±42.2	±140.5
MRT$_p$ (min)	6.5	15.5	34.0**	48.4**
	±1.1	±5.3	±4.7	±5.2
CL$_{tot}$ (mL/min)	20.7	17.9	5.7**	3.2**
	±3.1	±5.7	±0.7	±0.6

212 AUC$_p$: area under the plasma concentration-time profile, MRT$_p$: mean resistance time,

213 CL$_{tot}$: total body clearance.

214 The AUC$_p$, MRT$_p$, and CL$_{tot}$ represent the mean ± S.E. of at least four experiments.

215 ** $p < 0.01$: significantly different from the result at 37°C

216

217   Legend to Figures

218   Fig.1   (A) Hanes-Woolf plots of 4NP elimination from rat liver homogenate under

219   different temperatures and (B) Km and Vmax of 4NP at each temperature obtained by

220   Hanes-Woolf plots.   Each point represents the mean ± S.E. and bar represents the mean

221   + S.E. of at least three experiments.   Key: at 37°C (○), 34°C (■), 32°C (▲), or 28°C

222   (◇).   **: $p < 0.01$, significantly different from 37°C.

223

224
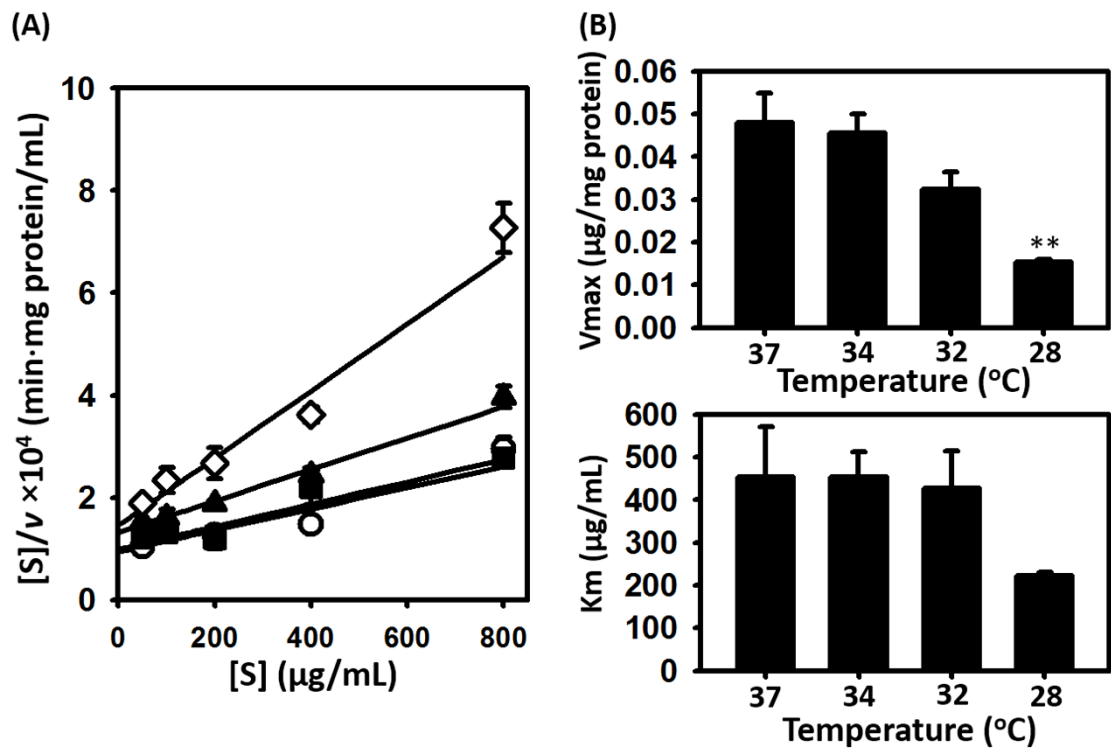
225   Fig.2   Relationship between the perfusate temperature and hepatic extraction ratio (E_h)

226   of 4NP at a dose of 0.2 mg in the rat liver perfusion system.   Each symbol represents the

227   mean $\pm$ S.E. of at least five experiments.   The solid line represents the regression curve.

228   Key: at 37°C ($\circ$), 34°C ($\blacksquare$), 32°C ($\blacktriangle$), or 28°C ($\diamondsuit$).
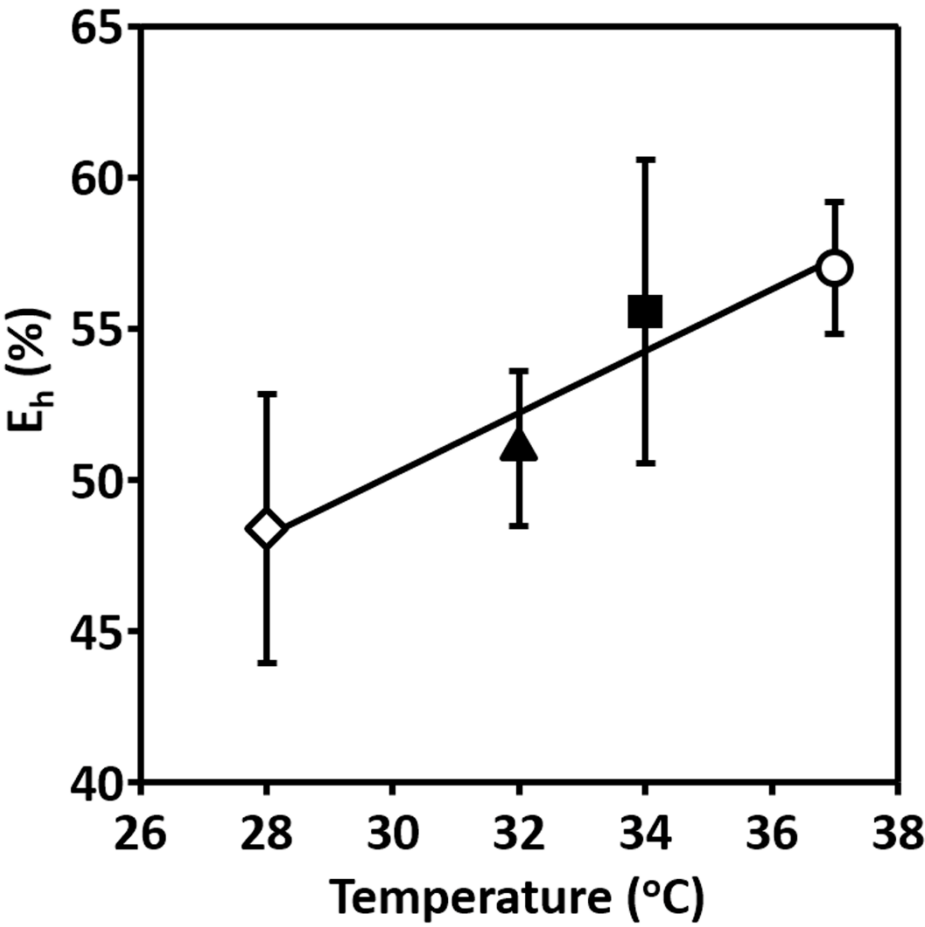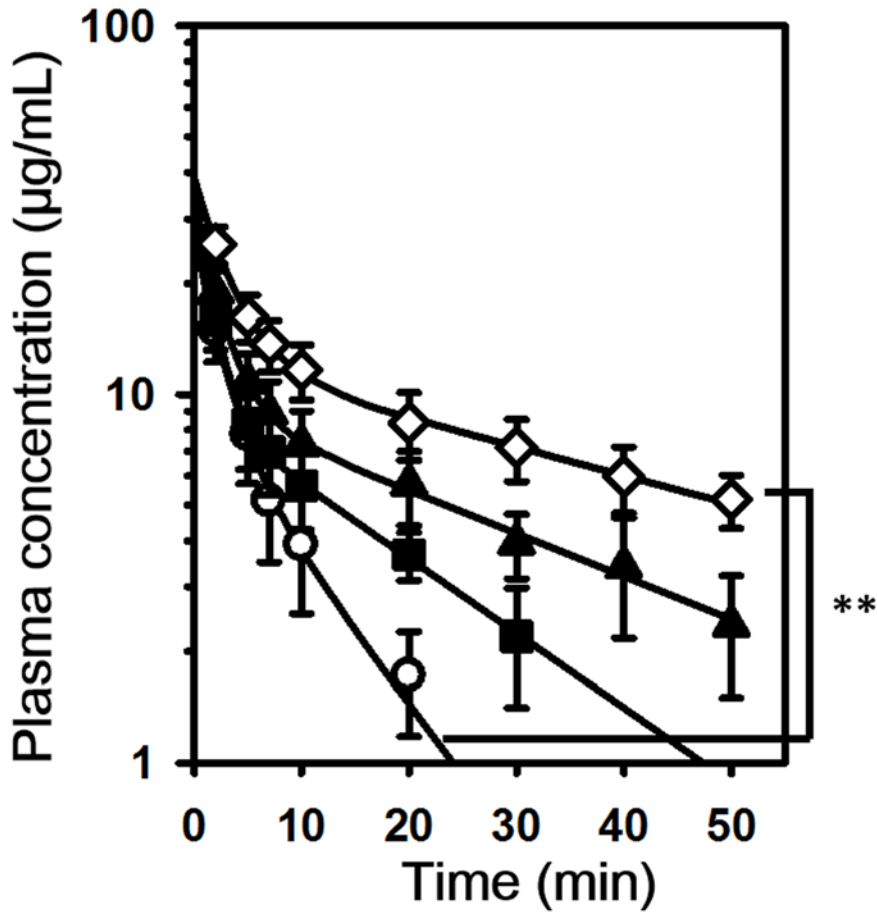

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230

231   Fig.3   Plasma concentration profiles of 4NP at a dose of 2 mg after its i.v. administration

232   to rats at 37°C (○), 34°C (■), 32°C (▲), or 28°C (◇).   Each point represents the mean ±

233   S.E. of at least five experiments.

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